

Short paper

First report of *Rhopalosiphum nymphaeae* (L.) (Hem.: Aphididae) on *Azolla filiculoides* from Iran and its male formation on secondary host plant

Atousa Farahpour-Haghani^{1*}, Mahdi Jalaeian¹ and Mohsen Mehrparvar²

- 1. Rice Research Institute of Iran (RRII), Agricultural Research, Education and Extension Organization (AREEO), Rasht, Iran.
- 2. Department of Biodiversity, Institute of Science and High Technology and Environmental Sciences, Graduate University of Advanced Technology, Kerman, Iran.

Abstract: To determine the effective bio-control agents of water fern, *Azolla filiculoides*, adults and nymphs of water lily aphid, *Rhopalosiphum nymphaeae* (L.), were collected on *A. filiculoides* in Guilan Province, Iran, during October 2013. This aphid has been collected extensively from almost all water fern samples during 2013 and 2014. Although *R. nymphaeae* was reported from numerous host plants in Iran, this species was collected for the first time from water fern. This aphid is heteroecious holocyclic with a sexual phase on *Prunus* spp., but it is the first report of its sexual phase formation on water fern as secondary host plant.

Keywords: Aphid, Rice, Water fern, Guilan

Introduction

Water lily aphid, *Rhopalosiphum nymphaeae* (Linnaeus, 1761) (Hem.: Aphididae), as a polyphagous species is a heteroecious holocyclic aphid, living on a large variety of water plants. This aphid is able to survive underwater (Holman, 2009; Blackman and Eastop, 2015). In large colonies that develop on water lilies, the aphids aggregate along the leaf veins and infest flowers as well. The sexual forms have been reported on *Prunus* spp. (Blackman and Eastop, 1994; 2006). The rates of development, natality and survivorship of *R. nymphaeae* have been studied in relation to its potential for virus transmission on both crops and aquatic weeds (Ballou *et al.*, 1986).

Handling Editor: Ehsan Rakhshani

* Corresponding author, e-mail: hpapiliona@gmail.com Received: 24 May 2015, Accepted: 31 July 2015 Published online: 28 September 2015 Recorded optimal temperature range for *R. nymphaeae*, is 21 to 27 °C (Hance *et al.*, 1994).

Spring colonies of the aphid infest young twigs, leaf petioles and fruit stalks of various *Prunus* spp. which cause curling of host plant leaves (Blackman and Eastop, 1994; 2006). This aphid species is tended by ants. Spring migrant alatae fly to secondary hosts in May-June. Rhopalosiphum nympheae is almost cosmopolitan and its activity on Azolla spp. was reported frequently (Lumpkin and Plunkett, 1980; Rostron, 1983; Center et al., 2002). It has been recommended as a biological control agent for water weeds in rice as well (Yano et al., 1983; Calilung and Lit, 1986; Lu et al., 1991; Oraze and Grigarick, 1992; Hance et al., 1994; Center et al., 2002). Beside these, R. nymphaeae is vector for some viruses including abaca mosaic, cabbage black ringspot, cauliflower mosaic, cucumber mosaic, and onion yellow dwarf viruses (Kennedy et al., 1962: Chan et al., 1991). It caused die-back of water lettuce in Nigeria (Pettet and Pettet, 1970). Moreover, the aphid transmitted eggplant mosaic virus in India (Seth and Raychaudhuri, 1973).

Rhopalosiphum nymphaeae has already been reported in Iran on Prunus domestica and Prunus divaricata (Hodjat, 1993) and Mokhtari et al. (2012) reported this aphid on Plantago lanceolata from central Alborz. This Aphid has been reported as a host for parasitoids Aphidius matricariae Haliday, Aphidius transcaspicus Telenga, Ephedrus cerasicola Starý, Lysiphlebus fabarum Marshall and Praon necans Mackauer in Iran (Mokhtari et al., 2000; Nazari et al., 2012; Barahoei et al., 2014).

Results

We collected *R. nymphaeae* (Fig. 1) on water fern, *Azolla filiculoides* Lam. (Pteridophyta: Salviniaceae), for the first time in October 2013 from natural habitats. Alate males (Fig. 2) were found among the samples collected during the autumn and winter 2014. This is the first time to report the sexual form of this aphid on the secondary host plant. Decreasing of day length and temperature are the most important factors that trigger sexual morph production in aphids (Kawada, 1987). Biometric data of parthenogenetic and sexual morphs collected on water fern in Iran are presented in Table 1.

Materials examined: In the current study, the specimens were collected for the first time on A. filiculoides in Iran from Guilan Province, in the Rice Research Institute of Iran (RRII) (N 37°12′22.2″, E 049°38′40.7″, 80 m). Aphids were collected weekly from October 2014 until February 2015. Each time at least 30 individuals were collected and preserved in 70% ethanol. Slide mounted specimens are deposited in Aphid Collection of Aphidology Research Group, Institute of Science and High Technology and Environmental Sciences, Graduate University of Advanced Technology, Kerman, Iran and preserved samples in alcohol are deposited in the Plant Protection Research Department, Rice Research Institute of Iran (RRII), Rasht, Iran.

Oraze and Grigarick (1992) recommended this aphid as a bio-control agent for duck salad

(Heteranthera limosa) in rice fields. However, we could not find any report concerning successful biological control of a weed by this aphid. It seems that despite of wide distribution, having a board host range makes it less favorable as a bio-control agent for weeds due to probable damage on non-target plants. Nevertheless, at areas that use of water fern as green fertilizer in rice fields this aphid known as a considerable pest for water fern (Lumpkin and Plunkett, 1980).



Figure 1 Rhopalosiphum nymphaeae on Water fern.



Figure 2 Alate male of *Rhopalosiphum nymphaeae* on Water fern.

Table 1 Biometric data of *Rhopalosiphum nymphaeae* collected on *Azolla filiculoides* in Iran.

| Morph | Character length | Biometric data | Ratio | Biometric data |
|-----------------------------|------------------|-------------------|----------------------------|----------------|
| Apterous viviparae (n = 24) | Body | 1.36-1.59 | PT/ANTVIb | 3.50-4.00 |
| | PT | 0.33-0.35 | URS/2HT | 1.36-1.45 |
| | ANTVIb | 0.08-0.10 | URS/Cauda | 1.00-1.33 |
| | URS | 0.14-0.15 | SIPH/Body length | 0.19-0.21 |
| | 2HT | 0.11-0.12 | SIPH/Cauda | 2.07-2.50 |
| | SIPH | 0.27-0.31 | Rhin. on ANTIII | 0 |
| | Cauda | 0.11-0.14 | | |
| Alate viviparae (n = 6) | Body | 1.73-1.76 | URS/2HT | 1.25-1.36 |
| | PT | 0.26-0.27 | URS/Cauda | 1.15 |
| | ANTVIb | 0.12-0.13 | SIPH/Body length | 0.14-0.15 |
| | URS | 0.14 | SIPH/Cauda | 1.92 |
| | 2HT | 0.11-0.12 | Rhin. on ANTIII | 12-16 |
| | SIPH | 0.24-0.26 | Rhin. on ANTIV | 2-6 |
| Alate males (n = 18) | Cauda Body | 0.12 1.48-1.61 | Rhin. on ANTV PT/ANTVIb | 0-1 3.76 |
| | PT | 0.26-0.45 | URS/2HT | 1.30-1.50 |
| | ANTVIb | 0.10-0.12 | SIPH/Body length | 0.13-0.16 |
| | URS | 0.12-0.14 | SIPH/Cauda | 2.22-2.78 |
| | 2HT | 0.10-0.11 | URS /Cauda | 1.56-1.67 |
| | SIPH | 0.18-0.24 | Rhin. on ANTIII | 28-40 |
| | Cauda | 0.08-0.09 | Rhin. on ANTIV | 12-22 |
| | | | Rhin. on ANTV | 7-10 |

Abbreviations: ANTIII, ANTIV, ANTV, ANTVIb, antennal segments III, IV, V, and the base of antennal segment VI, respectively; PT, processus terminalis; Rhin, Rhinaria; URS, ultimate rostral segment; 2HT, second segment of hind tarsus; and SIPH, siphunculus. Lengths are given in mm.

Acknowledgements

We would like to thank the Head of Plant Protection Research Department, Rice Research Institute of Iran for providing financial support. This research was supported by grant of the Institute of Science and High Technology and Environmental Sciences, Kerman, Iran.

References

Ballou, J. K., Tsai, J. H. and Center, T. D. 1986. Effects of temperature on the development natality, and longevity of *Rhopalosiphum* *nymphaeae* (L.) (Homoptera: Aphididae). Environmental Entomology, 15: 1096-1099.

Barahoei, H., Rakhshani, E., Nader, E., Starý, P., Kavallieratos, N. G., Tomanović, Z. and Mehrparvar, M. 2014. Checklist of Aphidiinae parasitoids (Hymenoptera: Braconidae) and their host aphid associations in Iran. Journal of Crop Protection. 3 (2): 199-232.

Blackman, R. L. and Eastop, V. F. 1994. Aphids on the World's Trees. CAB International with The Natural History Museum, London. Viii + 987 pages, 135 figures, 16 plates.

Blackman, R. L. and Eastop, V. F. 2006. Aphids on the World's Herbaceous Plants

- and Shrubs. Volume 2 The Aphids. John Wiley and Sons with the Natural History Museum, London. viii + pages 1025-1439.
- Blackman, R. L. and Eastop, V. F. 2015. Aphids on the World's Plants: An online identification and information guide. Available from: http://www.aphidsonworldsplants.info/ Access Date: 2015.04.19.
- Calilung, V. J. and Lit, I. L. Jr. 1986. Studies on the insect fauna and other invertebrates associated with *Azolla* spp. Philippine Agriculturist, 69 (4): 513-520.
- Center, T. D., Dray, Jr. F. A., Jubinsky, G. P. and Grodowitz, M. J. 2002. Insects and other arthropods that feed on aquatic and wetland plants. U. S. Department of agriculture, Agricultural research service, Technical Bulletin No. 1870. pp 209.
- Chan, C. K., Forbes, A. R. and Raworth, D. A. 1991. Aphid-transmitted viruses and their vectors of the world. Agriculture Canada Technical Bulletin 1991-3E. 1-216 pp.
- Hance, T., Nibelle, D. and Lebrun, P. H. 1994. Selection of *Azolla* forms resistant to the water lily aphid, *Rhopalosiphum nymphaeae*: Life history of *Rhopalosiphum nymphaeae*. Entomologia Experimentalis et Applicata, 70: 11-17.
- Hodjat, S. H., 1993. A list of aphids and their host plants in Iran. Shahid Chamran University, 148 pp.
- Holman, J. 2009. Host Plant Catalog of Aphids, Palaearctic Region. Springer Science and Business Media B. V. 1216 pp.
- Kawada, K. 1987. Polymorphism and morph determination. In: Minks, A. K. and Harrewijn, P. (Eds.), Aphids, their biology, natural enemies and control (pp. 255-266). Amsterdam: Elsevier.
- Kennedy, J. S., Day, M. F. and Eastop, V. F. 1962. A conspectus of aphids as vectors of plant viruses. Commonwealth Institute of Entomology, London.
- Lu, Z., Zhu, J., Wen, M. and Wang, D. 1991. Studies on the biology and population

- dynamics of water lily aphid. Acta Phytophylacica Sinica, 18 (4): 357-361.
- Lumpkin, T. A. and Plunkett, D. L. 1980. *Azolla*-Botany, physiology, and uses-Green manure. Economic Botany, 34: 111-153.
- Mokhtari, A., Nozari, J., Rezwani, A., Rasolian, G., Petrović-Obradović, O. and Rakhshani, E. 2012. Aphids (Hemiptera: Aphididae) associated with grasslands of central Alborz, Iran. Acta Entomologica Serbica, 17 (1/2): 1-22
- Mokhtari, A., Sahragard, A., Rezwani, A. and Salehi, L. 2000. Study on five parasitoid species of the stone fruit aphids in Gilan Province. Proceedings of the 14th Iranian Plant Protection Congress, Isfahan University of Technology, P. 273.
- Nazari, Y., Zamani, A. A., Masoumi, S. M., Rakhshani, E., Petrović-Obradović, O., Tomanović, S., Starý, P. and Tomanović, Ž. 2012. Diversity and host associations of aphid parasitoids (Hymenoptera: Braconidae: Aphidiinae) in the farmlands of western Iran. Acta Entomologica Musei Nationalis Pragae, 52: 559-584.
- Oraze, M. J. and Grigarick, A. A. 1992. Biological Control of Ducksalad (*Heteranthera limosa*) by the Water lily Aphid (*Rhopalosiphum nymphaeae*) in Rice (*Oryza sativa*). Weed Science, 40: (2) 333-336.
- Pettet, A. and Pettet, S. J. 1970. Biological control of *Pistia stratiotes* in Western State, Nigeria. Nature, 226: 282 p.
- Rostron, J. C. 1983. The distribution of *A. filiculoides* in Britain. Department of Human Sciences, Loughborough University.
- Seth, M. L. and Raychaudhuri, S. P. 1973. Further studies on a new mosaic diseases of Brinjal (*Solanum melongena* L.). Proc. Indian Natl. Acad Sci. B 39: 122-128.
- Yano, K., Miyake, T. and Eastop, V. F. 1983. The biology and economic importance of Rice aphids (Hemiptera: Aphididae): a review. Bulletin of Entomological Research, 73 (4): 539-566.

اولین گزارش شته (Hem.: Aphididae) (L.) (Hem.: Aphididae) روی آزولا در ایران و ایران و ایران و ایران و ایران دوم

آتوسا فرح پور حقانی * ، مهدی جلائیان 1 و محسن مهر پرور 7

۱- مؤسسه تحقیقات برنج کشور، سازمان تحقیقات ترویج و آموزش کشاورزی، رشت، ایران.

۲- گروه تنوع زیستی، پژوهشگاه علوم و تکنولوژی پیشرفته و علوم محیطی، دانشگاه تحصیلات تکمیلی صنعتی و فناوری پیشرفته، کرمان، ایران.

* پست الکترونیکی نویسنده مسئول مکاتبه: hpapiliona@gmail.com

دریافت: ۳ خرداد ۱۳۹۴؛ پذیرش: ۹ مرداد ۱۳۹۴

چکیده: به منظور یافتن عوامل کنترل بیولوژیک مؤثر از حشرات فعال روی گیاه آزولا Azolla ازولا ۱۳۹۱ در پاییز ۱۳۹۱ نمونه برداری هایی صورت گرفت. پورهها و حشرات کامل شتهی filiculoides در پاییز ۱۳۹۱ نمونه برداری هایی صورت گرفت. پورهها و حشرات کامل شته Rhopalosiphum nymphaeae (L.) به طور گسترده ای از روی کلیه ی نمونه های آزولا در سال های ۱۳۹۲ و ۱۳۹۳ جمع آوری شده این اولین این که تاکنون این شته از روی بسیاری از گیاهان میزبان در ایران جمع آوری شده است، اما این اولین گزارش از این شته روی گیاه آزولا می باشد. این شته، دارای تناوب میزبانی و با چرخه ی زندگی کامل بوده و مرحله ی جنسی آن روی گونه های مختلف گیاه . Prunus spp ایجاد می شود اما این اولین گزارش تشکیل مرحله ی جنسی آن روی آزولا به عنوان میزبان دوم است.

واژگان کلیدی: شته، برنج، آزولا،گیلان