

#### **Review Article**

# Variation, plasticity and possible epigenetic influences in species belonging to the tribe Gomphocerini (Orthoptera; Gomphocerinae): A review

#### Seyed Hossein Hodjat and Alireza Saboori\*

Jalal Afshar Zoological Museum, Department of Plant Protection, College of Agriculture and Natural Resources, University of Tehran, Karaj, Iran.

Abstract: Environmental conditions can cause variation in morphology, behavior, and possibly epigenetic in the numerous species of the Gomphocerinae, especially in mountain habitats. Plasticity and changes in morphology in many of the species in this subfamily is caused by character segregation through the female choice of copulation that has produced various clines, sub-species or species groups. The variation and plasticity, as a result of environmental stress, besides morphology, affect physiology and epigenetics of many insect species. Environmental stress and female assortative mating might be accompanied by hybridization in populations, resulting in character divergence and speciation after a long period of time. Contemporary evolution and/or epigenetic inheritance may be a reason for their variation in acoustic and morphology of Gomphocerinae and the main factor in the present situation of difficulty in their classification. We review possible effects of environmental stress on plasticity, hybridization, and speciation by the appearance of endemic species. About half of the insect pest species have reduced their impacts as pests under global warming. The present insect pest situation in Iran is discussed.

**Keywords:** Classification, Hybridization, Groups, Gomphocerini, Phenotype, Song

#### 1. Introduction

Geographical distribution of Gomphocerinae species is the source for variation, plasticity, and hybridization by environmental effects. Many endemic species and subspecies are created by acoustic divergence and sympatric speciation. The degree and direction of a trait is related to sexual dimorphism and female choice for copulation in insects (Robinson and Hall,

Handling Editor: Ali Asghar Talebi

\*Corresponding author: saboori@ut.ac.ir

Received: 16 September 2019, Accepted: 14 August 2020

Published online: 03 October 2020

2002; Mol *et al.*, 2003; Tishechkin and Bukhvalova, 2009; Vedenina and Helversen, 2009; Şirin *et al.*, 2010, 2014; Stillwell *et al.*, 2010; Vedenina and Mugue, 2011). Therefore morphology alone is not enough to define many species of Gomphocerini and taxonomists are using sound characteristics for identification. Biodiversity studies of Gomphocerinae species are difficult due to a lack of knowledge for accurately identifying them in Iran without recording their songs and considering other factors causing variation in their populations (Sultana *et al.*, 2013; Hodjat *et al.*, 2019).

More than 81 species of Gomphocerini grasshoppers are widely distributed in the world (Sergeev, 2011). In other words, Gomphocerini

is an assemblage of several similar grasshopper genera including Gomphocerus, Myrmeleotettix, Chorthippus, Pseudochorthippus, Stauroderus, and Aeropedellus (Jago, 1971; Defaut, 2011, 2012, 2017). The genus Chorthippus has 229 species and is divided into 2 or 3 subgenera Chorthippus (Altichorthippus) (20 species), Chorthippus (Chorthippus) (32 species), and for various authors (including orthoptera species file OSF), Chorthippus (Glyptobothrus) (57 species). The other species of Chorthippus are not classified into any subgenera. The Chorthippus (Glyptobothrus) biguttulus group has 12 species and C. binotatus group has two species. The genus Pseudochorthippus Defaut, 2012 with four species and two sub-species is closely related to Stenobothrus Fischer and Omocestus Fieber (subtribe Stenobothrina).

Character changes in Gomphocerini species are a reaction to environmental factors that its stress may have influenced their genetics, epigenetics and external morphology (Laiolo et al. 2013). Neurohormones regulate all life processes in Gomphocerinae and insects in general (Perica-mataruaga et al., 2006; Hodiat, 2006). Juvenile hormones (JHs) and the neurosecretory neurons (NSNs) secretions of corpora cardiac (CC), corpora allata (CA) and the brain (CNS) release methylsulfonylmethane (MSM) into the haemolymph. The gene expression is involved in C. biguttulus (L.) and chitin-binding. biogenic amines Transcriptomic methods of profiling show differential expression pattern in various stages of C. biguttulus life cycle and are described by Berdan et al. (2017), and Tatsuta and Butlin Gene expression pattern in C. albonemus change when pastures have been overgrazed in cases of food shortage (Qin et al., 2017). The increased activity in locust is initiated after adipokinetic neurohormones are secreted (Hodjat, 2016). The reaction to communication in response environmental conditions has been studied in C. biguttulus (Klappert and Reinhold, 2003). Species in the Chorthippus (Glyptopothrus) group communicate by special male songs to fend off competitors from the female, a behavior regulated by *corpora cardiac* secretion (Heinrich *et al.*, 2012).

**Populations** in various species of Gomphocerini may change their characteristics different choosing selection Differentially fixation of various characters on the population of a species may increase their adaptation and survival chance. The process of divergence in colour, morphology, and song in parallelus **Pseudochorthippus** (Zetterstedt) different produces traits genetic by polymorphism (Kohler et al., 2017). A long separation of their population increases their genetic distance, colour and morphological or acoustic diversification (Tregenza et al., 2000; Butlin and Hewitt, 2008; Vedenina and Mugue, 2011; Monge, 2017). Subspecies and species group formations are the results of genetic or epigenetic changes in Gomphocerini (Smith and Ritchie, 2013; Noguerales et al., 2016), through assortative mating and environmental conditions (Klapper and Reinhold, 2003; Burggren, 2017).

Rapid character divergence of grasshoppers in response to the male songs for copulation and other environmental factors are influencing taxonomists to clarify their classification. A new line of research on sound production, epigenetic, and stress has proved its influence Gomphocerinae species radiation (Franzke and Reinhold, 2012). We refer to some publications of research on species radiations creating endemic forms phylogenetic, taxonomic, stress and genetic or epigenetic investigations (Pravdin, 1969; Mark, 1983; Hodjat et al., 2018; Burggren, 2018; Riede, 2018).

#### 2. Phylogeny and classification difficulties

Orthoptera Species File (OSF) has provided a list of 20 tribes classified in the Gomphocerinae subfamily. Gomphocerini Fieber, 1853 or Chorthippini Shumakov, 1963 with 16 genera is among the largest group of grasshoppers (Cigliano *et al.*, 2018). Some investigations have placed Gomphocerini, Chrysochraontini, Dociostaurini, and Arcypterini in the phylogenetic structures of Gomphocerinae and included

Helversen

Omocestus, Myrmeleotettix, Stenobothrus, Chorthippus and Arcyptera in their phylogenetic tree (Vedenina and Mugue, 2011; Garcia-Navas et al., 2017; Rohde et al., 2017).

Considerable phenotypic plasticity is a major source of difficulty in developing a unique phylogenetic classification for Gomphocerini (Song, 2005; Zhang et al., 2013; Şirin et al., 2014; Defaut, 2017). Morphological characters of a few species in Chrysochraontini, Arcypterini, Acridinae. Melanoplinae, Locustinae, Calliptaminae and Cyrtacanthacridinae tribes or subfamilies are close to Gomphocerinae or overlapping in some morphological characters (Defaut, 2017). The most common phylogenetic method of their separation is by the NADH (nicotine amid adenindinuclrotide phosphate) mitochondrial analysis. Yet, with this method, the relative time of appearance of Ramburiellini, Dociostaurini, and Arcypterini could not be determined. However, it was estimated that the Gomphocerini and species in Stenobothrus, Myrmeleotettix, Omocestus, Stenobothrus, and Stauroderus genera are closely related (Defaut, 2017). The acoustic and phylogenetic relations of various polyphyletic taxa in Gomphocerinae are comparable (Nattier et al., 2011; Vedenina and Mugue, 2011).

Integrating genomic and phylogenetic data with molecular studies might result in species delimitation and find how various traits produced the evolutionary radiation grasshoppers (Contreras and Chapco, 2006; Nattier et al., 2011; Via, 2012; Noguerales et al., 2018). The common species Gomphocerini in Europe and the Middle East are C. albomarginatus, C. apricarius, C. maritimus, C. brunneus, C. biguttulus and Pseudochorthippus parallelus (Harris, 1835), the latter is classified as one of the P. parallelus subgroups (Sergeev, 2011).

Morphologically similar of groups Gomphocerini species are as follows:

#### 2. 1. Chorthippus albomarginatus species group Sound changes in this group of species is studied by Vedenina (2015); Vedenina and

2009);

(2003,

Vedenina

Shestakov (2014); Vedenina et al. (2001, 2007b). Hybridization of C. albomarginatus and C. oschei produces asymmetric populations and reduces the hybrid viability (Vedenina et 2007a). The founder effect of the population with different songs can establish sibling species and new cline (Helversen and Helversen, 1997). The main driving force of speciation in this group of species is sexual selection by the female choice of courtship.

#### 2. 2. Chorthippus (Glyptobothrus) biguttulus species group and subgroup

Habitat segregation, character diversification by the female choice of copulation, and hybridization are known to have established many similar species in this group or subgroup of species (Saldamando et al., 2005; Gottsberger 2007; Şirin et al., 2010; Gomez and Dyck, 2012). The complex C. biguttulus group is formed by a mixture of macropterous and brachypterous forms in occidental and central Europe (Ingrisch, 1995). Many species in this group share habitat with C. binotatus binotatus (Charpentier) and are similar to C. jacobsi (Harz), C. bornhalmi (Harz), C. ilkazi (Uvarov), C. miramae (Ramme) and C. rubratibialis (Schmidt). Hybridization is reported between these species in this group (Stumpner and Helversen, 1994; Klapper and Reinhold, 2003; Helversen et al., 2004; Saldamando et al., 2005; Gottsberger, 2007; Franzke and Reinhold, 2012b).

#### 2. 3. Pseudochorthippus parallelus species group

Pseudochorthippus montanus (Charpentier), P. curtipennis (Harris), P. tatrae (Harz), P. parallelus erythropus (Fieber, 1858), P. p. serbicus (Karanian, 1953), and P. p. tenuis (Brulle, 1832) are classified in this group. Hybrids between the last two subspecies are formed because of cytoplasmic incompatibility as a result of Wolbachiella infection (Bella et al., 2010). The differences in song and the hypothetical change of flight activity in separating flightless and winged forms have been studied in this group (Ritchie, 1990; Tregenza et al. 2000; Butlin and Hewitt, 2008; Kohler et al., 2017) Morphological

and

variations in *P. parallelus* from samples at 450-2500 m elevation indicate that tegmina and body length of samples was decreased by ascending the mountain but morphology of pronotum is unchanged.

#### 2. 4. Chorthippus (C.) dorsatus species group

Chorthippus loratus (Fischer von Waldheim) and C. dorsatus are sibling species in this group of species. Songs in C. albomarginatus group represent an advanced development of the C. dorsatus group (Stumpner and Helversen 1994). The complexity of the calling songs in these two groups of grasshoppers is comparable (V. Vedenina personal communication).

## 2. 5. Chorthippus (Glyptobothrus) binotatus species group

Three subspecies of *C. binotatus* and three subspecies of *Chorthippus* (*G.*) saulcyi (Krsuss) are classified in separate groups of species. The two groups are very similar and are usually classified into one group. The genetic lineages between populations in this group of species living in the mountainous region of the border between France and Spain were studied by Defaut (2011) and Noguerales *et al.* (2016, 2018). Isolation of populations living in different mountain habitats may have been the cause for species separation in this group (Noguerales *et al.*, 2016).

## 3. Factors affecting classification of the Gomphocerinae

#### 3. 1. Variation and Plasticity

The definitions of phenotypic plasticity and its relation to selection, coevolution, ecological speciation, genetic drift, phylogenetic inference, pleiotropy, reinforcement, sexual dimorphism and size constraints have been explicitly described with examples of Gomphocerini species (Whitman and Agrawal, 2009; Feng et al., 2015; Zinna et al., 2018). Plasticity is more liable to fix characters after insects acquire a trait. Variations are temporary morphological character dissociation that may fix on a part of a population for several generations. Plasticity and variation may cause speciation or contemporary evolution i.e. the evolution of traits over a short period of time (e.g. centuries rather than eons; Górür, 2005; Whitman and Agrawal, 2009). Plasticity can change individual, population or species epigenetic by environmental stress in insects (Hoffmann and Parsons, 1991; Oxford Research Group, 2015; Augustyniak *et al.*, 2016; Burggren, 2018).

Phenotypic plasticity reflects the "reaction norm" of individuals to climate and other environmental conditions (Laiolo et al., 2013). Reception of environmental stressful conditions by CNS can cause variation and plasticity in parts of species population (Ronacher, 1989; Romer, 1993; Skinner, 2014). The genotype of an organism in the reaction norm may also be affected by life incidents. That is, plasticity confers the capability of producing changes in phenotype in response to environmental changes (Sgro et al., 2016). Final decision for direction of movement towards a stimuli such as attraction to the opposite sex in C. biguttulus, Gomphocerus sibiricus (L.), Omocestus viridulus (L.) and Stenobothrus lineatus (Panzer, 1796) is taken by the brain signals that are involved to activate the appropriate muscles (Helversen and Helversen, 1997; Klapper and Reinhold, 2003). Therefore plasticity is a broad term that can apply to the genetic, epigenetic and contemporary evolution of Gomphocerini response to the environment.

#### 3. 1. 1. Plasticity and variation on insect pests

Increasing plant defense against herbivores and pathogens is related to DNA methylation and histone modification by the transgenerational defense. Changes in the phenotypic traits of grasshoppers are also linked to performance and gene profile. Genes in C. albonemus change expression pattern when pastures are overgrazed and in cases of food shortage (Qin et al., 2017). The effects of ambient and optimal temperatures on 31 species of insect pests have been compared by Lehmann et al., (2018). North Europe, South America, and Australia had fewer outbreaks of these pests compared to Asia and Africa. Aridity is an important factor in locust and

grasshoppers to reach the outbreak point (Uvarov, 1957). Repeated insecticide treatments without considering the importance of natural enemies can cause pest outbreaks (Hodjat, 1967). A pest control project using modeling to consider the efficiency of pest control methods for Iran is suggested by Hodjat (1974). Modares-Awal's (2012) list of Orthoptera pests in Iran indicated that most species do not seriously damage crops and hardly reach to outbreak position. Caution for not using repeated insecticide treatments without pest management strategies has reduced pest damage to crops (Hodjat, 1967).

At present, the locust outbreaks are occasionally reported only after a rainy winter and revival of natural pastures in Iran. Some pest species such as Eurygaster integriceps Puton, Bemisia tabaci suppressalis (Gennadius, 1889), Chilo(Walker) and occasionally other insect pests of crops and fruit trees need a continuous survey (Davatchi, 1954). However, locust phase polymorphism, phenotypic plasticity, including various mechanisms responsible for the insurgence of aggressive biotypes, or resistance to insecticides are the main factors behind the pest control problems (Song, 2011; Augustyniak, 2016; Hodjat, 2016; Sgro et al., 2016; Lehmann et al., 2018).

#### 3. 2. Epigenetic and stress influences

Various stressors trigger epigenetic modifications in animals and plants. Modifications allow gene transcription and remain in genome memory. Transgenerational epigenetic memory ensures plasticity and genetic variation (Tricher, 2015). Gomphocerini species may change colour, size, and activity in response to climatic conditions, and these changes are epigenetically inheritable (Rohde et al. 2017). In fact, such polymorphisms are common in Orthoptera (Wall, 1987; Mol et al., 2003; Laws and Belovsky, 2010; Stillwell et al., 2010; Gomez et al., 2012; Nettle and Bateson, 2015; Valverde and Schielzeth, 2015; Lonsdale, 2018). Adaptive mutations by stress in locust are non-directional but they may cause new traits in populations (Whitman and Agrawal, 2009). Such rapid phenotypic adaptation to new environmental conditions reduces the strength of divergent selection (Sword, 2002, 2003; Smadja and Butlin, 2011).

Methylation of DNA through epigenetic regulation of the neural transcriptome occurs in response to the stress of insects that become resistant to various toxins and escape unsuitable environmental conditions such as drought, heat and food restriction. In C. biguttulus DNA breaking and its repair with new arrangements of amino acids is the result of environmental stress effects (Bijlsma and Loeschke, 2005: Augustyniak et al., 2006; Burggren, 2017; Lonsdale, 2018). Environmental stress and epigenetic inheritance are closely related to the physiological change of organisms in reaction to the inhabiting conditions that affect individual in the population (Skinner, 2014). of Methylation DNA through epigenetic regulation of the neural transcriptome occurs in response to environmental stress in insects. Most insects become resistant to various environmental toxins and escape unsuitable living conditions such as drought, heat and food restriction. Food shortages in crowded conditions of locust may increase cyclooxygenase, or COX stress enzymes. The enzyme affects the metabolism of locust for a few generations and the gregarious phase of the locust is produced (Rogers et al., 2003; Hodiat, 2006, 2016; Song, 2011). DNA breaking and its repair with new arrangements of amino acids in C. biguttulus are the results of environmental stress effects (Bijlsma and Loeschke, 2005; Augustyniak et al., 2006; Burggren, 2017; Lonsdale, 2018).

The genes responsible for oxidative stress are expressed by superoxide dismutase (SOD) (Bijlsma and Loeschcke, 2005; Glastad *et al.*, 2011; O'Grady and Markow, 2012; Skinner, 2014; Burggren, 2017, 2018). Transgenerational morphological and genetic characters are modified by stress-related levels of cytochrome oxidase. The toxin stress effects on mitochondria and metabolism as a result of aging and zinc toxicity in insects are studied in detail (Augustyniak *et al.*, 2008, 2011, 2016;

Skinner, 2014; Bultman, 2015; Lonsdale, 2018). Food shortages in crowded conditions of locust may also increase COX stress enzymes. After a few generations of crowding, metabolic enzymes change appears in locust and may change them to the gregarious phase (Rogers *et al.*, 2003; Hodjat, 2006; Song, 2011). To maintain plasticity, stress-induced epigenetic modifications buffer against DNA sequence-based evolution (Tricker, 2015).

epigenetic regulatory system neurosecretions allows reaction against stress effects (Smith and Ritchie, 2013; Tetreau, 2018). Stressors with different intensity influence the insect nervous secretions and hormones such as biogenic amines controlling carbohydrate or lipid metabolism and causing variation in insect behavior and morphology (Perić-Mataruga et al., Insect pricking by injection wound 2006). antimicrobial produced phenoloxidase masked the effect of lipopolysaccharide (LPS) immune challenge (Monge, 2017).The embryogenesis in C. biguttulus by transcriptomic profiling at various stages of development shows that genes are upregulated in the imago of C. biguttulus and are mostly involved in aging and immunity (Berdan et al., 2017). In stressed insects, changes in NF-kB (nuclear factor kapperlight chain of activated B-cells) of proteins that bind to DNA can decrease the antioxidant activities in the Gluthathion redox system. Molecular and physiological effects of stress can impose novel trade-offs and character changes (Isaksson et al., 2011). Stress can produce asymmetric forms in insect populations (Parsons, 1991, 1992; Møller and Swaddle, 1997; Jentzsch et al., 2003; Hodjat, 2016) or asymmetric mate choice in grasshoppers (Hochkirch and Lemke, 2011). Multigenerational environmental stressors on insects may last for a few generations by epigenetic effects but after the disappearance of stress, populations can obtain the previous phenotypic form (Burggren, 2016).

### 3. 3. Phenotypic plasticity in a noisy and toxic environment

Environment and stressful conditions can change the phenotype of grasshoppers by

changing their song loudness, resisting the toxins in their surroundings or competing with other males in copulation with females. Males of C. biguttulus resist the road noise by increasing the loudness of their song. Female selection of these males for copulation will increase the chance of progeny survival (Einhaupl et al., 2011; Lampe et al., 2012, 2014). A male of Omocestus viridulus sings shorter songs when other males are close by (Eiriksson, 1992). Various morphological forms of Glyptobothrus pullus (Philippi) appear as a result of feeding on different host plants in Switzerland (Steiner, 2006). In C. biguttulus DNA breaking and its repair with new arrangements of amino acids are the results of environmental stress effects (Bijlsma and Loeschke, 2005; Augustyniak et al., 2006; Burggren, 2017; Lonsdale, 2018).

In unfavorable environmental conditions, DNA damage or its partial modification may cause phenotypic plasticity (Steiner, 2006; Gottsberger, 2007; Dowle et al., 2014; Rohde et al., 2015a). The genetic modification in the beet armyworm, Spodoptera exigua (Hübner) is taking place after about 120 generations in multigenerational microevolution. DNA damage by zinc treatment disrupted the development of C. brunneus, during induced embryonic disorders diapause or (Augustyniak et al., 2011). In stressed insects, Glutathione S-transferase (GSTs) and catalase (CAT) enzymes, important for metal defense against intoxication, were reduced. The cycle of protein digestion inside the cell and its effects on synthesizing antioxidant is reviewed by Isaksson et al., (2011). Common field grasshoppers usually can adapt to metal-polluted habitats (Augustyniak et al., 2008, 2011). Zinc-induced effects are transferred to the embryonic DNA of C. brunneus (Augustyniak et al., 2006, 2016). Chernobyl radiation caused genetic damage and mutation on C. albomarginatus, as well as oxidative stress (Bonisoli-Alquati et al., 2018).

#### 3. 4. Epigenetic modifications

So far the epigenetic influence on morphological changes or character divergence of *Chorthippus* species is not known.

Transgenerational defense induction epigenetic inheritance are cued by methylation signaling in the parental generation. The changes in the offspring genotype caused by gene imprinting and neural fingerprints transcribe experiences of insects by affecting maternal and paternal DNA methylation (Norouzitallab et al., 2019; Ernst et al., 2015; Burggren, 2017). Epigenetic modification is known to cause phenotypic changes by DNA methylation across insects (Bewick et al., 2017). DNA methylation can regulate genome at CG -sets on chromosomes. They can also regulate transcription at the CpG islands (Deaton and Bird, 2011). The epigenetic mechanisms can cause polymorphism, change the locust phase, and act on caste formation of social insects (Weiner, 2012; Ernst, et al., 2015; Mallon et al., 2016; Burggren, 2017; Lonsdale, 2018). Environmental epigenetic modification is closely related to the stress effects by stimuliresponse effects for defense on living organisms.

#### 4. Hybridization and endemism

Basic factors responsible for hybridization are influenced genomically by genetics epigenetics. They influence cell division by cytoplasmic and genetic incompatibility resulting in the presence of different genomes (Ishikawa and Kinoshita, 2009). Hybrid and endemic species of Gomphocerinae described in relation to asymmetry in female preferences of male songs. Geographical distribution of Gomphocerinae species is the source for variation, plasticity, and endemism by environmental stress. Many endemic species and subspecies are created by acoustic divergence and sympatric speciation mountain regions. Hybridization and molecular evolution by mDNA sequencing are creating new routes for investigating trait divergence in Gomphocerinae. Trans-generational induction and epigenetic inheritance are cued by environmental signaling in the parental generation. The changes in the offspring genotype caused by gene imprinting and neural fingerprints transcribe experiences of insects by

affecting maternal and paternal DNA methylation (Ernst *et al.*, 2015; Burggren, 2017). Some of the Gomphocerinae hybrid species are produced after females choose to copulate with preferable males songs (Gottersberg, 2007).

#### 4. 1. Hybridization

Hybridization and factors responsible for character diversification may ultimately result in evolution and sympatric speciation. Many publications investigate factors responsible for the separation of population characters in Gomphocerinae and their hybrids (Pravdin, 1969; Gottsberger, 2007; Vedenina et al., 2007a; Smadja and Butlin, 2011; Frank and Reinhold, 2012b; Rhode, 2015; Job et al., 2016). Positive selection in favor of increasing flight ability is involved in some grasshopper populations at high altitudes in the mountains of China (Li et al., 2018). The relative abundance of C. montanus and C. parallelus as specialist and generalist also produce hybrids that can trigger speciation (Rhode et al., 2015). Crosses of the following species can produce hybrids in Gomphocerinae species (Gottesberg, 2007; Bella et al., 2010; Rhode, 2015; Vedenina, 2015):

- C. brunneus X C. jacobsi; C. bigutulus X C. brunneus; C.eisentrauti X C. brunneus;
- C. biguttulus X C. mollis; C. albomarginatus X C. oschei; C. albomarginatus X;
- C. karelini; P. montanus X P. parallelus; Stenobothrus clavatus X S. rubicundus;
- P. parallelus parallelus X P. parallelus ervthropus.

In northeastern Ukraine, *C. albomarginatus* and *C. oschei* produce hybrid offspring. The two crossing species are morphologically very similar except for their songs and organs for song production. The differences in the song are caused by leg movements and the type of pegs involved on femur after hoppers change the angle of their body (Vedenina *et al.*, 2007b; Sardnick *et al.* 2016). Courting males can produce more complex sounds and are preferentially selected by the female's inheritance. Duplicate loci controlled the homologous elements in the songs of the two

copulating species of C. oschei from C. albomarginatus (Vedenina and Helversen, 2003; Vedenina et al., 2007b). C. karelini produced a song with two short elements alternating with a longer element. Even courtship songs of two males of C. karelini recorded from two different localities in Ukraine produced different songs. The songs from these localities had hybrid characteristics of the two populations (Vedenina, 2015). In the Picos district in the north Spain hybridization between C. brunneus and C. jacobsi produced offspring with different numbers of stridulatory pegs in different localities. Various peg numbers produced different cline in the Picos region. The changes were significant from populations approximately 30 km south of Picos de Alps. The differences for wingless meadow grasshoppers, P. parallelus were measured by the nested model for geographic variation. Mark and recapture methods for measuring the elation of stridulatory pegs differences in various habitats have been applied to measure the extent of differences (Bridle et al., 2001). Investigation of hybridization and molecular evolution by mDNA sequencing is creating new routes for finding genetic or phylogeographic trait divergence in Gomphocerinae (Hewitt, 2001). Asymmetric mate choice between the two closely related populations of Gomphocerini in the same locality is presumed to produce hybrid forms (Hochkirch and Limke, 2011; Hochkirch, 2013). Asymmetry is recorded in female preferences between two closely related Stenobothrus clavatus and S. rubicundus (Vedenina et al., 2014; Sardnick et al., 2016). Song of C. biguttulus characteristics in relation to male robustness and the attractiveness to females. The loudness of to neutralize also changes surroundings (Gottsberger, 2007; Einhaupl et al., 2011; Lampe et al., 2014).

Hybrid sterility and mate preferences between two closely related species of Gomphocerini are related to many factors such as genetic incompatibility, gene flow between sympatric species, attractiveness to intermediate phenotypes and plasticity (Gootsberger, 2007; Hochkirch, 2013; Sgro *et al.*, 2016). Courting males can produce more complex sounds and are

preferentially selected by the females. Inheritance implying duplicate loci that control the homologous elements in the song of two closely related species are involved in hybridization between them such as hybrids between *C. oschei and C. albomarginatus* (Vedenina and Helversen, 2003; Vedenina *et al.*, 2007b).

Even courtship songs of two males of C. karelini recorded from two different localities in Ukraine produced different songs. The songs from these localities had a hybrid characteristic of the two populations (Vedenina, 2015). Asymmetry is observed in female preferences between two closely related Stenobothrus clavatus and S. rubicundus (Vedenina et al., 2014; Sardnick et al., 2016). In the Picos district of north Spain, hybridization between C. brunneus and C. jacobsi has produced offspring with a different numbers stridulatory pegs in different localities. Various peg numbers produced different clines in the Picos region. The changes were significant from populations approximately 30 km south of Picos de Europa Mountains.

Mark and recapture method is used for measuring the relation of stridulatory peg numbers of C. brunneus living in different habitats in Santander mountain regions of north Spain. The principal component analysis for mean peg scores against the distance from the fitted cline center shows that C. brunneus can be easily distinguished from G. jacobsi by their peg numbers. The hybrid forms are found within 30 km distance (Bridle et al., 2001). Similar studies of female choice of male song for copulation have been conducted in other grasshopper species (Ritchie, 1989, 1990; Helversen and Helversen, 1997; Bridle et al., 2001; Benediktov, 2005, 2014; Einhaupl et al., 2011; Heinrich et al., 2012; Job et al., 2016; Kohler et al., 2017).

#### 4. 2. Endemism

Endemism is produced by divergence in morphology and the song of *Pseudochorthippus parallelus* (Zetterstedt) populations in mountains. Their populations by genetic polymorphism have chosen various traits. A long separation of population traits increased

their genetic distance and caused morphological or acoustic diversification in various directions. The variability in characters are signs for forming an endemic or sympatric species (Tregenza *et al.*, 2000; Butlin and Hewitt, 2008; Stillwell *et al.*, 2010; Vedenina and Mugue, 2011; Monge, 2017).

Chorthippus pullus by feeding on various host plants produced plastic forms in Switzerland (Steiner, 2006). Procedures for the appearance of endemic species in Chorthippus in Turkey (Mol et al., 2003) and the list of Acrididae endemic species of Iran (Hodjat et al., 2018) show the reality of having endemic forms of grasshoppers. The Euloryma Spearman, 2013 (Hemiacridinae; Acridinae) of the Cape Floristi region in South Africa produce endemic species in their narrow distribution area. Euloryma larsinorom and E. are congeneric flightless distributed in natural shrub and heath vegetation. E. umoja and E. ottei can tolerate vineyard and agricultural environment (Adu-Acheampong et al., 2017). DNA methylation leading to epigenetic inheritance of the modified phenotype in endemic grasshoppers requires of further investigation (Nettle and Bateson, 2015).

#### 5. Song and phenotypic variation

Sound in Gomphocerinae is produced by rubbing the hind femora with a row of the stridulatory pegs, up and down against the radial vein of the tegmina, which is more prominent than the other veins and is called the *plectrum*. The song of males in C. dorsatus, C. dichrousand C. loratus is also produced by movements of the hind femur against the tegmina but differs in various localities. The song character may change by the position of their upward body direction or the speed of rubbing the set of pegs. Detailed registration of song characters differs between Chorthippus (Glyptobothrus) bozdaghi (Uvarov), Chorthippus (Glyptobothrus) ilkazi (Uvarov) and Chorthippus (Glyptobothrus) helverseni (Mol et al., 2003). Temporal features of the song in C. brunneus and C. jacobsi depend on the neurological pattern that controls leg movement. The neuromuscular activity during stridulation of C. biguttulus, Gomphocerus sibiricus, Omocestus viridulus (L.) and Stenobothrus lineatus (Panzer, 1796) is not related to the number of pegs (Elsner, 1974; Saldamando et al., 2005; Perić-Mataruga et al., 2006).

Mating of female grasshoppers with a preferable male is because of their particular song characteristics and can initiate sympatric species for adapting to the prevailing conditions. Consequently, in large parts of central Europe, hybrid populations and sympatric species have originated (Hochkirch and Limke, 2011; Hochkirch, 2013; Rohde *et al.*, 2015b, 2017).

Chortoicetes terminifera song is changed by crowding conditions (Song, 2005, 2011; Chapuis et al., 2010; Ernst et al., 2015). The internal reaction to resist the road noise in males of *C. biguttulus* is causing an increase in the loudness of their song. Female selection of these males for copulation will also increase the chance of progeny survival (Klappert and Reinhold, 2003; Einhaupl et al., 2011; Heinrich et al., 2012; Lampe et al., 2012).

Limited condition dependence of male body condition is likely to be correlated with male acoustic signals in C. biguttulus (Franzke and Reinhold, 2012a). The embryogenesis in G. bigutullus by transcriptomic profiling at various stages of development shows that gene expression at various stages of the life cycle ranged from synthetic biogenic amines to chitin-binding. Genes upregulated in the imago of C. biguttulus were mostly involved in aging and immunity (Berdan et al., 2017). In stressed insects, changes in NF-kB (nuclear factor kapper-light chain of activated B-cells) of proteins that bind to DNA can decrease the antioxidant activities in the Gluthathion redox system. Molecular and physiological effects of stress can impose novel trade-offs and character changes (Isaksson et al., 2011). Stress can produce asymmetric forms in insect populations (Parsons, 1991, 1992; Møller and Swaddle, 1997; Jentzsch et al., 2003; Hodjat, 2016) or asymmetric mate choice in grasshoppers (Hochkirch and Lemke, 2011). Multigenerational environmental stress on insects may last for a few generations by epigenetic effects but in some cases after the disappearance of stress populations can show phenotypic modification (Burggren, 2018). Pharmacologically induced stridulation had little effect on the courtship song and only changed one or two of the sound elements (Vedenina *et al.*, 2001).

#### 6. Conclusion

Environmental and genetic factors have produced various endemic populations with new characters that may create endemic hybrid and sympatric species of grasshoppers (Hodjat et al., 2018). The problem for taxonomists is to classify Orthoptera by considering their phylogenetic or ancestral relationship. In taxonomy, mainly morphological abrupt changes between taxa are the measure for defining them but variation in morphological characters is the gradual results of gene flow into the population. Morphological differentiation despite gene flow is based on a selection of phenotypes in the survived population as a result of the environmental stress (Dowle et al., 2014). It is difficult to find the degree of morphological in each character changes taxon. physiological trade-offs and diversifying characters of populations are known to affect the taxonomy and life history at various localities in animals (Zera and Harshman, 2001). In stressed insects, changes in NF-kB (nuclear factor kapperlight chain of activated B-cells) proteins that bind to DNA can decrease the antioxidant activities in the Gluthathion redox system. Molecular and physiological effects of stress can impose novel trade-offs and character displacements (Isaksson et al., 2011).

Phase polymorphism and variation in locust morphometrics appear to be the result of environmental stress effects on their physiology (Hodjat, 2016). Epigenetics in contemporary evolution separates the characters of the migratory phase for several generations from the solitary phase (Mallon *et al.*, 2016). This phenomenon can also affect other Gomphocerini species and produce variation in their morphological characters in response to environmental stress (Parsons, 1992; Chapuis *et al.*, 2010; Ketmaier *et al.*, 2010; Murren *et al.*, 2015). Hormones and

physiological changes, affecting insects due to environmental conditions, are capable of being epigenetically inherited through methylation. A new line of research is proposed for investigating the relation of epigenetic changes to stress in insects (Merritt and Bewick, 2017).

#### Acknowledgments

We thank Dr. Bernard Defaut (Quartier Babi, hameaud' Aynat, 09400 Bédeilhac-et-Aynat, France) for taxonomic revision and reprints to improve the text. We acknowledge help of Warren Burggren the Dr. (Developmental Investigative **Biology** Research, University of North Texas, Denton USA) for many suggestions to improve the text. Dr. Varvara Vedenina (Russian Academy Science) for her suggestions, corrections, and sending us the PDFs of her publications.

Dr. Klaus Riede, Zoologisches Forschungs museum Alexander Koenig- Leibniz InstitutfürBiodiversität der Tierpe, Germany, helped us with the manuscript. We thank Dr. Maria Augustyniak (Department of Animal Physiology and Eco-toxicology, University of Silesia, Katowice, Poland).

#### References

Adu-Acheampong, S., Samways, M. J., Landmann, T., Kyerematen, R., Minkah, R., Mukundamago, M. and Moshobane, C. M. 2017. Endemic grasshopper species distribution in an agro-natural landscape of the Cape Floristic Region, South Africa. Ecological Engineering, 105: 133-140.

Augustyniak, M., Babczynska, A. and Augustiniak, M. 2011. Oxidative stress in newly-hatched *Chorthippus brunneus*- the effects of zinc treatment during diapauses, depending on the female's age and its origins. Comparative Biochemistry and Physiology–C. Toxicology and Pharmacology, 154(3): 172-179.

Augustyniak, M., Babczynska, A., Kozlowski, M., Sawczyn, T. and Augustiniak, M. 2008. Effects of zinc and

- female aging on nymphal life history in a grasshopper from polluted sites. Journal of Insect Physiology, 54: 41-50.
- Augustyniak, M., Juchimiuk, J., Przylowicz, W. J., Mesjasz-Przybylowicz, J., Babczynskia, A. and Migula, P. 2006. Zinc induced DNA damage and the distribution of metals in the brain of grasshoppers by the comet assay and micro-PIXE. Comparative Biochemistry and Physiology–C. Toxicology & Pharmacology, 44: 242-251.
- Augustyniak, M., Plachetka-Bozek A., Kafel, A., Babezynska, A., Tarnawska, M., Janiak, A., Loba, A., Dziewiecka, M., Karpe-Kaczmarek, J. and Zawisza-Raszka, A. 2016. Phenotypic plasticity, epigenetic or genetic modifications in relation to the duration of Cd-exposure within a microevolution time range in the beet armyworm. PlosOne, 11(12): e0167371. https://doi.org/10.1371/journal.pone.0167371.
- Bella, J., Martinez-Rodriguez, P., Arroyo-Yebras, F., Abernal, J. S., Fernandez-Calvin, B., Mason F. L. and Zabal-Aguirre, M. 2010. *Wolbachia* Infection in the *Chorthippus parallelus* Hybrid Zone: Evidence for its role as a reproductive barrier. Journal of Orthoptera Research, 19(2): 205-212.
- Benediktov, A. A., 2005. Fauna and acoustic signals of the genus *Chorthippus* Fieber (Orthoptera; Acrididae) from southern Siberia. Proceedings of the Russian Entomological Society, St. Petersburg, 76: 118-130.
- Benediktov, A. A., 2014. The new data on distribution of the pygmy grasshopper *Tetrix tartara* s. l. (Orthoptera: Tetrigoidea: Tetrigidae) and its vibrational signals. Russian Entomological Journal, 23(1): 1-4.
- Berdan, E. L., Finck, J., Johnston, P.R., Waurick, I., Mazzoni, C. J. and Mayer, F. 2017. Transcriptome profiling of ontogeny in the acridid grasshopper *Chorthippus biguttulus*. PLOS ONE, 12(5): e0177367. https://doi.org/10.1371/journal.pone. 0177367.
- Bewick, A. J., Vogel, K.J., Moore, A. J. and Scmitz, R. J. 2017. Evolution of DNA methylation across insects. Oxford Journal of Molecular Biology and Evolution, 34(3): 654-665.

- Bijlsma, R. and Loeschke, V. 2005. Environmental stress, adaptation and evolution: an overview. Journal of Evolutionary Biology, 18: 477-799.
- Bonisoli-Alquati, A., Ostermiller, S., Beasley, D. E., Welch, S. M., Møller, A. P. and Mousseaus, T. A. 2018. Faster development covaries with higher DNA damage in grasshopper (*Chorthippus albomarginatus*) from Chernobyl. Physiological and Biochemical Zoology, 91(2): 776-787.
- Bridle, J. R., Baird, S. J. E. and Butlin, R. K. 2001. Spatial structure and habitat variation in a grasshopper hybrid-zone. Evolution, 55(9): 1832-1843.
- Bultman, S. J. 2015. Metabolic inputs into epigenetics. In: Chadwick, B. P. (Ed.), Epigenetics, current research and emerging trends. Caister Academic Press, pp. 307-326. https://doi.org/10.21775/9781910190074.16.
- Burggren, W. W. 2017. Epigenetics in insects: Mechanisms, phenotypes and ecological and evolutionary implications. Advances in Insect Physiology, 53: 1-30.
- Burggren, W. W. 2018. Developmental phenotypic plasticity helps bridge stochastic weather events associated with climate change. Journal of Experimental Biology, 221: jeb161984. doi: 10.1242/jeb.161984
- Butlin, R. K. and Hewitt, G. M. 2008. Genetic divergence in the *Chorthippus* species group (Orthoptera; Acrididae). Biological Journal of the Linnaean Society, 31(4): 301-310.
- Chapuis, M. P., Crespin, L., Estoup, A., Auge-Sabatier, A., Foucart, A., Lecoq, M. and Michalakis, Y. 2010. Parental crowding influences life-history traits in *Locusta migratoria* females. Bulletin of Entomological Research, 100: 9-17.
- Cigliano, M. M., Braun, H., Eades, D. C. and Otte, D. 2018. *Orthoptera Species File*–Version 5.0/5.0. http://Orthoptera.Species File.org. (Accessed 10 June 2019).
- Contreras, D. and Chapco, W. 2006. Molecular phylogenetic evidence for multiple dispersal events in gomphocerine grasshopper. Journal of Orthoptera Research, 15(1): 91-98.

- Davatchi, A. 1954. Insect Pests of Cereals in Iran. University of Tehran Press. No. 211. Tehran, Iran
- Deaton, A. M. and Bird, A. 2011. CpG island and the regulation of transcription. Genes and Development, 25: 1010-1022.
- Defaut, B. 2011. Révision préliminaire des *Chorthippus* du groupe *binotatus* (Charpentier, 1825) (Caelifera, Acrididae, Gomphocerinae). Matériaux Orthoptériques et Entomocénotiques, 16: 17-54.
- Defaut, B. 2012. Implication taxonomiques et nomenclaturales de publications récentes en Phylogénie moléculaire: 1. Les Gomphocerinae de France (Orthoptera, Acrididae). Matériaux Orthoptériques et Entomocénotiques, 17: 15-20.
- Defaut, B. 2017. Notes de lectures phylogénétiques sur les Orthoptères. Matériaux Orthoptériques et Entomocénotiques, 22: 5-13.
- Dowle, E. J., Morgan-Richard, M. and Trewick, S. A. 2014. Morphological differentiation despite gene flow in an endangered grasshopper. Evolutionary Biology, 14: 216-231.
- Einhaupl, A., Stange, N., Henning, R. M. and Ronacher, B. 2011. Attractiveness of grasshopper songs correlates with their robustness against noise. Behavioral Ecology, 22(4): 791-799.
- Eiriksson, T. 1992. Density dependent song duration in the grasshopper *Omocestus viridulus*. Behavior, 122(1-2): 121-132.
- Elsner, N. 1974. Neuroethology of sound production in Gomphocerine grasshoppers (Orthoptera: Acrididae). I- Song patterns and stridulatorymovements. Journal of Comparative Physiology, 88(1): 67-102.
- Ernst, U. R., Van Hiel, M. B., Depuydt, G., Boerjan, B., De Loof, A. and Schoofs, L. 2015. Epigenetics and locust life phase transition. Journal of Experimental Biology, 218: 88-99.
- Feng, X. J., Jiang, G. F. and Fan, Z. 2015. Identification of outliers in a genomic scan for selection along environmental gradients in the bamboo locust, *Ceracris kiangsu*. Scientific Reports, 5: 13758.

- Franzke, A. and Reinhold, K. 2012a. Limited condition dependence of male acoustic signals in the grasshopper *C. biguttulus*. Ecology and Evolution, 2(8): 1914-1921.
- Franzke, A. and Reinhold, K. 2012b. Transgenerational effects of diet and environment on life-history and acoustic signals of a grasshopper *Chorthippus biguttulus*. Behavioral Ecology, 24(3): 734-739.
- Garcia-Navas, V., Noguerales, V., Cordero, P. and Ortego, J. 2017. Phenotypic disparity in Iberian short-horned grasshopper (Acrididae): The role of ecology and phylogeny. BMC Evolutionary Biology, 17: 109. doi: 10.1186/s12862-017-0954-7.
- Glastad, K. M., Hunt, B.G., Yi, S. V. and Goodisman, M. A. 2011. DNA methylation in insects: on the brink of the epigenetic era. Insect Molecular Biology, 20: 553-565.
- Gomez, G. S. My. and Dyck, H. V. 2012. Ecotypic differenciation between urban and rural populations of the grasshopper *Chorthippus brunneus* relative to climateand habitat fragmentation. Oecologia, 169: 125-133.
- Górür, G. 2005. The importance of phenotypic plasticity in herbivorous insect speciation. In: Ananthakrishnan, T. N. and Whitman, D. (Eds.), Insect phenotypic plasticity: diversity of responses. CRC Press, pp. 145-171.
- Gottsberger, B. 2007. Interspecific hybridization between the grasshoppers *Chorthippus biguttulus* and *C. brunneus* (Acrididae; Gomphocerinae). Doctoral thesis. Faculty of Natural Science, Fredich-Alexander University, Germany, 78 pp.
- Heinrich, R., Kunst, M. and Wirmer, A. 2012. Reproduction-related sound production of grasshopper regulated by internal state and actual sensory environment. Frontiers in Neuroscience, 6: 89.
- Helversen, D. von and Helversen, O. von. 1997. Recognition of the acoustic sex in communication of the grasshopper Chorthippus biguttulus (Orthoptera, Acrididae). Journal Comparative of Physiology, 180: 373-386.
- Helversen, D. von, Balackrishnan, R. and Helversen, O. von. 2004. Acoustic

- communication in a duetting grasshopper: receiver response variability, male strategies and signal design. Animal Behaviour, 68: 131-144.
- Hewitt, G., 2001. Speciation, hybrid zones and phylogeography or seeing genes in space and time. Molecular Ecology, 10: 537-549.
- Hochkirch, A. 2013. Hybridization and the origin of species. Journal of Evolutionary Biology, 26: 247-251.
- Hochkirch, A. and Lemke, I. 2011. Asymmetric mate choice, hybridization and hybrid fitness in two sympatric grasshopper species. Behavioral Ecology and Sociobiology, 65: 1637-1645.
- Hodjat, S. H. 1967. How to Test Pesticides. Entomology and Pest Control Section, Ahwaz Agricultural College Publication, Kayhan Press, Iran, No. 13, 90 pp.
- Hodjat, S. H. 1974. Use of population models in pest control. Shahid Chamran University Press, No. 69/17, 103 pp.
- Hodjat, S. H. 2006. Ecological stress and locust outbreak, an outlook at locust phase research. Scientific Journal of Agriculture, 29(3): 61-74.
- Hodjat, S. H. 2016. Effects of crowding and stress on locusts, aphids, armyworms and specifically the hemipteran *Dysdercus fasciatus* Sign. (Hemiptera: Pyrrhocoridae). Journal of Crop Protection, 5(3): 313-329.
- Hodjat, S. H., Saboori, A. and Husemann, M. 2019. A view on the historic and contemporary acridid fauna (Orthoptera, Caelifera, Acrididae) of Iran- A call for conservation. Journal of Crop Protection, 8(2): 135-142.
- Hodjat, S. H., Tork, M., Seiedy, M. and Defaut, B. 2018. A taxonomic review of recorded species of Caelifera (Orthoptera) in Iran. Matériaux Orthoptériques Etentomologiques, 23: 35-75.
- Hoffmann, A. A. and Parsons, P. A. 1991. Evolutionary Genetics and Environmental Stress. Oxford University Press, Oxford, UK. 284 pp.
- Ingrisch, S. 1995. Evolution of the *Chorthippus biguttulus* group (Orthoptera) in the Alps,

- based on morphology and stridulation. Revue Suisse de Zoologie, 102(2): 475-535.
- Isaksson, C., Sheldon, B. C. and Uller, T. 2011. The challenges of integrating oxidative stress into life-history biology. BioScience, 61: 194-202.
- Ishikawa, R. and Kinoshita, T. 2009. Epigenetic programming: The challenge to species hybridization. Mollecular Plant, 2(4): 589-599.
- Jago, N. D. 1971. A review of the Gomphocerinae of the world, with a key to the genera. Proceedings of the Academy of Natural Sciences of Philadelphia, 123(8): 205-343.
- Jentzsch, A., Kohler, G. and Schumacher, J. 2003. Environmental stress and fluctuating asymmetry in the grasshopper *Chorthippus parallelus*. Zoology, 106: 117-125. doi: 10.1078/0944-2006-00106.
- Job, J. R., Myers, K., Naghshineh, K. and Gill S. A. 2016. Uncovering spatial variation in acoustic environments using sound mapping. PLOS ONE, 11(7): e0159883.
- Ketmaier, V., Stukas, H., Hempel, J., Landeck, I., Tobler, M., Plath, M. and Tiedemann R. 2010. Genetic and morphological divergence among gravel bank grasshoppers, *Chorthippus pullus* (Acrididae), from contrast environments. Organism Diversity & Evolution, 10: 381-395.
- Klappert, K. and Reinhold K. 2003. Acoustic preference: functions and sexual selection on the male calling song in the grasshopper *Chorthippus biguttulus*. Animal Behaviour, 65: 225-233.
- Kohler, G., Sameitz, J. and Schielzeth, H. 2017. Morphological and colour morph clines along an altitudinal gradient in the meadow grasshopper *Pseudochorthippus parallelus*. PLOS ONE, 12(2): e0189815. https://doi.org/10.1371/journal.pone. 0189815.
- Laiolo, P., Illera, J. C. and Obeso, J. R. 2013. Local climate determines intra and variation interspecific in sexual size dimorphism grasshopper in mountain communities. Journal **Evolutionary** Biology, 26: 2171-2183.

- Lampe, U., Reinhold K. and Schmoll, T. 2014. How grasshoppers respond to road noise: Developmental plasticity and population differentiation in acoustic signaling. Functional Ecology, 28: 660-668.
- Lampe, U., Schmoll, T., Franzke, A. and Reinhold, K. 2012. Staying tuned: grasshoppers from many roadside habitats produce courtship signals with elevated frequency components. Functional Ecology, 26: 1948-1354.
- Laws, N. and Belovsky, E. B. 2010. How will species respond to climate change? Examining the effects of temperature and population density on herbivore insects. Environmental Entomology, 39(2): 312-319.
- Lehmann, P., Ammunet, T., Madeleine, B., Battist, A., Eigenbrode, S. D., Jepsen, J. U., Kalinkat, G., Neuvonen, S., Niemelä, P., Økland, B., Terblanche, J. S. and Björkman, C. 2018. Complex responses of global insect pests to climate change. bioRxiv, doi: https://doi.org/10.1101/425488.
- Li, X. D., Jiang, G. F., Yan, L. Y., Li, R., Mu, Y. and Deng, W. A. 2018. Positive selection drove the adaptation of mitochondrial genes to the demands of flight and high altitude environments in grasshoppers. doi: 10.3389/fgene.2018.00605.
- Lonsdale, Z. N. 2018. Epigenetic mechanisms of insect polymorphisms. University of Leicester Doctoral Thesis, Leicester, UK, 232 pp.
- Mallon, E. B., Amarasinghe, H. E. and Ott, S. R. 2016. Acute and chronic gregarisation are associated with distinct DNA methylation fingerprints in desert locusts. Scientific Reports, 6: 35608.
- Mark, K. A. 1983. Morphological variation in relation to habitat in some British grasshoppers. Journal of Natural History, 17(1): 75-85.
- Merritt, T. J. S. and Bewick, A. J. 2017. Genetic diversity in insect metal tolerance. Frontiers in Genetics, 8: 172. doi: 10.3389/fgene.2017.00172.
- Modares Awal, M. 2012. List of agricultural pests and their natural enemies in Iran. Third

- edition, Ferdowsi University of Mashhad Press, Mashhad, Iran, 778 pp.
- Mol, A., Ciplak, B. and Şirin, D. 2003. Song and morphology of the three Anatolian endemic species of the genus *Chorthippus* (Orthoptera: Acrididae, Gomphocerinae). Annales de la Société Entomologique de France (N. S.), 39(2): 121-128.
- Møller, A. P. and Swaddle, J. P. 1997. Asymmetry, developmental stability, and evolution. Oxford University Press, Oxford, UK, 304 pp.
- Monge, J. P. V. 2017. Phenotypic plasticity and condition-dependent trait expression in sexual ornaments and body colour in alpine grasshopper *Gomphocerus sibiricus*. Ph. D. Dissertation, University of Bielefeld, Bielefeld, Germany, 97 pp.
- Murren, C. J., Auld, J. R., Callahan, H., Ghalmbor, C. K., Handelsman, C. A., Heskel, M. A., Kingsolver, J. G., Maclean, H. J., Mase, J., Maughan, H., Pfening, D. W., Relyea, R. A., Seiter, S., Snell-Rood, E., Steiner, U. K. and Schlichting, C. D. 2015. Constraints on the evolution of phenotypic plasticity: limits and costs of phenotype and plasticity. Heredity, 115: 293-301.
- Nattier, R., Robillard, T., Amédégnato, C., Couloux, A. and Cruaud, C. 2011. Evolution of acoustic communication in the Gomphocerinae (Orthoptera:Caelifera: Acrididae). Zoologica Scripta, 40(5): 479-497.
- Nettle, D. and Bateson, M. 2015. Adaptive developmental plasticity, what is it, how can we recognize it and when can it evolve? Proceedings of the Royal Society B, 282: 20151005. doi: 10.1098/rspb. 2015.1005.
- Noguerales, V., Cordero P. J. and Ortego, J. 2016. Hierarchical genetic structure shaped by topography in a narrow-endemic mountain grasshopper. BMC Evolutionary Biology, 16: 96.
- Noguerales, V., Cordero, P. J. and Ortego, J. 2018. Integrating genomic and phenotypic data to evaluate alternative phylogenetic and species delimitation hypothesis in a recent evolutionary radiation of grasshopper.

- Molecular Ecology, 27(5): 1229-1244. doi: 10.1186/s12862-016-0663-7.
- Norouzitallab, P., Baruelh, B., Vanrompey, D. and Bossier, P. 2019. Can epigenetics translate environmental cues into phenotypes? Science of The Total Environment, 647: 1281-1293.
- O'grady, P. M. and Markow, T. A. 2012. Rapid morphological, ecological and behavioral evolution in *Drosophila*: Comparison between the *Cactophilic replete* species group and the endemic Hawaian *Drosophila*. In: Singh, R. S., Xu, J. P. and Kulathinal, R. J. (Eds.), Rapidly Evolving Genes and Genetic Systems. Oxford Scholarship Online, Oxford, UK., pp. 176-186.
- Oxford Research Group, 2015. Effects of environmental stressors. Oxford research encyclopedia of environmental science on various topics. doi: 10.1093/acrefore/9780199389414.013.1
- Parsons, P. A. 1991. Evolutionary rates: stress and species boundaries. Annual Review of Ecology and Systematics, 22: 1-18.
- Parsons, P. A. 1992. FA a biological monitor of environmental and genomic stress. Heredity, 68: 117-125.
- Perić-Mataruga, V., Nenadovic, V. and Ivanovic, J. 2006. Neurohormones in insect stress: a review. Archives of Biological Sciences, 58(1): 1-12.
- Pravdin, F. N. 1969. The formation of subspecies in Acridoidea (Orthoptera) in the mountains of Middle Asia. Beiträgezur Entomologie, 19: 647-657.
- Qin, X., Ma, J., Huang, X., Kallenbach, R. L., Lock, T. R., Ali, Md. P. and Zhang, Z. 2017. Population dynamics and transcriptomic responses of *Chorthippus albonemus* (Orthoptera: Acrididae) to herbivore grazing intensity. Frontiers in Ecology and Evolution, 5: 136. doi: 10.3389/fevo. 2017.00136.
- Riede, K., 2018. Acoustic profiling of Orthoptera: present state and future needs. Journal of Orthoptera Research, 27(2): 203-215.
- Ritchie, M. G. 1989. Assortative mating across a hybrid zone in *Chorthippus parallelus*

- (Orthoptera; Acrididae). Journal of Evolutionary Biology, 2(5): 339-352.
- Ritchie, M. G. 1990. Are differences in song responsible for assortative mating between subspecies of the grasshopper *Chorthippus parallelus* (Orthoptera; Acrididae)? Animal Behaviour, 39: 685-691.
- Robinson, D. J. and Hall, M. J. 2002. Sound signaling in Orthoptera, In: Evans, P. (Ed.), Advances in insect physiology. Volume 29. Elsevier, pp. 151-278.
- Rogers, S. M., Matheson, T., Despland, E.,
  Dodgson, T., Burrows, M. and Simpsons,
  M. 2003. Mechanosensory-induced
  behavioural gregarization in the desert locust
  Schistocerca gregaria. Journal of
  Experimental Biology, 206: 3991-4002.
- Rohde, K., Dreher, E. and Hochkirch, A. 2015a. Sex-specific phenotypic plasticity in response to the trade-off between developmental time and body size supports the dimorphic niche hypothesis. Biological Journal of the Linnaean Society, 115: 48-57.
- Rohde, K., Hau, Y., Kranz, N., Weinberger, J., Elle, O. and Hochkirch, A. 2017. Climate effect on population decline of a rare wetland species and the role of spatial and temporal isolation as barrier to hybridization. Functional Ecology, 31(6): 1262-1274.
- Rohde, K., Weyer, H. J. and Hochkirch, A. 2015b. Wide prevalence of hybridization in two sympatric grasshopper species may be shaped by their relative abundances. BMC Evolutionary Biology, 15: 191. doi: 10.1186/s12862-015-0460-8.
- Romer, H. 1993. Environmental and biological constraints for the evolution of long-range signaling and hearing in acoustic insects. Philosophical Transactions of the Royal Society B, 226: 179-185.
- Ronacher, B. 1989. Stridulation of acridid grasshoppers after hemisection of the thoracic ganglia; Evidence for hemiganglionic occillators. Journal of Comparative Physiology, 164: 723-736.
- Saldamando, C. I., Miyaguchi, S., Tatsuta, H., Kishino, H., Bridle, J. R. and Butlin, R. K.

- 2005. Inheritance of song and stridulatory peg number divergence between *C. brunneus* and *C. jacobsi*, two naturally hybridizing grasshopper species (Orthoptera; Acrididae). Journal of Evolutionary Biology, 18: 703-712.
- Sardnick, J., Klofel, A., Elner, N. and Vedenina, V. 2016. Variation in complex mating signals in an "island" hybrid zone between *Stenobothrus* grasshopper species. Ecology and Evolution, 6(14): 5057-5075.
- Sergeev, M. 2011. Distribution patterns of grasshoppers and their kin in the Boreal Zone. Psyche, 324130.
- Sgro, C. M., Terblanche, S. and Hoffmann A. A. 2016. What can plasticity contribute to insect responses to climate change? Annual Review of Entomology, 61: 431-451.
- Şirin, D., Helversen O. von and Ciplak, B. 2010. *Chorthippus brunneus* subgroup (Orthoptera, Gomphocerinae) in Anatolia with description of two species: data suggest an Anatolian origin for the lineage. Zootaxa, 2410: 1-28.
- Şirin, D., Mol, A. and Akyildiz, G. 2014. The morphological and behavioral analysis of geographically separated *Rammeihippus turcicus* (Orthoptera, Acrididae Gomphocerinae) populations: data result in taxonomic conflict. Journal of Insect Science, 14: 145. doi: 10.1093/jisesa/jeu007.
- Skinner, M. K. 2014. Environmental stress and epigenetic transgenerational inheritance. BMC Medicine, 12: 153.
- Smadja, C. and Butlin, R. 2011. A framework for comparing processes of speciation in the presence of gene flow. Molecular Ecology, 20(24): 5123-5140.
- Smith, G. and Ritchie, M. G. 2013. How might epigenetics contribute to ecological speciation? Current Zoology, 59(5): 686-696.
- Song, H. 2005. Phylogenetic perspectives on the revolution of locust phase polymorphism. Journal of Orthoptera Research, 14(2): 235-245.
- Song, H. 2011. Density-dependent phase polymorphism in nonmodel locusts: A mini review. Psyche, 741769.

- Steiner, A. 2006. Contribution to the nutritional ecology of the endangered grasshopper *Chorthippus pullus* (Philippe, 1830). Ph. D. Thesis, University of Bern, Bern, Switzerland.
- Stillwell, R. C., Blanckenhorn, W. U., Teder, T., Davidowitz, G. and Fox, C. W. 2010. Sex differences in phenotypic plasticity of body size affect variation in sexual size dimorphism in insects: from physiology to evolution. Annual Review of Entomology, 55: 227-245.
- Stumpner, A. and Helversen, O. von. 1994. Song production and song recognition in a group of sibling grasshopper species (*Chorthippus dorsatus*, *C. dichrous* and *C. loratus*: Orthoptera, Acrididae). Bioacoustics, 6(1): 1-23.
- Sultana, R., Wagan, Y. S. and Waga, M. S. 2013. Orthopteran biodiversity of Thar Desert, Sindh, Pakistan. Pakistan Journal of Zoology, 45(2): 299-304.
- Sword, G. A. 2002. A role of phenotypic plasticity in the evolution of aposematism. Proceedings of the Royal Society of London B, 269: 1639-1644.
- Sword, G. A. 2003. To be or not to be a locust? A comparative analysis of behavioral phase change in nymphs of *Schistocerca Americana* and *S. gregaria*. Journal of Insect Physiology, 49: 709-717.
- Tatsuta, H. and Butlin, R. K. 2001. Amplified fragment length polymorphism for the analysis of genetic structure in grasshopper populations. Journal of Orthoptera Research, 10(2): 203-207.
- Tetreau, G. 2018. Interaction between insects, toxins, and bacteria: Have we been wrong so far? Toxins, 10(7): 281-311.
- Tishechkin, D. Yu. and Bukhvalova M. A. 2009. Acoustic communication in grasshopper communities (Orthoptera: Acrididae: Gomphocerinae): segregation of acoustic niches. Russian Entomological Journal, 18(3): 165-188.
- Tregenza, T., Prichard, V. L. and Butlin, P. K. 2000. Pattern of trait divergence between populations of the meadow grasshopper

- *Chorthippus parallelus*. Evolution, 44(2): 574-585.
- Tricker, P. J. 2015. Transgenerational inheritance or resetting of stress-induced epigenetic modifications: two sides of the same coin. Frontiers in Plant Science, 6: 699.
- Uvarov, B. P. 1957. The aridity factor in the ecology of locusts and grasshoppers of the world. Arid Zone Research, 8: 164-198.
- Valverde, J. P. and Schielzeth, H. 2015. What triggers colour change? Effects of background colour and temperature on the development of an alpine grasshopper. BMC Evolutionary Biology, 515: 168.
- Vedenina V. Yu. and Mugue, N. 2011. Speciation in Gomphocerinae Grasshoppers; Molecular phylogeny versus bioacoustic courtship behavior. Journal of Orthoptera Research, 20(1): 109-125.
- Vedenina V. Yu., Kuligina, N. K. and Panyutin, A. K. 2007a. Isolation mechanism in closely related Grasshopper species *Chorthippus albomarginatus* and *Ch. oschei* (Orthoptera; Acrididae). Entomological Review, 87(3): 263-272.
- Vedenina V. Yu., Panyutin A. K. and Helversen, O. von. 2007b. The unusual inheritance pattern of the courtship songs in closely related grasshopper species of the *Chorthippus albomarginatus* group (Orthoptera; Gomphocerinae). Journal of Evolutionary Biology, 20: 260-277.
- Vedenina, V. Yu. 2015. Courtship song analysis in two hybrid zones between sibling species of the *Chorthippus albomarginatus* group (Orthoptera, Gomphocerinae). Entomological Review, 95(2): 94-108.
- Vedenina, V. Yu. and Helversen, O. von. 2003. Complex courtship in bimodal grasshopper hybridzone. Behavioral Ecology and Sociobiology, 54(1): 44-54.
- Vedenina, V. Yu. and Helversen, O. von. 2009. A re-examination of the taxonomy of the *Chorthippus albomarginatus* group in Europe on the basis of song and morphology (Orthoptera; Acrididae). Tijdschrift voor Entomologie, 152(1): 65-97.

- Vedenina, V. Yu. and Shestakov, L. S. 2014. Stable and variable parameters in courtship songs of grasshoppers of the subfamily Gomphocerinae (Orthoptera, Acrididae). Entomological Review, 94(1): 1-20.
- Vedenina, V. Yu., Heinrich, R. and Elsner, N. 2001. Pharmacological activation of stridulation in the grasshopper *Chorthippus albomarginatus* (Orthoptera; Gomphocerinae). Journal of Evolutionary Biochemistry and Physiology, 37(4): 388-397.
- Via, S. 2012. Divergence hitchhiking and the spread of genomic isolation during ecological speciation with gene flow. Philosophical Transactions of the Royal Society B: Biological Sciences, 367(1587): 451-460.
- Wall, R. 1987. Individual variation and the effects of population density in the grasshopper *Chorthippus brunneus*. Oikos, 49(1): 15-27.
- Werner, S. A. and Toth, A. L. 2012. Epigenetics in social insects: A new direction for understanding the evolution of casts. Genetics Research International, 609810. http://dx. doi.org/10.1155/2012/609810.
- Whitman, D. W. and Agrawal A. A. 2009. What is phenotypic plasticity and why is it important? In: Whitman, D. W. and Ananthakiishnan, T. N. (Eds.), Phenotypic plasticity of insects, mechanisms and consequences. CRC Press, Boca Raton, USA, pp. 1-63.
- Zera, A. J. and Harshman, L. G. 2001. The physiology of life history trade-offs in animals. Annual Review of Ecology and Systematics, 32: 95-126.
- Zhang, H. L., Huang, Y., Lin, L. L., Wang, X. Y. and Zheng, Z. M. 2013. The Phylogeny of the Orthoptera (Insecta) deduced from mitogenomic gene sequences. Zoological Studies, 52: 37.
- Zinna, R. A., Gotoh, H., Kojima, T. and Niimi, T. 2018. Recent advances in understanding the mechanisms of sexually dimorphic plasticity: insights from beetle weapons and future directions. Current Opinion in Insect Science, 25:35-41.

## مروری بر تأثیرات تغییر، انعطافپذیری و اپیژنتیک احتمالی در گونههای قبیله Gomphocerini (Orthoptera; Gomphocerinae)

سیدحسین حجت و علیرضا صبوری\*

موزه جانورشناسی استاد جلال افشار، گروه گیاهپزشکی، پردیس کشاورزی و منابع طبیعی دانشگاه تهران، کرج، ایران. پست الکترونیکی نویسنده مسئول مکاتبه: saboori@ut.ac.ir دریافت: ۲۵ شهریور ۱۳۹۸؛ پذیرش: ۲۴ مرداد ۱۳۹۹

چکیده: شرایط محیطی موجب تغییر در ریختشناسی، رفتار و احتمالاً اپیژنتیک در بسیاری از گونههای زیرخانواده Gomphocerinae بهویژه در زیستگاههای کوهستانی میشود. انعطاف پذیری و تغییرات در ریختشناسی بسیاری از گونهها در این زیرخانواده در اثر جدایی ویژگیها در طبی انتخباب ماده بیرای جفتگیری ایجاد میشود که کشانههای مختلف، گروههای زیرگونهای یا گونهای تشکیل میدهد. تغییرات و انعطاف پذیری در اثر استرسهای محیطی افزون بر ریختشناسی، بر فیزیولوژی و اپیژنتیک بسیاری از گونههای حشرات تأثیر میگذارد. استرسهای محیطی و جفتگیری با ماده مناسب ممکن است با تولید دورگ در جمعیت همراه شده و منجر به جدایی صفات و گونهزایی در درازمدت شود. تکامل معاصر و یا وراثت اپیژنتیک ممکن است دلیل بر تغییرات در آوازخوانی و ریختشناسی Gomphocerinae بوده و عامل اصلی در مشکلات کنونی در ردهبندی آنهاست. در این مقاله، اثرات احتمالی استرسهای محیطی، دورگ گیری و گونهزایی با ظهور گونههای بومی مرور میشود. تأثیر حدود نیمی از گونههای حشرات آفت در بهعنوان آفت در اثر تغییر در شرایط محیطی کنونی کهرنگ شده است. وضعیت کنونی حشرات آفت در ایران مورد بحث قرار گرفته است.

واژگان کلیدی: ردهبندی، دورگ گیری، Gomphocerini، فنوتایپ، آواز