doi: 10.18869/modares.jcp.5.3.431

Research Article

Demographic traits of *Hippodamia variegata* (Goeze) (Coleoptera: Coccinellidae) fed on *Sitobion avenae* Fabricius (Hemiptera: Aphididae)

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Abstract: Lady beetles are the most important predatory species among arthropods, so studying their population parameters gives a clear picture about their life span. The life histories of variegated lady beetle, Hippodamia variegata (Goeze), fed on the English grain aphid, Sitobion avenae Fabricius, on different host plants were studied under laboratory conditions, at 25 ± 1 °C, $65 \pm 5\%$ Relative Humidity and a photoperiod of 16:8 (Light: Dark) h. Developmental, survival and fecundity data were analyzed using the age-stage, two-sex life table method. According to the age-stage, two-sex life table, the net reproductive rates (R_0) of ladybird fed on S. avenae reared on Triticum aestivum var. tajan, Hordeum vulgare var. Dasht, Zea mays var. single cross 704 and Sorghum durra var. Speed feed were estimated to be 235.38 ± 36.15 , 190.74 ± 33.37 , 293.54 ± 36.15 46.66 and 137.46 ± 23.66 female offspring / female, respectively. Based on the results, the intrinsic rate of increase as an index of population increase of H. variegata, was 0.181 ± 0.006 , 0.179 ± 0.006 , 0.163 ± 0.006 and 0.162 ± 0.006 h preyed on host reared on T. aestivum var. tajan Z. mays var. single cross 704 S. durra var. Speed feed and H. vulgare var. Dasht, respectively. Our findings demonstrate that by using the age-stage, two-sex life table we can accurately describe the growth, survival and development of the predator. Our finding may provide basic information for developing aphid biological control programs.

Keywords: Two Sex, Demography, *Hippodamia variegata*, *Sitobion avenae*, host plant

Introduction

Biological control is the major component of Integrated Pest Management (IPM) strategies. The aim of biological control is to reduce pest populations through natural enemies, such as predators, parasitoids and pathogens (Gilkeson and Kelin, 2001). A successful natural enemy is

Handling Editor: Jafar Mohaghegh Neyshabori

*Corresponding author, e-mail: ghafourim@uoz.ac.ir Received: 3 April 2015, Accepted: 1 April 2016 Published online: 28 June 2016 one, which has high reproduction rate, and good searching ability for its host, adaptability in different environmental conditions synchronization with its host (Buchanan, 1996). Among all beneficial insects and arthropods, lady beetles are the most well-known predators all around the world (William, 2002). In fact, during their adult and larval stages, coccinellids attack to a variety of soft bodied herbivorous pests, including aphids, whiteflies, psyllids, and scales. Moreover they ingest fungal spores, pollen and nectar as complementary or even alternative foods when the preys are rare (Ali et al., 2009). In many studies, it has been shown that *Hippodamia variegata* (Goeze) is an efficient predator and can be used for the biological control of aphids (Franzman, 2002). This coccinellid is an effective predator of aphids and is used as biocontrol agent against a wide range of aphid species in different crops such as wheat, cotton, tobacco, vegetables and orchards either in outdoor farms or in greenhouses (Cabral *et al.*, 2008). Due to the big size and high feeding capacity, this generalist-variegated beetle is proposed as valuable biological control agent (Kontodimas and Stathas, 2005) and is useful in insect pest management programs.

Due to the importance of predatory ladybird beetles in aphid biological control, it is necessary to investigate the factors influencing the resource quality of aphids as their prey. Among these factors, plant species can affect not only the herbivorous insects feeding on them, but also parasitoids and predators directly indirectly, through multi-trophic or/and interactions (Price et al., 1980). Plant biochemistry may influence the nutritional value of herbivorous insects and in turn affect the various life history traits of their natural enemies (Giles et al., 2002). Prey quality has a direct impact on the growth, development and reproduction of predatory insects (Thompson, 1999), and this effect is displayed by their differential efficiencies (Babu, 1999). The relative suitability of a prey species can be evaluated by measuring its impact on the biological characteristics of the predator (Kalushkov and Hodek, 2001). Preys are categorized as essential, alternative or rejected on the basis of quantitative data on the developmental rate, survival and reproductive capacity (Hodek and Honek, 1996). Recently, some information has become available on the relative suitability of aphid species for some important ladybird beetles (e.g., Takizawa et al., 2000; Evans et al., 2004; Tsaganou et al., 2004; Kalushkov and Hodek, 2001, 2004; Omkar and Srivastava, 2003; Omkar and Bind, 2004; Pervez and Omkar, 2004; Ali and Rizvi, 2007; Golizadeh and Jafari-Behi, 2012). However, supporting data are limited (Wu et

al., 2010). Some studies indicated that the suitability of prey was affected not only by the host plant species (Rü and Mitsipa, 2000; Francis *et al.*, 2001; Al-Zyoud *et al.*, 2005), but also by the cultivar (Du *et al.*, 2004).

Demography has been applied to study enemies' potential in biocontrol programs (Bellows et al., 1992) improvement of rearing techniques in the mass production of many species (Carey and Vargas, 1985; Lido and Carey, 1994). A proper assessment of predation potential should include the evaluation of a life table and only life tables can provide a comprehensive description of the development, survival, and fecundity of a population (Farhadi et al., 2011). For most insects, the developmental rates vary among individuals and between the sexes (Istock, 1981; Chi and Liu, 1985; Carey, 1993). Chi and Liu (1985) and Chi (1988) developed an age-stage, two-sex life table that takes into consideration stage differentiation and the male population. The age-stage, two-sex life table has been used to describe the population characteristics of many insect and mite species under the influence of a variety of physical conditions. Chi and Liu (1985) and Chi (1988) also demonstrated that the traditional female age-specific life tables (e.g., Lewis, 1942; Leslie, 1945; Birch, 1948) ignore the males and the stage differentiation.

Although past publications on H. variegata focused mainly on the biological characteristics (Fan et al., 1995; An et al., 2000), functional response (Fan and Zhao, 1988; Feng et al., 2000; Pang et al., 2000), life table (Lanzoni et al., 2004; Kontodimas and Stathas, 2005) and the influence of temperature on its development (Michels and Bateman, 1986; Michels and Flanders, 1992), no studies have been carried out on influence of different host plants of wheat green aphid, Sitobion avenae (Fabricius) (Hemiptera: Aphididae) on some population parameters of ladybird, *H. variegata*. Only Wu et al. (2010) investigated Influence of five host plants of Aphis gossypii Glover on some population parameters of *H. variegata*.

As the fitness of a mass-produced natural enemy depends on many factors as well as food

quality, the present research was carried out to determine development, mortality, longevity, reproduction and the most suitable prey for ladybird *H. variegata* as influenced by host plants species in the laboratory.

Materials and Methods

Collecting and rearing of *H. variegata* and *S. avenae*

The English grain aphid, *S. avenae* Fabricius, was collected from wheat fields at the experimental field of Mohaghegh Ardabili University in the suburb of Ardabil city, Iran, in April 2012 and transferred onto four different host plants (*Triticum aestivum* var. *tajan, Hordeum vulgare* var. Dasht, *Zea mays* var. Single cross 704 and *Sorghum durra* var. Speed feed) cultivated in a greenhouse at 25 ± 5 °C, 50-70% RH and natural light conditions at the Department of Plant Protection, College of Agriculture, University of Mohaghegh Ardabili. These four plant species are main hosts of *S. avenae* in Iran.

H. variegata adults used in this study were originally collected from wheat fields at the above mentioned experimental field and brought to the laboratory. Then some pairs of ladybirds were selected for oviposition. Mating pairs were kept together in separate clear plastic cups (15 cm diameter, 8 cm height), with moist filter papers at the bottom (Wu et al., 2010). Each pair was provided with S. avenae populations from four different host plants described above and maintained in a growth chamber set at 25 ± 1 °C, 65 ± 5 % RH and 16L: 8Dh photoperiod. Laid eggs of each pair were removed from the culture every 24 h and were kept separately in Petri dishes and were used to rear a new generation. The adult predators and larvae were fed daily with the aphid species. Aphids were replaced daily with fresh ones to avoid microbial contamination (Omkar and James, 2004). Predator populations were reared for at least one generation on aphid prey before being used for life table study.

Pre-imaginal development and survival

Only eggs laid within 24 h were used for the

experiments. For investigation of pre-imaginal development and survival on different host plants, groups of about 50 eggs of H. variegata were obtained from the adults reared on four different host plants, and placed separately in Petri dishes (15 cm diameter, 8 cm height) in an incubator cabinet at 25 ± 1 °C, $60 \pm 10\%$ RH and 16:8h [L: D] photoperiod. Egg hatching and incubation period were recorded daily. After hatching, all emerged first instar larvae for each plant species were carefully transferred and isolated individually into a Petri dish with a fine hairbrush to avoid cannibalism. The first instars were supplied with a specific host plant aphid complex until their pupation. Aphids and host plant leaves were replaced daily with fresh ones to avoid any microbial contamination. Larval, pupal periods and survival were recorded daily.

Reproductive attributes and longevity

After adult emergence, each pair was isolated in a Petri dish for mating and provided with the above-mentioned specific host plant-aphid complex daily. Eggs were counted daily and transferred to another Petri dish, which was labelled and cultured under the same conditions to calculate egg hatching rates on different host plants. Number of eggs deposited was counted daily and the experiments were continued until the death of all individuals. In the current study, adult pre-oviposition period (APOP: the period of time between the emergence of an adult female insect and the initiation of its oviposition), total pre-oviposition period (TPOP: the duration from egg to first oviposition), oviposition period, total fecundity (eggs during the reproductive period) and adult longevity were recorded. Based on the data obtained from the above experiments, life tables and population growth parameters of H. variegata on four host plants were calculated (Birch, 1948; Kontodimas and Stathas, 2005).

Age-stage, two-sex life table

Variables were tested for normality using the Kolmogorov-Smirnov test before subjecting them to analysis (SPSS, 2007). The raw life

history data for *H. variegata* on the developmental time of all individuals, including males, females, and those dying before the adult stage, survivorship, longevity, and female daily fecundity of individuals were analyzed according to the theory of age-stage, two-sex life table (Chi and Liu, 1985; Chi, 1988) using the computer program TWOSEX-MSChart (Chi, 2013)

Following Chi and Liu (1985), the age-stage specific survival rate (s_{xj}) (where x = age and j =stage), the age-specific survival rate (l_x) , the agestage specific fecundity (f_{xj}) , the age-specific fecundity (m_x) , and the population parameters (r_m) the intrinsic rate of increase; λ , the finite rate of increase,; R_0 , the net reproductive rate; T, the mean generation time) were calculated survival accordingly. The age-specific includes both male and female, the age-specific fecundity, and net reproductive rate calculated according to Chi and Liu (1985) as:

$$l_x = \sum_{i=1}^m S_{xi} \tag{1}$$

and

$$m_{x} = \frac{\sum_{j=1}^{m} S_{xi} f_{xj}}{\sum_{j=1}^{m} S_{xj}}$$
 (2)

and

$$R_o = \sum_{x=0}^{\infty} l_x m_x \tag{3}$$

In this paper, the intrinsic rate of increase was estimated using the iterative bisection method and the Euler–Lotka equation with the age indexed from 0 (Goodman, 1982):

$$\sum_{n=0}^{\infty} e^{-r(x+1)} l_x m_x = 1 \tag{4}$$

The mean generation time is defined as the length of time that a population needs to increase to R_0 -fold of its original size as the stable age distribution and the stable increase rate are reached, i.e., $e^{rT} = R_0$ or $\lambda^T = R_0$. Thus, it is calculated as $T = (\ln R_0)/r$. The gross reproductive rate (GRR) is calculated as GRR = 1

 $\sum m_x$. The finite rate (λ) was calculated as follows:

$$\lambda = e^r \tag{5}$$

Because life table studies are extremely time consuming and replication is impractical, we used the bootstrap techniques (Efron and Tibshirani, 1993) to calculate the means and standard errors of the population parameters.

Bootstrap technique

In the bootstrap procedure, we randomly take a sample of n individuals from the cohort with replacement and calculate the R_{θ} , i-boot for this bootstrap sample as:

$$R_{o,i-boot} = \sum_{x=0}^{\infty} l_x m_x \tag{6}$$

where the subscript *i-boot* represents the *i*th bootstrap and lx and m_x are calculated from the n individuals selected randomly with replacement. Generally, the data on the same individual are repeatedly selected. We repeat this procedure m times (m > 200, according to Meyer $et\ al.$, 1986) and compute the mean of these m bootstraps as:

$$R_{o,B} = \frac{\sum_{i=1}^{m} R_{0,i-bool}}{m} \tag{7}$$

The variance S_B^2 and standard error se $(R_{\alpha B})$ of these m bootstraps can be calculated as:

$$S_B^2 = \frac{\sum_{i=1}^m (R_{o,i-boot} - R_{o,B})^2}{m-1}$$
 (8)

$$Se(R_{o,B}) = \sqrt{S_B^2} \tag{9}$$

The same method is used for the corresponding estimates of the gross reproductive rate, i.e., GRR_{i-boot} , GRR_{B} , and the variance and standard error of GRR_{B} .

Effects of host plants on the developmental time, reproduction period and female and male adult longevity were analyzed with one-way ANOVA. If significant differences were detected, multiple comparisons were made using the SNK procedure (P < 0.05). Statistical

analysis and draw figures were performed using the SPSS v. 16.0 statistical program (SPSS, 2007) and Systat SigmaPlot v. 12.2, respectively.

Results

Age-stage, two-sex life table

The means of developmental periods, adult longevity and fecundity of H. variegata fed by aphids reared on four host plants are given in Table 1. All four host plant species of S. avenae were suitable for the development of *H. variegata* (Table 1). The mean durations of egg incubation, first, second and fourth instars development times varied significantly. There were also significant differences among the pupal periods on the four aphid host plant species. However, third instar periods did not vary significantly among the four aphid host plants. Moreover, host plant species of S. avenae significantly influenced the total developmental period of predators; the complete pre-adult duration was longest (16.31 \pm 0.15 days) when reared on *H. vulgare* var. Dasht and shortest $(14.89 \pm 0.13 \text{ days})$ when reared on T. aestivum var. *tajan*. Female and male adults of *H. variegata* when reared on *Z. mays* var. Single cross 704 lived an average of 53.28 ± 1.31 and 47.88 ± 1.47 days, which was slightly longer than that of the female and male adults when reared on *S. durra* var. Speed feed i.e., 39.10 ± 1.54 and 32.66 ± 1.50 days (Table 1). The total recorded immature survival rates were 88, 94, 92 and 86 % on the examined four host plants, respectively.

The APOP of the *H. variegata* was not significantly different regarding the host plants. However, the host plants tested showed significant effects on the TPOP (P < 0.05) of this predator, which was the longest on *S. durra* var. Speed feed (20.68 \pm 0.33 days) and the shortest on *T. aestivum* var. *tajan* (19.36 \pm 0.35 days). There was a significant difference in the total number of eggs per individual of *H. variegata* on the four host plants (P < 0.05), which was highest on *Z. mays* var. Single cross 704 (587.31 \pm 15.71) and lowest on *S. durra* var. Speed feed (362.47 \pm 16.42) (Table 1). Mating happened all along the adult life (Table 1).

Table 1 Developmental durations in days of the different stages, adult longevity, fecundity, preovipostional period, total preovipostional period of *Hippodamia variegata* fed on aphids reared on four host plants at 25 °C.

Host plant	<i>Triticum aestivum</i> var. Tajan	Hordeum vulgare var. Dasht	Zea mays var. Single cross 704	Sorghum durra var. Speed feed
Egg (d)	2.18 ± 0.05^{b}	2.20 ± 0.06^{b}	2.54 ± 0.07^a	2.48 ± 0.07^a
1 st instar (d)	2.35 ± 0.08^b	2.90 ± 0.10^a	2.70 ± 0.09^a	2.82 ± 0.09^a
2 nd instar (d)	2.34 ± 0.08^b	2.52 ± 0.08^b	2.57 ± 0.09^{b}	2.81 ± 0.08^a
3 rd instar (d)	1.89 ± 0.10^a	1.83 ± 0.11^{a}	1.86 ± 0.08^a	1.71 ± 0.09^{a}
4 th instar (d)	3.16 ± 0.11^{b}	3.66 ± 0.07^a	3.13 ± 0.09^{b}	3.23 ± 0.12^{b}
Pupal period (d)	2.86 ± 0.05^{c}	3.15 ± 0.06^{a}	3.11 ± 0.08^{ab}	2.93 ± 0.05^{bc}
Total pre-adult (d)	14.89 ± 0.13^{b}	16.31 ± 0.15^{a}	15.89 ± 0.14^{a}	16.02 ± 0.09^a
Female adult longevity(d)	48.40 ± 1.60^b	44.40 ± 1.27^b	53.28 ± 1.31^a	39.10 ± 1.54^{c}
Male adult longevity (d)	42.28 ± 1.64^{b}	38.40 ± 1.31^{b}	47.88 ± 1.47^{a}	$32.66 \pm 1.50^{\circ}$
Fecundity (eggs/female)	470.34 ± 19.08^b	432.79 ± 14.38^{c}	587.31 ± 15.71^{a}	362.47 ± 16.42^d
Oviposition period (d)	31.32 ± 0.25^{b}	29.68 ± 0.18^{b}	36.68 ± 0.19^a	24.37 ± 0.28^{c}
$APOP^{1}(d)$	4.32 ± 0.28^a	4.27 ± 0.24^a	3.88 ± 0.21^a	4.63 ± 0.29^a
$TPOP^{2}(d)$	19.36 ± 0.35^{b}	$20.73 \pm 0.33^{\rm a}$	19.76 ± 0.23^{ab}	20.68 ± 0.33^{a}

¹⁻ APOP: Adult pre-oviposition period, 2- TPOP: total pre-oviposition period. Means followed by the same letters in a row are not significantly different, (P < 0.05, SNK).

To illustrate the difference between male and female survival, the survival rates (sxj) of both male and female individuals are shown in Fig. 1. The means and standard summarized errors the descriptive statistics (Table 1), whereas the survival curves depict the detailed survival and stage differentiation process of the cohort. The overlaps between different stages during a development period demonstrate the variable developmental rates among individuals (Fig. 1). For adult male 0.388, 0.508, 0.420, 0.488 and female 0.506, 0.447, 0.508 and 0.382 on mentioned host plants, respectively. The

number of offspring produced by an individual H. variegata of age x and stage j is shown in Fig. 2. Because only females produce eggs, there is only a single curve, f_{x7} , which represents females in the seventh life stage. The parameters l_x , m_x , and agespecific maternity ($l_x m_x$) are also plotted in Fig. 2. The curve of l_x is a simplified version of the age-stage survival rate (s_{xj}). The most daily fecundity (f_{x7}) obtained 16.36, 18.18, 17.00 and 16.84, which occurred in days 42, 39, 41 and 25 respectively. The curves of m_x show roughly periodic peaks in reproduction on barley and corn.

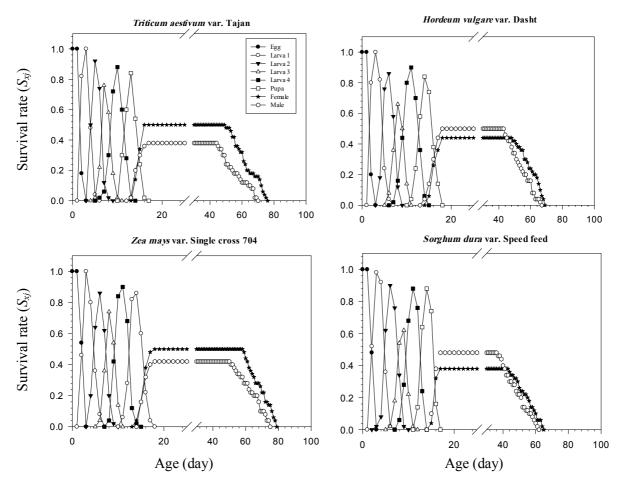


Figure 1 Age-stage specific survival rates of *Hippodamia variegata* fed on aphids reared on four host plants at 25 °C.

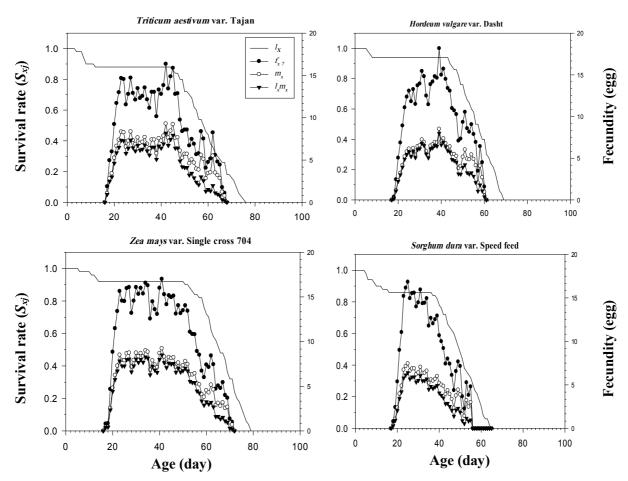


Figure 2 Age specific survival rates (l_x) , fecundities (m_x) , maternities $(l_x m_x)$ and age-stage specific fecundities (f_{x7}) of *Hippodamia variegata* fed by aphids reared on four host plants at 25 °C.

Population parameters

The means and standard errors of the population parameters that were estimated by employing the bootstrap techniques (Efron and Tibshirani, 1993) are listed in Table 2. The GRR of H. variegata showed significant differences among four host plants, and the highest value was observed on Z. mays var. Single cross 704 (334.80 \pm 48.62 eggs). The R_0 of H. variegata was significantly different and highest when fed by aphids reared on Z. mays var. Single cross 704 (291.14 \pm 46.66) and lowest on S. durra var. Speed feed (134.30 \pm 23.66) (Table 2). Similarly, the r and λ value exhibited a significant difference on four host plants, which were highest on aphid- T. aestivum var. tajan whereas the lowest was on the aphid- H. vulgare var. Dasht diet. The T was shortest on the aphid- S. durra var. Speed feed diet (29.87 \pm 0.33) and longest on aphid-H. vulgare var. Dasht (32.03 \pm 0.44). Therefore, based on the intrinsic rate of population increase as an index of suitability of prey host plant species on the population increase of H. variegata, the suitability in decreasing order was: T. aestivum var. tajan > Z. mays var. tajan > Z. ta

Chi (1988) demonstrated that the relationship between the mean female fecundity (F) and the net reproductive rate (R_0) can be described as:

$$R_0 = \left(\frac{N_f}{N}\right) F \tag{10}$$

where N is the total number of individuals used at the beginning of the life table study (i.e., 50 eggs), and N_f is the number of female adults emerging from these N eggs. It also means $N_f \times F = R_0 \times N$. In other words, the total number of offspring produced by all females equals the net reproductive rate times the cohort size. In our results i.e., in wheat $N_f \times F = 11,758.5$ and $R_0 \times N = 11,752.5$. This minor difference is due to rounding-off. This relationship shows the precision obtainable in the age-stage, two-sex life table analysis.

Discussion

consistently Prev quality influenced the development, reproduction and adult longevity of H. variegata. The differences in the life history data of *H. variegata* on four host plant species of S. avenae were significant, indicating the major differences in their suitability. In the this study, H. variegata completed its development from egg to adult stage in 14.89-16.31 days at 25 °C. depending on the host plant of its prey; this range is shorter than those reported elsewhere at the same temperature on three aphids species [15.16-18.85 days in Golizadeh and Jafari-Behi (2012) or 18.1 days in Lanzoni et al. (2004) on M. persicae], but longer than those reported by Wu et al. (2010) on A. gossypii (12.6-14.5 days depending on the host plant of its prey). On the other hand, the pre-adult survival of H. variegata ranged from 86.0 to 94.0%, depending on the host plant of its prey and this was higher than the rates reported by others such as 81.0 % (Golizadeh and Jafari-Behi, 2012), 65.9 % (Farhadi et al., 2011), 44.06-58.97 % (Wu et al., 2010), 49.1% (Lanzoni et al., 2004) and 61.8% (ElHag and Zaitoon, 1996). The differences in reported development time and survival rate between different studies can be attributed to the differences experimental methods, conditions and prey species. In addition, host plants on which the prey fed might affect the nutrition of the prey and in turn affect the predation, development and survival of the predator (Zhang et al., 2010; Wu et al., 2010). Because the variable developmental among individuals rate occurring

incorporated into the age-stage, two-sex life table, the survival curve of a cohort shows stage overlapping (Chi and Yang, 2003). When H. variegata was fed on A. gossypii that had been reared on five host plant species, the suitability of aphids from various host plants differed for this predator (Wu et al., 2010). Similar inferences were made by Francis et al. (2001) for A. bipunctata (L.) on M. persicae. The difference can be attributed to the nature of the host plants used in the experiment. Francis et al. (2001) suggested that host plant allelochemical substances may not only affect the herbivores, but also the pest predators. Lanzoni et al. (2004), and ElHag and Zaitoon (1996) used cylindrical containers and plastic containers, respectively, to rear ladybird beetles. In current study, the male adults lived longer compared to the mean longevity of female adults. Female and male adult longevity of H. variegata was prey dependent and ranged from 39.10 to 53.28 days and 32.66 to 47.88 days depending on four host plants. Mean female adult longevity on corn was 53.28 days, which was longer than that reported by Golizadeh and Jafari-Behi (2012) (64.75-73.10 days) and Farhadi et al. (2011) (44.9 days).

Our results showed that aphid host plant species affected significantly TPOP and oviposition rate of *H. variegata*. Rü and Mitsipa (2000) and Al-Zyoud *et al.* (2005) also obtained similar results. The longevity, and oviposition period and total fecundity of *H. variegata* did vary significantly among four host plants. However, taking into account other reproductive attributes of the predator, our results are different from others. The difference may be attributed to the nature of the host plants used in the experiment.

In this study, total fecundity of *H. variegata* ranged from 362.47 to 587.31 eggs per female, depending on host plant, which was lower than that of Lanzoni *et al.* (2004) (841.7) and (1139.2) on *A. fabae* in Farhadi *et al.* (2011), but higher than that of Kontodimas and Stathas (2005) (76.3–294.4). Also, the recorded total fecundity on Corn, wheat and barley were 587.31, 470.34 and 432.79 eggs, respectively, which were close to the results of Golizadeh and Jafari-Behi (2012).

Table 2 Population parameters of *Hippodamia variegata* fed on aphids reared on four host plants at 25 °C, estimated by using all individuals and the bootstrap techniques.

Parameter	Triticum	Hordeum vulgare	Zea mays	Sorghum durra
	aestivumvar. Tajan	var. Dasht	var. Single cross 704	var. Speed feed
r (day ⁻¹)	0.181 ± 0.006^{a}	0.162 ± 0.006^{c}	0.179 ± 0.006^{b}	0.163 ± 0.006^{c}
$^{\lambda}$ (day ⁻¹)	1.199 ± 007^a	1.176 ± 0.007^{c}	1.197 ± 0.008^{b}	1.177 ± 0.008^{c}
R_{θ} (offspring/individual)	235.05 ± 36.15^{b}	183.97 ± 33.37^{c}	291.14 ± 46.66^{a}	134.30 ± 23.66^{d}
T(day)	29.95 ± 0.41^{c}	32.03 ± 0.44^{a}	31.48 ± 0.40^{b}	29.87 ± 0.33^{c}
GRR (offspring)	297.39 ± 39.53^{b}	215.31 ± 36.05^{c}	334.80 ± 48.62^{a}	173.48 ± 28.92^{d}

Means followed by different letters in a row are significantly different, (SNK; P < 0.05).

R: the intrinsic rate of increase, λ : the finite rate of increase, R_0 : the net reproductive rate, T: the mean generation time, GRR: gross reproductive rate.

In this study, a decrease was shown in survival rate by spending time in both sexes. The curves of m_x and $l_x m_x$ (Fig. 2) showed roughly periodic peaks in reproduction on sorghum that were similar to those exhibited by H. variegata, in study by Farhadi $et\ al$. (2011). The periodic reproductive peaks are apparent in the raw data of the daily fecundity of individual females, in which reproductive peaks are separated by zeros, which represent periods of no reproduction (Yu $et\ al$., 2013).

The population parameters of *H. variegata* are significantly influenced by aphid species. In general, life table can provide a comprehensive description of the development, survival and fecundity of a population. Among the population parameters, the intrinsic rate of increase is the most useful for comparing the population growth potential under specific climatic and food conditions, and it reflects the overall effects of prey on development, reproduction and survival characteristics of a population (Southwood and Henderson, 2000). The values of net reproductive rate (R_0) and intrinsic rate of increase (r) were lower than those reported by Kontodimas and Stathas (2005) ($R_0 = 425.9$, r = 0.178) and Farhadi et al. (2011) $(R_0 = 387.6, r = 0.204)$, but higher than those of Lanzoni et al. (2004) ($R_0 = 52.75$, r =0.114), and ElHag and Zaitoon (1996) ($R_0 = 45.6$, r= 0.082). The R_0 value in Golizadeh and Jafari-Behi (2012) was similar (290.97) to the calculated value in the current study (291.14-183.23). The differences in life table parameter values are possibly a result of different experimental conditions, different prey species, different prey species host plants or geographical populations

(Kontodimas and Stathas 2005; Farhadi *et al.*, 2011; Golizadeh and Jafari-Behi, 2012).

Meyer et al. (1986) discussed the application of the jackknife and bootstrap techniques to population parameters. Based on results for cladoceran species, Meyer et al. (1986) suggested that the precision for reporting r values should be limited in most cases to two significant figures. According to the means and standard errors of our current and previous studies, we suggest that four significant figures are necessary to confirm the values' differences, precision, and variability. Moreover, the frequency distribution of the net reproductive rate that was estimated by employing the bootstrap technique met the assumptions of normality, which is an important premise for further statistical analysis. Because Yu et al. (2013) and Huang and Chi (2012b) zero pseudo values were obtained for R_0 by using the jackknife technique. A R_0 of zero means that the population cannot reproduce offspring and that no intrinsic rate can be estimated. This results in a serious contradiction in the application of the jackknife technique to the estimation of population parameters. Huang and Chi (2012b)mathematically invalidated the use of the jackknife technique in estimating the net reproductive rate. As proven by Chi (1988), based on the two-sex life table, the relationship between R_0 and mean female fecundity (F) is given in equation (10), and our data for N, N_f , F and R_θ (Table 3) were consistent with Eq. (10), for which all individuals are included in the calculation of R_0 . In fact, when the bootstrap technique was employed in the calculation of R_0 , minor differences were apparent due to the application of the resampling technique.

Chi and Yang (2003) also noticed that the application of a jackknife technique resulted in degrees of discrepancy between the estimated means and their definitions. Eq. (10) is valid for the age-stage, two sex life table, as well as the traditional female age-specific life table. It shows a simple and robust relationship between the common statistics of mean fecundity and the net reproductive rate that was defined more than 100 years ago (Boeckh, 1890).

In using a predaceous ladybird beetle for biological control, individuals in the larval stage are more effective control agents than the adults (Trouve et al., 1997), unless the adults are flightless (Kuroda and Miura, 2003; Seko et al., 2008). Inayat et al. (2011) reported that the overall larval feeding rate was twofold greater than that of the adult. In greenhouse conditions, Kuroda and Miura (2003) released H. axyridis egg sheets on cucumber plants, and the emerged larvae effectively decreased the density of A. gossypii. Therefore, the larval stage of *H. variegata* has the potential to be of use in biological control programs. The advantage of a predation study based on the age-stage, two-sex life table is evident in its proper stage grouping compared to the traditional female age-specific life table. To assess the population growth potentials and the efficacies of insect rearing programs, different methods and parameters have been used. However, in comparing the efficiency of a predator, we have to consider not only the predator's population growth rate but also its predation rate. Although the net reproductive rate R_0 was highest for beetles preying aphids fed on corn, the highest intrinsic rate and the finite rate were observed in beetles that preyed on aphids reared on wheat when the effect of time, i.e., the reproductive age, was considered. A faster intrinsic rate of increase or finite rate does not necessarily represent an efficient predator.

Conclusion

In conclusion, this study showed that all of S. avenae from four host plant species were suitable for the growth, development and reproduction of H. variegata. Prey host plants affected significantly the biological and ecological characteristics and performance of *H. variegata*. Wheat was the most suitable host among the tested host plants, followed by corn; sorghum and barley were least suitable as host plants. The obtained life history data in this study are of basic importance for efforts to develop a biological control or integrated pest management approach for the examined host plant species. Our study provides an opportunity for better understanding the tritrophic interactions of the plant-aphid-predator relationship and showed that successful biological control of pests should integrate each trophic level. Not only the food prey but also the host plants of prey should be carefully selected in predator's production for biological control. Moreover, age-stage two sex life table information for *H. variegata* must be integrated with its behavioral characteristics and other information, such as its capacity to establish and disperse. The mechanisms on the influence of prey's host plants on the predator remains to be investigated. Like other studies, we demonstrate that an accurate description of the survival, development, and population parameters can be achieved by the age-stage, two-sex life table. Thus, further investigation is needed on predation capacity and functional response.

Table 3 Relation between F and R_0 of *Hippodamia variegata* fed by aphids reared on four host plants at 25 °C.

Host plants	N	N_{f}	F	R_0	$R_0 = \left(\frac{N_f}{N}\right) F$
Triticum aestivumvar. Tajan	50	25	470.34	235.05	235.05
Hordeum vulgarevar. Dasht	50	22	432.79	183.97	183.97
Zea maysvar. Single cross 704	50	25	587.31	291.14	291.14
Sorghum durravar. Speed feed	50	19	362.47	134.30	134.30

N: the total number of individuals used for life table study; N_f : The number of female adults emerged from N; R_0 : the net reproductive rate; F: the mean fecundity of the female.

Acknowledgment

This research was funded by the Department of Plant Protection, University of Mohaghegh Ardabili, which is greatly appreciated.

References

- Ali, A. and Rizvi, P. Q. 2007. Age specific survival and fecundity table of *Coccinella septempunctata* L. (Coleoptera: Coccinellidae) on different aphid species, Annals of Plant Protection Sciences, 15: 329-334.
- Ali, A. and Rizvi, P. Q. 2009. Life table studies of *Menochilus sexmaculatus* Fabr. (Coleoptera: Coccinellidae) at varying temperature on *Lipaphis erysimi* Kalt. World Applied Sciences Journal, 7: 897-901.
- Al-Zyoud, F., Tort, N. and Sengonca, C. 2005. Influence of host plant species of *Bemisia tabaci* (Genn.) (Hom., Aleyrodidae) on some of the biological and ecological characteristics of the entomophagous *Serangium parcesetosum* Sicard (Col, Coccinellidae). Journal of Pest Science, 78: 25-30.
- Amir-Maafi, M. and Chi, H. 2006. Demography of *Habrobracon hebetor* (Hymenoptera: Braconidae) on two pyralid hosts (Lepidoptera: Pyralidae). Annals of the Entomological Society of America, 99: 84-90.
- An, R. J., Li, X. W., Zhang, J. X. and Zhang, L. J. 2000. Study on bionomics of *Hippodamia variegata* (Goeze). Journal Zhelimu Animal Husbandry College, 10: 14-16.
- Babu, A. 1999. Influence of prey species on feeding preference, postembryonic development and reproduction of *Coccinella transversalis* F. (Coccinellidae: Coleoptera). Entomon, 24: 221-228.
- Bai, Y. Y., Jiang, M. X. and Cheng, J. A. 2005. Effects of transgenic cry1Ab rice pollen on fitness of *Propylea japonica* (Thunberg). Journal of Pest Science, 75: 123-128.
- Bellows, T. S., van Driesche, R. G. and Elkinton, J. S. 1992. Life table construction and analysis in the evaluation of natural enemies. Annual Review of Entomology, 37: 587-614.

- Birch, L. C. 1948. The intrinsic rate of natural increase in an insect population. Journal of Animal Ecology, 17: 15-26.
- Boeckh, R. 1890. Die statistische Messung der ehelichen Fruchtbarkeit. In: Bulletin de l'Institut International de Statistique, vol. 5, first section.
- Buchanan, G. A. 1996. Beneficial insects in the home yard and garden. Georgia Extension Publications and Bulletins, 1140: 1-5.
- Cabral, S., Garcia, P. and Soares, A. O. 2008. Effects of pirimicarb, buprofezin and pymetrozine on survival, development and reproduction of *Coccinella undecimpunctata* (Coleoptera: Coccinellidae). Biocontrol Science and Technology, 18: 307-318.
- Carey, J. R. and Vargas, R. I. 1985. Demographic analysis of insect mass rearing: a case study of three tephritids. Journal of Economic Entomology, 78: 523-527.
- Carey, J. R. 1993. Applied Demography for Biologists with Special Emphasis on Insects. Oxford University Press, New York.
- Chi, H. 2013. TWOSEX-MSChart: a computer program for the age-stage, two-sex life table analysis. Available on: http://quarantine.entomol.nchu.edu.tw/Ecology/Download/TWOSEX MSChart.rar.
- Chi, H. 1988. Life-table analysis incorporating both sexes and variable development rates among individuals. Environmental Entomology, 17: 26-34.
- Chi, H. and Liu, H. 1985. Two new methods for the study of insect population ecology. Bulletin of the Institute of Zoology, Academia Sinica, 24: 225-240.
- Chi, H. and Su, H. Y. 2006. Age-stage, two-sex life tables of *Aphidius gifuensis* (Ashmead) (Hymenoptera: Braconidae) and its host *Myzus persicae* (Sulzer) (Homoptera: Aphididae) with mathematical proof of the relationship between female fecundity and the net reproductive rate. Environmental Entomology, 35: 10-21.
- Chi, H. and Yang, T. C. 2003. Two-sex life table and predation rate of *Propylaea japonica* Thunberg (Coleoptera: Coccinellidae) fed on *Myzus persicae* (Sulzer) (Homoptera:

- Aphididae). Environmental Entomology, 32: 327-333.
- Dillwith, J. W., Neese, P. A. and Brigham, D. L. 1993. Lipid biochemistry in aphids. In: Stanley-Samuelson D. W. and Nelson. D. R (Eds.), Insect Lipids: Chemistry, Biochemistry, and Biology. University of Nebraska Press, Lincoln, pp 389-434.
- Efron, B. and Tibshirani, R. J. 1993. An Introduction to the Bootstrap. Chapman and Hall, New York.
- ElHag, E. A. and Zaitoon, A. A. 1996. Biological parameters for four coccinellid species in Central Saudi Arabia. Biological Control, 7: 316-319.
- Evans, E. W., Richards, D. R. and Kalaskar, A. 2004. Using food for different purposes: female responses to prey in the predator *Coccinella septempunctata* (Coleoptera: Coccinellidae). Ecological Entomology, 29: 27-34.
- Fan, G. H., Liu, B. X., Song, Q. B. and Ma, G. R. 1995. Studies on biology of *Adonia variegata* Goeze. Entomological Journal of East China, 4: 70-74.
- Fan, G. H. and Zhao, J. F. 1988. Functional response of *Adonia variegata* (Goeze) (Coleop: Coccinellidae) to cotton aphids. Natural Enemies Insects, 10: 187-190.
- Farhadi, R., Allahyari, H. and Chi, H. 2011. Life table and predation capacity of *Hippodamia variegata* (Coleoptera: Coccinellidae) feeding on Aphis fabae (Hemiptera: Aphididae). Biological Control, 59: 83-89.
- Febvay, G., Pageaux, J. F. and Bonnot, G. 1992. Lipid composition of the pea aphid, *Acyrthosiphon pisum* (Harris) (Homoptera: Aphididae), reared on host plant and on artificial media. Archives of Insect Biochemistry and Physiology, 21: 103-118.
- Feng, H. Z., Wang, L., Xiong, R. C., Wang, B. and Ji, C. S. 2000. A study on the population dynamics and predacious function of *Hippodamia* (*Adonia*) variegata (Goeze). Entomological Knowledge, 37: 223-227.
- Francis, F., Haubruge, E., Hastir, P. and Gaspar, C. 2001. Effect of aphid host plant on development and reproduction of the third trophic level, the predator *Adalia bipunctata*

- (Coleoptera: Coccinellidae). Environmental Entomology, 30: 947-952.
- Franzmann, B. A. 2002. *Hippodamia variegata* (Goeze) (Coleoptera: Coccinellidae), a predacious ladybird new in Australia. Australian Journal of Entomology, 41: 375-377.
- Giles, K. L., Madden, R. D., Stockland, R., Payton, M. E. and Dillwith, J. W. 2002. Host plants affect predator fitness via the nutritional value of herbivore prey: investigation of a plant-aphid-ladybeetle system. Biocontrol, 47: 1-21.
- Gilkeson, L. and Kelin, M. 2001. Natural enemies of insect pests. Cooperative Extension, Cornell University, Ithea, N. Y. P. 63.
- Golizadeh, A. and Jafari-Behi, V. 2012. Biological traits and life table parameters of variegated lady beetle, *Hippodamia variegata* (Coleoptera: Coccinellidae) on three aphid species. Applied Entomology and Zoology, 47: 199-205.
- Goodman, D. 1982. Optimal life histories, optimal notation, and the value of reproductive value. American Naturalist, 119: 803-823.
- Hodek, I. and Honek, A. 1996. Ecology of Coccinellidae, Boston: Kluwer Academic, 464 pp.
- Huang, Y. B. and Chi, H. 2012a. Age-stage, two-sex life tables of *Bactrocera cucurbitae* (Coquillett) (Diptera: Tephritidae) with a discussion on the problem of applying female age-specific life tables to insect populations. Journal of Insect Science, 19: 263-273.
- Huang, Y. B. and Chi, H. 2012b. Life tables of *Bactrocera cucurbitae* (Diptera: Tephritidae): with an invalidation of the jackknife technique. Journal of Applied Entomology, http://dx.doi.org/10.1111/jen.12002.
- Inayat, T. P., Rana, S. A., Rana, N., Ruby, T., Sadiqui, M. J. I. and Abbas, M. N. 2011. Predation rate in selected coccinellid (Coleoptera) predators on some major aphidid and cicadellid (hemipteran) pests. International Journal of Agriculture and Biology, 13: 427-430.
- Istock, C. A. 1981. Natural selection and life history variation: theory plus lessons from a

- mosquito. In: Denno, R. F. and Dingle, H. (Eds.), Insect Life History Patterns: Habitat and Geographic Variation. Springer-Verlag, New York, pp. 113-127.
- Kalushkov, P. and Hodek, I. 2001. New essential aphid prey for *Anatis ocellata* and *Calvia quatuorde-cimguttata* (Coleoptera: Coccinellidae). Biocontrol Science and Technology, 11: 35-39.
- Kalushkov, P. and Hodek, I. 2004. The effects of thirteen species of aphids on some life history parameters of the ladybird Coccinella septempunctata. Biocontrol, 49: 21-32.
- Kontodimas, D. C. and Stathas, G. J. 2005. Phenology, fecundity and life table parameters of the predator *Hippodamia variegata* reared on *Dysaphis crataegi*. Biocontrol, 50: 223-233.
- Kuroda, T. and Miura, K. 2003. Comparison of the effectiveness of two methods for releasing *Harmonia axyridis* (Pallas) (Coleoptera: Coccinellidae) against *Aphis* gossypii Glover (Homoptera: Aphididae) on cucumbers in a greenhouse. Applied Entomology and Zoology, 38: 271-274.
- Lanzoni, A., Accineli, G., Bazzocchi, G. and Burgio, G. 2004. Biological traits and life table of the exotic Harmonia axyridis compared with *Hippodamia variegata*, and *Adalia bipunctata* (Col, Coccinellidae). Journal of Applied Entomology, 128: 298-306.
- Leslie, P. H. 1945. On the use of matrices in certain population mathematics. Biometrika, 33: 183-212.
- Lewis, E. G. 1942. On the generation and growth of a population. Sankhya, 6: 93-96.
- Lido, P. and Carey, J. R. 1994. Mass rearing of *Anastrepha* (Dip: tephritidae). Fruit flies: a demographic analysisJournal of Economic Entomology, 87: 176-180.
- Lotfalizadeh, H. 2001. Sex determination in some ladybirds (Col.: Coccinellidae) fauna of Moghan region. Journal of Entomological Society of Iran, 21: 69-88 (in Persian with English abstract).
- Lundgren, J. G., Hesler, L. S., Tilmon, K., Dashiell, K. and Scott, R. 2009. Direct effects of soybean varietal selection and Aphis glycines-resistant

- soybeans on natural enemies. Arthropod-Plant Interactions, 3: 9-16.
- Meyer, J. S., Ingersoll, C. G., McDonald, L. L. and Boyce, M. S. 1986. Estimating uncertainty in population growth rates: jackknife vs. bootstrap techniques. Ecology, 67: 1156-1166.
- Michels, G. J. and Bateman, A. C. 1986. Larval biology of two imported predators of the greenbug, *Hippodamia variegata* Goeze and *Adalia flavomaculata* Degeer, under constant temperatures. Southwestern Entomologist, 11: 23-30.
- Michels, G. J. and Flanders, R. V. 1992. Larval development, aphid consumption and oviposition for five imported coccinelids at constant temperature on Russian wheat aphids and greenbugs. Southwestern Entomologist, 17: 233-243.
- Moser, S. E., Harwood, J. D. and Obrycki, J. J. 2008, Larval feeding on Bt hybrid and non-Bt corn seedlings by *Harmonia axyridis* (Coleoptera: Coccinellidae) and *Coleomegilla maculata* (Coleoptera: Coccinellidae). Environmental Entomology, 37: 525-533.
- Omkar, S. and Bind, R. B. 2004. Prey quality growth, dependent development reproduction of biocontrol a agent, Cheilomenes sexmaculata (Fabrcius) (Coleoptera: Coccinellidae). Biocontrol Science and Technology, 14: 665-673.
- Omkar, S. and Srivastava, S. 2003. Influence of six aphid prey species on development and reproduction of a ladybird beetle, *Coccinella septempunctata*. Biocontrol, 48: 379-393.
- Omkar, S. and James, B, E. 2004. Influence of prey species on immature survival, development, predation and reproduction of *Coccinella transversalis* Fabricius (Col, Coccinellidae). Journal of Applied Entomology, 128: 150-157.
- Pang, B. P., Zhao, J. X., Meng, R. X., Meng, H. W. and Yi, W. D. 2000 Predation of *Adonia variegata* (Goeze) on the wheat aphid, *Macrosiphum avenae* (F.): intraspecific interference and space heterogeneity. Journal of Inner Mongolia Agricultural University, 21: 13-17.

- Pervez, A. and Omkar, S. 2004. Prey-dependent life attributes of an aphidophagous ladybird beetle, *Propylea dissecta* (Coleoptera: Chrysomelidae). Biocontrol Science and Technology, 14: 385-396.
- Price, P. W., Bouton, C. E., Gross, P., McPheron, B. A., Thompson, J. N. and Weis, A. E. 1980. Interactions among three trophic levels: influence of plant on interactions between insect herbivores and natural enemies. Annual Review of Ecology, Evolution, and Systematics, 11: 41-65.
- Rü, B. L. and Mitsipa, A. 2000. Influence of the host plant of the cassava mealy bug *Phenacoccus manihoti* on life-history parameters of the predator *Exochomus flaviventris*. Entomologia Experimentalis et Applicata, 95: 209-212.
- Seko, T., Yamashita, K. and Miura, K., 2008. Residence period of a flightless strain of the ladybird beetle *Harmonia axyridis* Pallas (Coleoptera: Coccinellidae) in open fields. Biological Control, 47: 194-198.
- Southwood, T. R. E. and Henderson, P. A. 2000. Ecological methods. Blackwell, Oxford.
- SPSS Inc .2007. SPSS base 16.0 user's guide. SPSS Incorporation, Chicago.
- Takizawa, T., Yasuda, H. and Agarwala, B. K. 2000. Effects of parasitized aphids (Homoptera: Aphididae) as food on larval performance of three predatory ladybirds (Coleoptera: Coccinellidae). Applied Entomology and Zoology, 35: 467-472.
- Thompson, S. N. 1999. Nutrition and culture of entomophagous insects. Annual Review of Entomology, 44: 561-592.
- Trouve, C., Ledee, S., Ferran, A. and Brun, J. 1997. Biological control of the damson-hop aphid, *Phorodon humuli* (Hom.: Aphididae), using the ladybeetle *Harmonia axyridis*. Entomophaga, 42: 57-62.
- Tsaganou, F. C., Hodgson, C. J., Athanassiou, C.

- G., Kavallieratos, N. G. and Tomanovic, Z. 2004. Effect of *Aphis gossypii* Glover, *Brevicoryne brassicae* (L.), and *Megoura viciae* Buckton (Hemiptera: Aphidoidea) on the development of the predator *Harmonia axyridis* (Pallas) (Coleoptera: Coccinellidae). Biological Control, 31: 138-144.
- Wang, Y. H., Liu, B. S., Fu, Z. H. and Gu, L. N. 1984. Studies on the life history and occurrence of *Adonia variegata* (Goeze). Entomological Knowledge, 21: 19-22.
- William, F. L. 2002. Lady beetles. Ohio State
 University Extension Fact Sheet,
 Horticulture and Crop Science. Division of
 Wildlife, 2021 Coffey Rd. Columbus, Ohio.
- Wu, X. H., Zhou, X. R. and Pang, B. P. 2010. Influence of five host plants of *Aphis gossypii* Glover on some population parameters of *Hippodamia variegata* (Goeze). Journal of Pest Science, 83: 77-83.
- Yu, J. Z., Chi, H. and Chen, B. H. 2005. Life table and predation of *Lemnia biplagiata* (Coleoptera: Coccinellidae) fed on *Aphis gossypii* (Homoptera: Aphididae) with a proof on relationship among gross reproduction rate, net reproduction rate, and preadult survivorship. Annals of the Entomological Society of America, 98: 475-482.
- Yu, J. Z., Chi, H. and Chen, B. H. 2013. Comparison of the life tables and predation rates of *Harmonia dimidiata* (F.) (Coleoptera: Coccinellidae) fed on *Aphis gossypii* Glover (Hemiptera: Aphididae) at different temperatures. Biological Control, 64: 1-9.
- Zhang, S. Z., Jiang, R. X., Zhang, F., Xu, X. L. and Liu, T. X. 2010. Suitability of *Aphis craccivora* (Hemiptera:Aphididae) and *Bemisia tabaci* (Hemiptera: Aleyrodidae) biotype-B as prey for *Chrysopa pallens* (Neuroptera: Chrysopidae). Biocontrol Science and Technology, 20: 961-971.

ویژگی های دموگرافیک کفشدوزک (Goeze) طای دموگرافیک

(Coleoptera: Coccinellidae) با تغذیه از شته سبز گندم

Sitobion avenae Fabricius (Hemiptera: Aphididae)

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چکیده: کفشدوز کها یکی از مهم ترین گونه های شکار گردر میان بندپایان می باشند. بنابراین مظالعه پارامترهای رشد جمعیت آنها یک تصویر شفافی درباره طول عمر آنها به ما می دهد. تاریخچه زندگی کفشدوز ک Sitobion avenae با تغذیه از شته سبز گندم Sitobion avenae روی گیاهان میزبان مختلف در شرایط آزمایشگاهی در دمای 1 ± 5 درجه سلسیوس، رطوبت نسبی 1 ± 5 درصد و مختلف در شرایط آزمایشگاهی در دمای 1 ± 5 درجه سلسیوس، رطوبت نسبی 1 ± 5 درصد و دوره نوری 1 + 5 ساعت روشنایی و 1 + 5 ساعت تاریکی مورد بررسی قرار گرفت. اطلاعات دوره نشو و نماه بقاء و باروری با استفاده از جدول زندگی دوجنسی مورد تجزیه و تحلیل قرار گرفتند. بر طبق جدول زندگی دوجنسی نرخ خالص تولیدمثل 1 + 5 کفشدوز ک با تغذیه از گندم (رقم تجن)، جو (دشت)، ذرت رسینگل کراس 1 + 5 و سور گوم (اسپید فید) به ترتیب 1 + 5 به به عنوان یک شاخص ماده اماده انسل به دست آمد. هم چنین براساس نرخ ذاتی افزایش جمعیت 1 + 5 به به عنوان یک شاخص مناسب در افزایش جمعیت این گفشدوز ک می باشد، گندم نسبت به سایر گیاهان میزبان دارای بیش ترین نرخ ذاتی افزایش جمعیت بود و با تغذیه از گیاهان میزبان ذکر شده در بالا به ترتیب 1 + 5 به برای در ک به ترمی توان نشو و نما و بقای شکار گر را با دقت توصیف کرد. هم چنین این مطالعه برای در ک به ترمی تومکنش گیاه – شته – شکار گر فرصت خوبی را ارائه کرد، هم چنین این مطالعه برای در ک به ترمی تومکنش گیاه – شته – شکار گر فرصت خوبی را ارائه کرده است.

واژگان کلیدی: دو جنسی، دموگرافیک، Sitobion avenae Hippodamia variegata گیاهان میزبان