

# Identification of new maize inbred lines with resistance to *Striga hermonthica* (Del.) Benth.

Haron Karaya<sup>1</sup>, Njoroge Kiarie<sup>2</sup>, Stephen Mugo<sup>1</sup>, Fred Kanampiu<sup>1\*</sup>, Emmanuel Ariga<sup>2</sup> and John Nderitu<sup>2</sup>

- 1. International Maize and Wheat Improvement Center (CIMMYT), P. O. Box 1041-00621, Nairobi, Kenya.
- 2. University of Nairobi, Faculty of Agriculture, Upper Kabete Campus, P. O. Box 29053, Nairobi, Kenya.

**Abstract:** Striga hermonthica is a noxious, obligate hemi-parasite of cereal grasses that causes severe grain yield loss in susceptible maize cultivars in Africa. The development of host plant resistance is one of the most practical Stiga control strategies. In this study experiments on 36 maize inbred lines were conducted in pots and in field during the two rainy seasons of 2009 at Kibos and Alupe stations in Kenya. This study was carried out in order to determine the variation in Striga emergence, and the correlation between the attachments of the parasite to the roots. Significant differences (P < 0.001) were detected among the inbred lines for grain yield under Striga-free environment. The Striga damage rating (SDR) was significant (P < 0.05) among the inbred lines. A highly significant and negative correlation coefficient was observed between grain yield and Striga damage rating (r = -0.67). Positive correlation coefficients were observed between grain yield and ear aspect (r = 0.46) and plant aspect (r = 0.46)0.75), respectively. For the experiment in pots, highly significant differences (P < 0.01) were observed among the inbred lines for Striga resistance traits. Striga attachments were found to be correlated with the number of emerged Striga plants. A significant correlation was found between Striga attachments and Striga counts in pots at the 10th week after planting (WAP) (r = 0.25) and the  $14^{th}$  WAP (r = 0.31). Inbred lines JI-30-19 and OSU231//56/44-6-4-17-3 were identified as the most resistant lines as they consistently performed well in both Striga-free and Striga-infested environments. These inbred lines could be used for breeding Strigaresistant maize varieties.

**Keywords:** maize inbred lines, resistance, *Striga hermonthica*, Kenya

### Introduction

Maize (Zea mays) is one of the major staple food crops in sub-Saharan Africa. The demand for this cereal in the world is

Handling Editor: Dr. Majid Agha Alikhani

\*Corresponding author, e-mail: f.kanampiu@cgiar.org Received: 18 February 2012, Accepted: 20 June 2012 expected to increase to about 504 million tons by 2020, thus surpassing the demand for both wheat and rice (IFPRI, 2000). Among the most serious biotic constraints to maize production in the land holdings of resource-poor farmers is the root hemiparasitic weed *Striga hermonthica*. The parasite decimates maize, pearl millet, sorghum and upland rice in Africa wherever it exists. *Striga* is an obligate parasite which has a deleterious effect on its host as well as robbing it of

water and nutrients (Amusan *et al.*, 2008). This root-attaching parasite affects over 100 million people (Kanampiu and Friesen, 2003; Berner *et al.*, 1995)

Grain yield losses in maize from S. hermonthica infestation in Africa range from 20 to 80 % (Berner et al., 1995), but can sometimes reach 100 % in susceptible maize cultivars under severe field infestation al..1990: (Ransom etHaussmann et al., 2000). The development of host plant resistance and tolerance is one of the most feasible and effective Striga control strategy, and is a potentially practical option for reducing yield loss from S. hermonthica for farmers who lack the financial means to use high-input management practices and other options to control Striga in maize fields (Doggett, 1988; Ramaiah et al., 1991).

The International Institute for Tropical Agriculture (IITA) has developed artificial field infestation techniques that impart uniform infestation with the parasite and accurately identify cultivars resistant to S.hermonthica from diverse germplasms (Kim, 1991). The IITA has also developed many maize inbred lines, hybrids and populations with improved field tolerance and resistance to Striga (Kim, 1994; Badu-Apraku et al., 2007). Tolerant germplasm supports a number of emerged Striga plants which may ultimately flower and set seeds, resulting in an increase in the Striga seed bank in the soil. This therefore calls for further screening towards high Striga resistance levels, as Striga-resistant varieties reduce the seed reproduction of the parasites and contribute to the depletion of the soil seed bank (Haussmann, 2000). To obtain resistant germplasm, a good source of resistance was obtained from elite tropical germplasms as well as from populations obtained from local maize collections in Africa and an accession of Zea diploperennis as donor parents (Berner et al., 1995). Subsequently resistant inbred lines with high resistance levels were developed through

intensive screening of the germplasm in the field.

An ideal maize inbred line with the desired levels of resistance under field conditions should allow the emergence of only a few parasitic plants and show very low parasitism and little loss in grain yield (Kim, 1991; Kim, 1994). Such an inbred line would probably have low levels of Striga emergence stimulants, resulting in a low emergence. It is therefore of paramount importance to understand the relationship between the number of emerged Striga plants in the field, and the attachment of the germinated Striga seeds to host roots. The aim of this study was therefore to identify new maize inbred lines with good levels of Striga resistance by screening maize inbred lines from diverse sources (IITA and Kenyan germplasm) under artificial infestation in pots and field trials. The study sought firstly to confirm the efficacy of the IITA-sources of resistance under conditions in eastern Africa, and secondly to explore the possible presence of field resistance in germplasm obtained from Kenyan sources.

### **Materials and Methods**

### **Field experiment**

A total of 36 maize inbred lines from various sources, which included the Kenya Agricultural Research Institute (KARI), International Wheat Maize and **Improvement** Center (CIMMYT) International Institute for **Tropical** Agriculture (IITA) (Table 1), were used in this study. The inbred lines were first planted at Kiboko site (37<sup>0</sup>75'E, 2<sup>0</sup> 15'S) in a seed increase nursery for adaptation during short rainy season of 2008. The inbred lines were evaluated on-station at Kibos (0°4'S, 34°48'E) and Alupe (0°29'N, 34° 2'E) under both artificial Striga infestation and in Striga-free environments during the long rainy season and the short rainy season of 2009. Artificial infestation was conducted in a specially developed field facility in order to screen large numbers of breeding lines. Plants were artificially infested with *S. hermonthica* seeds. *Striga* seeds were added to each plot to ensure that each maize plant was exposed to a minimum of 2,000 viable *Striga* seeds.

These seeds were added in a sand/seed mixture and placed in an enlarged planting hole at a depth of 7–10 cm directly below the maize seed.

Table 1 Maize inbred lines tested under both Striga-free and Striga-infested conditions.

Entry	Genotype	Source	Entry	Genotype	Source
1	OSU231//56/44-6-4-17-3	KARI	19	JI-30-17	KARI (MUGUGA)
2	TESTR 152	IITA	20	TESTR 139	IITA
3	Л-30-19	KARI (MUGUGA)	21	CML444-IR	CIMMYT
4	JI-30-1-19	KARI (MUGUGA)	22	DT//56/4-6-1-15-2	KARI (MUGUGA)
5	F1-14-14-24-4-5-4	KARI (MUGUGA)	23	CML395	CIMMYT
6	CML444	CIMMYT	24	JI-30-21	KARI (MUGUGA)
7	F1-14-79-4-1-3	KARI (MUGUGA)	25	JI-30-7	KARI (MUGUGA)
8	TESTR 153	IITA	26	JI-30-8	KARI (MUGUGA)
9	JI-304	KARI (MUGUGA)	27	TESTR 149	IITA
10	Л-30-18	KARI (MUGUGA)	28	TESTR 132	IITA
11	Л-303	KARI (MUGUGA)	29	CML202IR	CIMMYT
12	TESTR 156	IITA	30	MGA19-4-1	KARI (MUGUGA)
13	CML204-IR	CIMMYT	31	TESTR 136	IITA
14	EARLY-N-POP-7-13-5-1	KARI (MUGUGA)	32	TESTR 151	IITA
15	JI-30-22	KARI (MUGUGA)	33	E11-133/7/44-6-3-17-3-2	KARI (MUGUGA)
16	TESTR 150	IITA	34	TESTR 133	IITA
17	JI-30-16	KARI (MUGUGA)	35	CML206//56/44-6-3-7-1	KARI (MUGUGA)
18	JI-30-7	KARI (MUGUGA)	36	CML395-IR	CIMMYT

The genotypes were planted in 5m single row plots, with a spacing of 75cm between rows and 25 cm between hills with two seeds per hill, and later thinned to one plant per hill to give a population of approximately 53,333 plants per hectare. The experimental design used was an alpha lattice (0, 1) design with 2 replicates. Di-ammonium phosphate (18-46-0) was applied during planting at 50 and 128 kg N and P<sub>2</sub>O5/ha, and top dressing was done using calcium ammonium nitrate (CAN) at 50 kg N/ha. Normal crop husbandry was carried out; weeding was done three weeks after planting and thereafter hand pulling was done only to remove other types of weed other than Striga.

Data were recorded from each plot on agronomic traits which included: grain yield, days to 50 % anthesis, days to 50 % silking, anthesis silking interval (calculated as the difference between days to 50 % anthesis and days to 50 % silking), plant height and ear height. Reaction to two major diseases gray leaf spot caused by Cercospora zeae-maydis and Northern leaf blight caused by Exserohilum turcicum was recorded using a scale of 1-5, where 1 = nodisease and 5 = severely diseased. The Striga damage rating was recorded using a scale of 1 - 9 (where 1 - 3 = no damage, 4 - 3 = no damage) 6 = extensive leaf blotching, wilting and some stunting, and 7 - 9 = completescorching). The Striga count data was recorded by counting the number of Striga plants emerged per plot starting at 8 weeks and then after every two weeks up to 14 weeks after planting (WAP).

### Pot experiment

The 36 maize inbred lines were planted in pots 20cm in diameter and 30cm in height. The pots were filled with sandy soils up to 25cm from the bottom. The *Striga* inoculum was applied in each pot using a tablespoonful to ensure about 2000 viable *Striga* seeds per pot. An enlarged hole was made in the sand in

each pot and the maize seeds were placed directly on top of the inoculum. Four maize seeds were sown in each pot and later thinned to two to ensure a uniform stand. The data recorded included *Striga* counts at 10, 12 and 14 WAP, flowering *Striga* plants at 12, 14 and 15 WAP and *Striga* plants setting seeds at 12, 14 and 15 WAP. *Striga* attachment was recorded after washing the maize roots of each plant and later counting individual attachments.

### **Statistical analysis**

Striga count per square meter was calculated and the data transformed using  $log_{10}(X+1)$ , where X= count per meter squared. An adjustment of grain yield to 15 % moisture content was done after harvesting. The data were then subjected to analysis of variance (ANOVA) using the General Linear Model (GLM) procedure of SAS (SAS, 2003) for individual locations and across locations. Means were separated using Duncan's Multiple Range Test at p < 0.05. Least significant differences (LSD 0.05) values based on analysis of variance were also calculated to allow pair-wise multiple comparisons among means.

### **Results**

## Field experiments *Striga*-free environment

Grain yield was highly significant (P < 0.001) among the inbred lines evaluated. The mean grain yield was 1.4 t/ha and the range 4.3 t/ha. Inbred was 0.1to OSU231//56/44-6-4-17-3 gave the highest grain yield while CML395-IR gave the lowest yield. Among the top 10 inbred lines in terms of grain yield, seven were from KARI, one was from CIMMYT and two were from IITA (Table 2). Highly significant (P < 0.01) differences were observed in days to 50 % anthesis, days to 50 % silking, plant aspect, plant height and ear height.

**Table 2** Performance of selected maize inbred lines under *Striga*-free conditions across sites.

Genotypes	Grain yield (t/ha)	Days to 50 % anthesis (days)	Days to 50 % silking (days)	Plant height (cm)	Ear height (cm)	E. turcicum (score 1-5)	Plant aspect (score 1-5)
OSU231//56/44-6-4-17-3	4.3	64.3	70	200	102.5	2.4	3.5
TESTR 152	4.1	70.0	76	145	80.0	2.5	3.8
JI-30-19	4.0	65.8	74	184	101.3	2.4	4.0
JI-30-20	3.9	68.5	77	199	102.5	2.5	3.5
F1-14-14-24-4-5-4	2.4	68.0	75	190	102.5	2.1	3.3
CML444	2.3	74.3	81	135	77.5	2.5	2.8
F1-14-79-4-1-3	2.2	67.3	72	195	113.8	2.6	3.5
TESTR 153	1.9	68.5	71	169	93.8	3.0	3.3
JI-304	1.6	69.3	73	111	73.8	2.3	2.5
JI-30-18	1.5	78.3	81	146	87.5	2.0	2.8
JI-303	1.5	75.8	81	136	75.0	2.4	3.0
TESTR 156	1.4	71.5	77	146	72.5	2.4	2.5
CML204IR	1.4	77.8	86	134	83.8	2.4	3.5
EARLY-N-POP-7-13-5-1	1.4	74.8	84	133	83.8	2.5	2.8
JI-30-22	1.3	75.8	81	138	87.5	2.1	3.8
TESTR 151	0.4	80.0	82	165	87.5	3.1	2.5
E11-133/7/44-6-3-17-3-2	0.2	73.5	87	159	92.5	2.0	2.0
CML206//56/44-6-3-7-1	0.1	75.0	85	150	90.0	2.1	2.5
CML395-IR	0.1	88.8	92	87	62.5	2.3	2.0
TESTR 133	0.1	73.3	80	93	60.0	3.0	2.3
Mean	1.4	73.8	79.6	140.3	80.9	2.6	2.6
CV (%)	30.5	7.7	8.9	25.5	22.6	16.3	20.9
LSD( 0.05)	1.94	8.10	11.13	50.19	25.61	0.58	1.12
Significance( GxE)	***	***	***	***	***	***	***

<sup>\*, \*\*</sup>and \*\*\* indicate differences that are significant at P < 0.05, P < 0.01 and P < 0.001, respectively.

Stem lodging and ear aspect were significant at P < 0.05. Reaction to E. turcicum was highly significant (P < 0.001) among the inbred lines. The worst affected inbred lines were TESTR 133, TESTR 136, TESTR 151, TESTR 153, TESTR 150 and TESTR 132. These materials have not been screened for E. turcicum blight disease and had E. turcicum scores of 3 to 4. However, inbred lines TESTR 149, TESTR 139, TESTR 152 and TESTR 156 gave a score of less than 3. Most of the inbred lines with good E. turcicum scores were the KARI-Muguga lines (Table 2).

### *Striga*-infested environment

Significant differences among the inbred lines were observed (P < 0.05) for grain yield (Table 3). The mean grain yield was 2.1 t/ha and the range was 0.2 to 2.9 t/ha. Inbred lines JI-30-19, OSU231//56/44-6-4-17-3, F1-14-14-24-4-5-4, JI-30-18 and TESTR 156 were the top five best performers. They gave desirable grain yields of between 1.9 and 2.9 t/ha under artificial *Striga* infestation. Inbred line JI-30-20 gave the lowest yield (0.2 t/ha). Highly significant differences (P < 0.001) were observed for days to 50 % anthesis (AD) and days to 50 % silking (SD). The mean for AD was 68.4 days and with 65 to 86.8 days,

while the mean SD was 71.2 days and the range was from 69.5 to 72.8 days. Reaction to *E.turcicum* differed significantly among the inbred lines, similarly to what was observed under *Striga*-free conditions in the present study (Table 3). Thus *Striga* infestation does not appear to interfere with the manifestation of resistance or susceptibility to *E. turcicum*.

Significant genetic (P < 0.05) variations were observed in reaction to Striga infection among the maize inbred lines. A mean of 5.1 for Striga damage rating (SDR) was observed, and the range was from 2.5 to 6.5 in a scale of 1

to 9. Inbred lines with desirable SDR scores were identified as JI-30-18, CML 202IR, JI-30-19, JI-30-20, JI-30-22, TESTR 150, JI-30-21 and JI-30-16. These inbred lines had a score of between 2.5 to 4, which is considered resistant on a scale of 1 to 9 (Kim, 1994). Inbred line OSU231//56/44-6-4-17-3 had a score of 6 although it was among the top five best in terms of grain yield. This particular inbred line could be considered tolerant as the effect of *Striga* on grain yield performance was minimal.

**Table 3** Performance of selected maize inbred lines under artificial field *Striga* infestation across sites.

Genotypes	Grain yield (t/ha)	Days to 50% anthesis (days)	Days to 50 % silking (days)	E. turcicum (score 1-5)	Striga damage rating (score 1-9)	Striga count 8 WAP (m²)	Striga count 10 WAP (m²)	Striga count 12 WAP (m²)
JI-30-19	2.9	75.8	79.2	2.3	3.5	4.5	13.9	11.8
OSU231//56/44-6-4-17-3	2.4	65.0	72.7	3.0	6.0	10.7	27.9	46.7
F1-14-14-24-4-5-4	2.2	68.5	71.7	3.5	5.3	7.4	22.9	32.3
TESTR 156	1.9	73.8	80.7	3.5	5.3	3.6	5.6	9.4
JI-30-18	1.9	77.5	80.0	1.8	2.5	3.0	8.3	12.6
EARLY-N-POP-7-13-5-1	1.7	80.5	81.6	1.8	4.3	1.4	11.3	19.3
F1-14-79-4-1-3	1.7	75.5	76.2	3.5	4.5	10.9	16.4	24.1
JI-30—4	1.6	68.0	79.5	3.3	4.3	6.9	16.8	29.0
JI-30—3	1.6	78.0	79.9	2.5	4.8	3.1	9.7	15.1
CML206//56/44-6-3-7-1	1.6	76.5	78.3	1.8	4.8	2.1	11.2	21.1
TESTR 153	1.5	73.5	73.6	3.8	4.8	6.4	14.3	24.1
CML395-IR	1.2	74.6	82.3	3.0	4.3	2.2	7.8	19.8
CML202IR	1.2	75.0	79.7	2.8	3.3	2.3	9.2	14.4
JI-30-8	1.1	77.5	83.7	2.0	4.5	1.6	8.5	13.5
JI-30-16	1.1	78.3	80.5	2.0	4.0	3.9	11.3	27.4
TESTR 133	0.7	68.5	70.2	3.8	6.0	4.4	9.4	14.3
TESTR 152	0.7	85.2	90.9	2.5	5.8	4.0	9.6	11.0
CML444-IR	0.5	75.5	84.2	3.0	5.5	2.3	13.7	21.2
TESTR 132	0.4	75.5	77.8	4.0	5.0	7.1	8.6	10.0
JI-30-20	0.2	73.3	76.5	3.8	4.0	11.1	18.8	25.5
MEAN	1.2	73.8	77.5	2.8	4.5	5.9	15.2	22.3
CV (%)	27.4	8.7	9.1	24.7	30.4	23.5	24.8	29.9
LSD (0.05)	1.36	9.25	11.86	1.44	2.02	7.24	14.87	19.47
Significance (GxE)	*	***	***	**	*	**	*	**

<sup>\*, \*\*</sup>and \*\*\* indicate differences that are significant at P < 0.05, P < 0.01 and P < 0.001, respectively.

Data on *Striga* counts was highly significant (P < 0.001) 12 weeks after planting (WAP). Genetic variations among the inbred lines were observed in *Striga* counts at the 8, 10 and 12<sup>th</sup> WAP. The mean *Striga* count at 12 WAP was 22.3 *Striga* plants per m<sup>2</sup>, and the range was from 2.9 to 46.7 *Striga* plants per m<sup>2</sup> (Table 3). Inbred lines TESTR 139, TESTR 151, TESTR 152, TESTR 132, TESTR 150, TESTR 136, TESTR 156, TESTR 149, JI-30-21 and JI-30-19 gave the least number of *Striga* plants per m<sup>2</sup>.

Further assessment on the resistance of the maize inbred lines was done by examining the relationship between the yield performance and the *Striga* resistance traits. This was investigated through use

of a simple linear correlation coefficient in a combined analysis for the two sites. A highly significant (P < 0.001) and negative correlation was observed between grain yield and SDR (r = -0.67\*\*\*). A positive but not significant correlation coefficient between grain yield and *Striga* counts was observed (Table 4). A positive and significant correlation coefficient was observed between ears per plant (EPP) and grain yield (r = 0.39\*\*\* and days to 50 % anthesis (r = 0.33\*\*\*) while a negative and significant correlation between EPP and anthesis silking interval (ASI) (r = -41\*\*\*) was observed. *Striga* counts at 8 WAP was highly correlated to *Striga* counts at 10 WAP (r = 0.81\*\*\*\*) and the 12WAP (r = 0.77\*\*\*\*) (Table 4).

**Table 4** Correlation between grain yield, agronomic traits, and *Striga* resistance traits under *Striga*-infested conditions across sites.

Traits	YLD	AD	SD	ASI	PH	ЕН	EPP	GLS	RUST	TURC	EA	PA	SDR	STR8	STR10
YLD	1.00														
AD	-0.23														
SD	-0.38*	0.76***													
ASI	-0.13	-0.12	0.37*												
PH	0.35	-0.40**	-0.38**	-0.15											
EH	0.58***	-0.40**	-0.47***	-0.32*	0.43**										
EPP	0.39**	0.33**	-0.03	-0.41**	-0.10	0.20	1.00								
GLS	0.34**	-0.09	-0.08	-0.14	0.06	0.39**	0.14	1.00							
RUST	0.14	-0.16	-0.15	-0.25	0.03	0.29	0.26	0.33**	1.00						
TURC	-0.26	-0.47	-0.20	0.17	0.22	0.01	-0.57***	0.17	0.12						
EA	0.46***	0.11	0.21	0.18	-0.08	-0.30	-0.44	-0.32*	-0.25	0.28	1.00				
PA	0.75***	-0.36**	-0.57***	-0.10	0.33*	0.59***	0.31	0.22	0.11	-0.07	-0.20				
SDR	-0.67***	-0.16	-0.06	0.25	-0.05	-0.49***	-0.24	-0.24	-0.16	0.28	0.16	-0.28	1.00		
STR8	0.17	0.04	0.19	0.37*	0.11	0.12	-0.17	-0.11	-0.15	0.04	0.18	0.01	0.22	1.00	
STR10	0.27	-0.13	-0.04	0.27	0.24	0.23	-0.17	-0.11	-0.14	-0.01	0.21	0.13	0.15	0.81***	1.00
STR12	0.21	-0.15	-0.01	0.35*	0.15	0.18	-0.17	-0.21	-0.24	-0.08	0.22	0.12	0.15	0.77***	0.95***

<sup>\*, \*\*</sup> and \*\*\* indicate differences that are significant at P < 0.05, P < 0.01 and P < 0.001, respectively. Yld = Grain yield, AD = days to 50 % anthesis, SD = days to 50 % silking, ASI = anthesis silking interval, PH = Plant height, EH = ear height, GLS = Gray leaf spot, Turc = E.turcicum, SDR = Striga damage rating, STR8 = Striga counts 8WAP, STR10 = Striga counts 10WAP and STR12 = Striga counts 12 WAP

The *Striga* count at 10WAP was positively and highly correlated to *Striga* count at 12 WAP (r = 0.95\*\*\*). Grain yield was also found to be positively correlated to ear aspect (r = 0.46\*\*\*) and plant aspect (r = 0.75\*\*\*). It was clear that for the more resistant genotypes, *Striga* counts peaked at week 12 and declined towards the 14<sup>th</sup> week. Therefore the assessment of resistance at 12 WAP should be considered adequate. It is noted that the decline of *Striga* plants from 12 WAP could be attributed to plants dying after the host has succumbed to infestation at the maximum level and probably dying of host maize roots.

### **Pot experiment**

Highly significant differences were observed among the inbred lines in *Striga* counts at 10 WAP (Table 5), but not at 12 WAP. However at 14WAP the number of *Striga* plants which emerged was highly significant (P < 0.01). Flowering *Striga* plants per pot was not

significant at the 12 and 14 WAP, but it was highly significant at the 15 WAP. The number of *Striga* plants setting seeds per pot was not significant at 12 WAP, although significant differences were exhibited (P < 0.05) at 14 and 15 WAP. The number of *Striga* attachments observed was not significant. The mean number of *Striga* attachments per pot was 20.71 and the range was from 0 to 74.5 *Striga* attachments per pot (Table 5). These observations were similar to those observed in the field.

A simple linear correlation between the *Striga* resistance traits was computed. *Striga* attachments were found to be significantly correlated to *Striga* counts at 10 WAP (r = 0.25\*\*) and the 14 WAP (r = 0.31\*) (Table 6). The number of *Striga* plants setting seeds at 15 WAP was also significantly correlated to the number of attachments per pot.

**Table 5** Striga resistance traits in a pot experiment under artificial Striga infestation across seasons.

Genotypes	Striga count 10 WAP /m <sup>2</sup>	Striga count 12 WAP /m <sup>2</sup>	Striga count 14 WAP /m <sup>2</sup>	Flowering Striga plants 14 WAP /m <sup>2</sup>	Flowering Striga plants 15 WAP/m <sup>2</sup>	Striga plants setting seeds 14 WAP/m <sup>2</sup>	0 <b>1</b>	Striga attachments
CML202IR	0.00	0.00	-0.04	0.00	0.00	0.00	0.00	0.50
CML204IR	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
CML395-IR	0.00	0.00	0.00	0.00	0.00	0.00	0.00	1.50
CML444-IR	0.00	0.00	0.00	0.00	0.00	0.00	0.00	1.50
TESTR 151	1.05	1.05	1.30	0.15	0.15	0.15	0.15	28.00
TESTR 149	2.50	3.05	2.60	0.00	0.00	0.00	0.00	29.50
TESTR 150	2.80	2.95	2.70	0.35	0.35	0.00	0.15	7.50
CML444	2.55	2.85	2.75	0.40	0.50	0.30	0.40	32.50
MGA19-4-1	3.15	3.05	2.75	0.30	0.30	0.00	0.25	73.50
TESTR 139	2.50	2.80	2.85	0.00	0.00	0.15	0.30	25.00
TESTR 132	4.70	510.90	2.90	185.50	0.00	0.00	0.15	5.50
JI-30—3	2.50	2.70	2.90	0.15	0.15	0.00	0.00	19.50
JI-30-19	2.75	3.05	2.90	0.00	0.15	0.30	0.40	22.50
CML395	1.75	1.75	2.95	0.50	0.55	0.40	0.45	4.00
JI-30-19	3.35	3.30	2.95	0.15	0.15	0.30	0.30	9.50
F1-14-79-4-1-3	3.20	3.30	3.35	0.55	0.65	0.15	0.50	74.50
JI-30-7	2.65	3.10	3.40	0.00	0.00	0.15	0.30	44.00
JI-30-17	3.25	3.45	3.45	0.40	0.45	0.15	0.55	23.00
JI-30—4	3.45	3.50	3.55	0.45	0.65	0.15	0.50	14.50
JI-30-8	3.70	3.70	3.60	0.65	0.90	0.00	0.15	14.50
Mean	2.29	27.73	2.34	9.48	0.25	0.11	0.23	21.55
CV (%)	29.84	30.10	14.54	29.20	27.50	20.30	24.70	19.80
LSD (0.05)	1.61	243.20	0.83	88.70	0.49	0.28	0.45	59.87
Significance	***	NS	***	NS	**	*	*	NS

<sup>\*, \*\*, \*\*\*</sup> and NS indicate differences that are significant at P < 0.05, P < 0.01, and P < 0.001, respectively.

**Table 6** Correlation between *Striga* resistance traits in a pot experiment.

Traits	1	2	3	4	5	6	7	8	9
1. Striga count 10 WAP									
2. Striga count 12 WAP	0.33*								
3. Striga count 14 WAP	0.92***	0.04							
4. Flowering Striga plants 12WAP	0.34*	0.99***	0.06						
5. Flowering Striga plants 14WAP	0.32*	0.10***	0.03	0.99***					
6. Flowering Striga plants 15WAP	0.50***	-0.21	0.57***	-0.17	-0.21				
7. Striga setting seeds 12 WAP	0.33*	0.99***	0.03	0.99***	0.99***	-0.23			
8. Striga setting seeds 14 WAP	0.15	-0.14	0.25	-0.10	-0.14	0.36**	-0.10		
9. Striga setting seeds 15 WAP	0.41**	-0.07	0.47***	-0.06	-0.07	0.53***	-0.07	0.73***	
10. Striga attachments	0.25**	-0.13	0.31*	-0.15	-0.13	0.20	-0.14	0.12	0.31*

### **Discussion**

A broad range of genetic variation in Striga resistance traits was exhibited in this study particularly in the number of Striga plants emerged and the number of Striga plants attached. Similar results were reported by Amusan et al., (2008). Under Striga infested conditions, the days to 50 % flowering for the most susceptible inbred lines was delayed by about 5 days and some maize inbred lines did not reach days to 50 % silking. Cases of delayed flowering while testing several maize cultivars under different nitrogen levels were also reported by Kim et al., (1997). Our results also agree with these results in which some inbred lines never silked leading to reduction in yield due to lack of fertilization. The delay in flowering is a common observation in maize subjected to stresses other than Striga, for example drought stress.

The ear aspect of the tolerant and resistant inbred lines was significantly superior compared to that of the susceptible inbred lines. The usefulness of the ear aspect in the assessment of host plant response to *Striga* infection was also reported by other workers (Kim *et al.*, 1997). The inbred line JI-30-19 exhibited the best ear aspect and also gave the highest grain yield. The number of ears harvested from the maize inbred lines tested in

this study proved to be a major component of grain yield under *Striga* infestation as was previously reported by Kim, (1991).

Most of the inbred lines with field resistance to Striga had significantly fewer attached parasites as opposed to the susceptible inbred lines. These results were consistent with previous observations reported in maize (Kim, 1999; Amusan et al., 2008). Striga emergence in some moderately susceptible inbred lines was found to be similar to Striga emergence in some resistant and tolerant lines, as was observed in inbred lines tested in the field (Table 4). Previous results from several studies have shown that Striga emergence counts from tolerant maize cultivars and from moderately susceptible cultivars were significantly different. This discredits the use of Striga emergence counts as the only criterion to distinguish genetic control of Striga tolerance in maize (Kim, 1994; Kim and Adetimirin, 1997). This is probably because resistance may often be confounded by tolerance existing in the same germplasm.

A significant and negative correlation has been shown between grain yield and Striga damage rating (SDR) (Kim and Adetimirin, 1997; Amusan et al., 2008). Similar observations were made in the present study where a significant (P < 0.001) and negative correlation was recorded between grain yield and SDR (r = - 0.67\*\*\*). However there was no

significant correlation between grain yield and *Striga* counts as would have been expected.

In the present study, the observed significant and positive correlation between the attached and emerged *Striga* plants with the *Striga* damage rating and reduction in grain yield of the maize plants indicated that the possibility exists of selecting maize inbred lines with low SDR scores and *Striga* emergence, and with higher grain yields under *Striga* infection.

As was found in this study, the number of Striga attachments has similarly been shown in the past to correlate with the number of emerged parasites in the pots (Kim, 1999; Amusan et al., 2008). Several previous studies have revealed a strong correlation between attached Striga plants in pots and the number of emerged parasites in both pots and field. In the present study inbred lines TESTR 139, TESTR 151, TESTR 152, TESTR 132, TESTR 150, TESTR 136, TESTR 156, TESTR 149, JI-30-21 and JI-30-19 had significantly fewer emerged Striga plants compared to the susceptible lines. These results suggest the possibility of selection for field resistance to Striga by using both attached Striga and emerging Striga either in the pot or in the field.

Striga-resistant maize inbred lines were identified from among the diverse range of inbred lines tested. The maize inbred lines with fewer emerged Striga plants and low SDR scores were considered as the resistant lines, which confirm many previous studies in maize research. The IITA inbred lines were confirmed as having resistance since most of them supported very few emerged Striga plants. However the use of Striga counts as a criterion in selection for Striga resistance was found not to be the most appropriate. On many occasions a small number of emerged Striga plants caused heavy Striga damage in some of the inbred lines tested. A significant and negative correlation between grain yield and Striga damage rating was observed. The number of emerged Striga plants was found to be highly correlated to the number of Striga attachments on the maize roots. Through the use of the observed significant and positive correlation of the attached and emerged Striga plants with the Striga damage rating and reduction in grain yield of the maize plants, it is therefore possible to select maize inbred lines with low SDR scores and Striga emergence, and with higher yields under Striga infection.

### Acknowledgements

The authors would like to thank CIMMYT and KARI technical staff in all research stations where these materials were tested. Special thanks go to IITA and KARI for providing most of the inbred lines used in this study. This work was funded by a grant from Pioneer Hi-bred international through CIMMYT and the Syngenta Foundation for Sustainable Development partially funded the preparation of this manuscript.

### References

Amusan, I. O., Patrick, J. R., Menkir, A., Thomas, H., Ejeta, G. 2008. Resistance to *Striga hermonthica* in a maize inbred line derived from *Zea diploperennis*. New Phytologist, 178: 157–166.

Badu-Apraku, B., Fakorede, M. A. B., Lum, A. F. 2007. Evaluation of experimental varieties from recurrent selection for *Striga* resistance in two extra early maize populations in the savannas of West and Central Africa. Experimental Agriculture, 43: 183-200.

Berner, D. K., Kling, J. G. and Singh, B. B. 1995. *Striga* Research and Control. A perspective from Africa. Plant Disease, 79: 652-660.

Doggett, H., 1988. Sorghum, 2nd ed. Longman, London, UK.

Haussmann, B. I. G., Hess, D. E., Welz, H. G. and Geiger H. H. 2000. Improved methodologies for breeding *Striga* resistant sorghums. Field Crops Research, 66: 195-211.

Haussmann, B. I. G., Hess, D. E., Omanya, G.
O., Folkertsma, R. T., Reddy, B. V. S.,
Kayentao, M., Welz, H. G. and Geiger, H.
H. 2004. Genomic regions influencing resistance to the parasitic weed *Striga*

- hermonthica in two recombinant inbred populations of Sorghum. Theoretical and Applied Genetics, 109: 1005-1016.
- IFPRI, 2000. 2020 Projections IFPRI, Washington, DC, IRRI (1998) IRRISTAT for Windows, version 4.0. International Rice Research Institute, Manila.
- Kanampiu, F. and Friesen D. 2003. A New Approach to *Striga* Control. The Royal society of Chemistry, pp. 51-53.
- Kim, S. K. and 1991. Breeding maize for *Striga* tolerance and the development of a field infestation technique, Combating *Striga* in Africa, IITA, Ibadan, pp: 96-108.
- Kim, S. K. 1994. Genetics of maize tolerance of *Striga hermonthica*. Crop Science, 34: 900-907.
- Kim, S. K., Adetimirin, V. O., 1997. Responses of tolerant and susceptible maize hybrids to timing and rate of nitrogen under *Striga hermonthica* infestation. Agronomy Journal, 89: 38-44.

- Kim, S. K., Adetirimin, V. O., Akintunde, A. Y, 1997. Nitrogen effects on *Striga hermonthica* infestation, grain yield, and agronomic traits of tolerant and susceptible maize hybrids. Crop Science, 37: 711-716.
- Kim, S. K., Akintunde, A. Y. and Walker, P 1999. Responses of maize inbreds during development of *Striga hermonthica* infestation. Maydica, 441: 333-339.
- Ramaiah, K. V., Childley, V. L. and House L. R. 1991. A time course study of early establishment stages of the parasitic angiosperm *Striga asiatica* on susceptible sorghum roots. Annals of Applied Biology, 118: 403-410.
- Ransom, J. K., Eplee, R. E. and Langston, M. A. 1990. Genetic variability for resistance to *Striga asiatica* in maize. Cereal Research Communications, 18: 329-333.
- SAS, 2003. SAS ® 9.1 for Windows, Cary, NC.

### شناسایی رگههای خویش آمیخته ذرت مقاوم به علف هرز Striga hermonthica

هارون کارایا ٰ، نیوروگ کیاری ٔ ، استفان ماگو ٰ، فرد کانامپیو ٰ ٔ ، امانوئل آنیگا ؑ و جان ندریتو ؑ

۱- مرکز بینالمللی اصلاح ذرت و گندم (CIMMYT) صندوق پستی: ۱۰۴۱-۰۰۶۲۱، نایروبی، کنیا ۲- دانشگاه نایروبی، دانشکده کشاورزی، پردیس Kabete صندوق پستی: ۲۲۰۵۳، نایروبی، کنیا \* پست الکترونیکی مسئول مکاتبه: f.kanampiu@cgiar.org

چکیده: علف جادوگر (Striga hermonthica) یک علف هرز نیمه انگلی اجباری و خطرناک در غلات است که خسارت سنگینی به ارقام حساس ذرت در آفریقا وارد میکند. ایجاد مقاومت در گیاهان یکی از کاربردی ترین روشهای کنترل علف هرز Striga میباشد. در این تحقیق آزمایشها روی ۳۶ رگه خویش آمیخته ذرت در داخل گلدان و مزرعه در طی دو فصل زراعی پر باران در سال ۲۰۰۹ در ایستگاههای تحقيقاتي Kibos و Alupe كنيا انجام شد. هدف اصلى تحقيق تعيين نوسانات جوانهزني علف جادوگر و رابطه آن با میزان انگلی شدن ریشههای ذرت بود. اختلاف معنیدار (P<٠/٠٠١) بین رگههای خویش آمیخته ذرت از نظر میزان تولید در شرایط عاری از علف هرز Striga وجود داشت. میزان خسارت علف جادوگر (SDR) در ارقام مختلف ذرت دارای تفاوت معنی دار (P < ٠/٠۵) بود. ارتباط منفی و کاملاً معنی-داری بین میزان محصول و نرخ خسارت ناشی از علف هرز جادوگر مشاهده شد (r = -1/6۷). همچنین  $(r = \cdot /V\Delta)$  ووضعیت گیاه  $(r = \cdot /FA)$  و اندازه بلال  $(r = \cdot /FA)$  و وضعیت گیاه ( $(r = \cdot /V\Delta)$ وجود داشت. در آزمایشهای گلدانی تفاوت بسیار معنی داری (P < ۰/۰۱) بین لاینهای خویش آمیخته ذرت از نظر مقاومت به علف جادوگر مشاهده شد. میزان انگلی شدن ریشه ذرت با علف جادوگر با تعداد گیاهچههای سبز شده ارتباط معنی داری داشت همبستگی معنی داری بین تعداد مکینههای علف جادوگر و تعداد گیاهچههای سبز شده در هفته دهم  $(r = \cdot/\Upsilon 0)$  و چهاردهم  $(r = \cdot/\Upsilon 1)$  بعد از کشت ذرت در گلدان مشاهده شد. رگههای خویش آمیخته JI-C-19 و IJ-C-19-0SU231//56/44-6-4-17-3 به عنوان مقاوم ترین رگهها (لاینها) نسبت به علف هرز جادوگر بودند زیرا در شرایط بدون علف هرز و همچنین در حضور علف جادوگر عملکرد مشابه و قابل قبولی داشتند. این رگههای خویش آمیخته ذرت را میتوان برای تهیه واریتههای ذرت مقاوم به علف جادوگر مورد استفاده قرار داد.

واژگان کلیدی: رگههای خویش آمیخته، ذرت، علف هرز جادوگر، کنیا