

# Fungi associated with root and crown rot of wheat in Khuzestan province, Iran

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Abstract: To identify the fungi associated with foot and root rot of wheat in the Khuzestan province, diseased samples were collected at all growth stages in three growing seasons of 2004-2007. Pieces of infected parts of the root and foot were surface sterilized and cultured on acidic and non acidic PDA media. One hundred and fifteen isolates were obtained and on the basis of macroscopic and microscopic characters and valid keys were identified as Fusarium solani, F. equiseti, F. moniliforme, F. subglutinans, F. sambucinum, F. culmorum, F. proliferatum, F. pseudograminearum, F. longipes, F. avenacium, F. nygamai, F. semitectum, F. lateritium, Rhizoctonia solani, Bipolaris sorokiniana and Pythium sp. F. equiseti, F. solani and F. culmorum with occurrence frequencies of 16.07, 16.07 and 12.5 % respectively were more frequent than all other species. Results of Pathogenicity tests indicated that F. pseudograminearum and F. culmorum were the main fungi associated with wheat root and crown rot disease in khuzestan while other Fusarium spp. such as F. equiseti are probably aggravated by moisture stress at different growth stages of crop due to poor irrigation management. Rhizoctonia solani with occurrence frequency of 8.03 % in some regions such as Ahvaz, Shoush, Shoushtar and Baghmalek was important agent of crown and root rot of wheat second to Fusarium species. Bipolaris sorokiniana and Pythium sp also caused crown and root rot but with less frequency percent.

Keywords: Fusarium spp., wheat, frequency, water stress, Khuzestan

#### Introduction

Wheat (*Triticum aestivum L*.) is the most important crop in Khuzestan province. Root and crown rot diseases are one of the most common and destructive diseases of wheat which occur every growing season. They cause early maturity and result in white heads and incomplete grain fill (Wallwork, 2000). Whole plants or individual tillers may be stunted. Necrotic brown lesions on seminal roots, crown, subcrown internodes and stem tissues are observed. Several different fungi cause root and crown rot of wheat, including Fusarium, Rhizoctonia, Gaeumannomyces, Drecheslera, Exserohilum, and Bipolaris spp. (Smilev et al., 2005: Cook et al., 2002: Freeman and Ward, 2004). Fusarium is one of the most important fungi of soil microflora (Leslie and Summerell, 2006). Some Fusarium species cause crown and root rot of wheat in temperate and subtropical regions (Windles and Holen, 1989). In recent years several researches were done in many wheat producing areas of Iran in this regard. For instance Rhizoctonia solani, R. cerealis, F. graminearum and Gaeumannomyces graminis were isolated from Mzandaran (Foroutan et al., 1995). Pathogenicity of F. culmorum, F. avenaceum and F. acuminatum were confirmed in

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Fars province (Ravanlou and Banihashemi, 1999). The predominant pathogen causing common root and crown rot in Moghan area, northwest of Iran was identified to be *Bipolaris sorokiniana* (Hajigharari, 2009). *Rhizoctonia solani* is more severe in sandy soils where rainfall is lower (Gill *et al.*, 2001). Take-all caused by *Gaeumannomyces graminis* var *tritici* is the least common agent of root rot but may be the most damaging, taking all of the yield. Take-all is typically a greater problem on winter wheat growing in wet soils (Cook, 2001).

This research was conducted to identify and to evaluate the fungi associated with root and crown rot of wheat in Khuzestan province.

# **Materials and Methods**

# Sample collection

In this study the Khuzestan province was divided into four regions on the basis of geographical position; north and northwest, south and southwest, northeast and southeast. Wheat plants showing symptoms of stunting, chlorosis and necrosis on sub-crown internodes and crown at the seedling to maturity stages were collected (5 to 20 fields were selected on the basis of the area under cultivation). Ten symptomatic plants were collected from each field and each sample was removed with as much of the root system as possible. Samples were stored in a dry and cold place. All of the samples were collected between November till April during 2004-2007.

# **Fungal isolation**

Isolation of fungi associated with root and crown rot were made after surface sterilization. Each sample was rinsed under running water for 15 min. Five tissue pieces from root, crown and internodes (5mm) were surface sterilized with 1.5 % sodium hypochlorite for 1-2 min and washed two times in sterile distilled water. These pieces were plated onto Potato Dextrose Agar (PDA) supplemented with 1 drop lactic acid. Cultures were incubated at 20 °C for three days (Singleton *et al.*, 1990). *Fusarium* spp. from infected tissue were isolated by Nash & Snyder medium and we used Rifampicin - amended PDA for isolation of *Gaeumannomyces graminis*  var *tritici* (Duffy & Weller 1994) from suspected plants. Pure cultures of all fungi were prepared by hyphal tip and single spore culture methods (Nelson *et al.*, 1983) and were subcultured on PDA for inoculum production.

#### **Identification of fungi**

Fusarium species were transferred onto Carnation-Leaf Agar (CLA), incubated under a combination of white and near-ultraviolet fluorescent lights with a 12 h photoperiod and 23/25 °C night/day temperature cycle. The cultures were identified based on general colony morphology, morphology of microconidia, macroconidia, conidiophores, false heads, sporodochia, chlamydospore formation, and using Fusarium diagnostic keys (Leslie and Summerell, 2006). Other fungi except Pythium sp. were identified according to their descriptions (Windels & Holen, 1989; Sneh et al., 1991).

#### Pathogenicity test

Pathogenicity tests were conducted on seedlings of cv. Chamran. One flask was filled with 200g of moistened wheat seeds. Seeds were autoclaved twice at 121 °C for 2 h. A plug of PDA culture, 5mm in diameter, was put in each flask. The flasks were incubated at 25 °C for 15 days and were shaken occasionally by hand. Then the colonized seeds were ground in a blender and mixed with pasteurized soil (1:5 v/v). Pots (30 cm in diameter) were filled with soil. Ten surface sterilized seeds were sown per pot. The experiment was conducted in greenhouse using complete randomized design with three replications. The pots were divided into two groups on the basis of irrigation. One group was irrigated normally using 100 ml of water/day. The second group was irrigated normally in the early stages of growth but two periods (10 days) of water stress were imposed afterwards (Rice and Geraldj, 1981). Seedlings in each pot were checked after four weeks for disease symptoms and were graded according to a 2 - digit pathogenicity scale (0 and 1) as recommended by Rice and Geraldi, 1981; whereby 0 indicates without symptoms, 1 = with symptom.

#### Results

A total of 112 fungal isolates were collected from wheat growing fields (Tables 1, 2) and identified on the basis of Fusarium diagnostic keys and other relevant keys (figure 1).

Table 1 Wheat root - and crown rot - associated fungi isolated from different regions in Khuzestan

| Region                 | Species                                | No. of<br>Isolates | Frequency<br>% |
|------------------------|--|--------------------|----------------|
| Ahvaz                  | Fusarium solani                        | 5                  | 38.46          |
| Anvaz                  | F. equiseti                            | 3                  | 23.07          |
|                        | F. moniliforme                         | 2                  | 15.38          |
|                        |  | 1                  |                |
|                        | F. subglutinans                        |                    | 7.69           |
| D L                    | Rhizoctonia solani                     | 2                  | 15.38          |
| Dasht<br>azadegan      | F. solani                              | 3                  | 23.07          |
|                        | F. equiseti                            | 2                  | 15.38          |
|                        | F. sambucinum                          | 2                  | 15.38          |
|                        | F. culmorum                            | 6                  | 46.15          |
| Shoush                 | F. proliferatum                        | 2                  | 28.57          |
|                        | F. pseudograminearum                   | 1                  | 14.28          |
|                        | Pythium sp.                            | 2                  | 28.57          |
|                        | R. solani                              | 2                  | 28.57          |
| Andimeshk              | F. solani                              | 3                  | 37.5           |
|                        | F. moniliforme                         | 2                  | 25             |
|                        | F. longipes                            | 2                  | 25             |
|                        | B. sorokiniana                         | 1                  | 12.5           |
| Dezful                 | F. equiseti                            | 3                  | 42.85          |
|                        | F. avenaceum                           | 1                  | 14.28          |
|                        |  | 1                  | 28.57          |
|                        | F. pseudograminearum<br>B. sorokiniana | 1                  | 14.28          |
| 01 1/                  | Б. sorokiniana<br>F. culmorum          | 3                  |                |
| Shoushtar              |  |                    | 25             |
|                        | F. solani                              | 3                  | 25             |
|                        | F. sambucinum                          | 2                  | 16.66          |
|                        | F. nygamai                             | 1                  | 8.33           |
| Masjedsoleyman<br>Izeh | R. solani                              | 3                  | 25             |
|                        | F. moniliforme                         | 2                  | 25             |
|                        | F. equiseti                            | 3                  | 37.5           |
|                        | F. longipes                            | 3                  | 37.5           |
|                        | F. proliferatum                        | 2                  | 22.22          |
|                        | F. moniliforme                         | 2                  | 22.22          |
|                        | F. culmorum                            | 5                  | 55.55          |
| Baghmalek              | R. solani                              | 2                  | 40             |
|                        | F. solani                              | 1                  | 20             |
|                        | F. semitectum                          | 2                  | 40             |
| Mahshahr               | F. subglutinans                        | 1                  | 25             |
|                        | F. avenaceum                           | 3                  | 75             |
| Hendijan               | F. equiseti                            | 3                  | 30             |
|                        | F. pseudograminearum                   | 4                  | 40             |
|                        | F. solani                              | 2                  | 20             |
|                        | F. lateritium                          | 1                  | 10             |
| Ramhormoz              | F. solani                              | 1                  | 25             |
| IIIOL                  | F. proliferatum                        | 3                  | 23<br>75       |
|                        | F. subglutinans                        | 2                  | 66.66          |
|                        |  | 2                  | 33.33          |
| Haftkel                | F. nygamai<br>F. avanaaaum             | 1                  | 33.33<br>18.18 |
|                        | F. avenaceum                           | 2                  | 18.18          |
|                        | F. longipes                            |                    |                |
|                        | F. lateritium                          | 3                  | 27.27          |
|                        | F. equiseti                            | 4                  | 36.36          |

| Species              | Total<br>Number | Total Frequency<br>(%) |  |
|----------------------|-----------------|------------------------|--|
| F. solani            | 18              | 16.07                  |  |
| F. equiseti          | 18              | 16.07                  |  |
| F. culmorum          | 14              | 12.5                   |  |
| Rhizoctonia solani   | 9               | 8.03                   |  |
| F. moniliforme       | 8               | 7.14                   |  |
| F. proliferatum      | 7               | 6.25                   |  |
| F. pseudograminearum | 7               | 6.25                   |  |

7

6

4

4

4

2

2

2

2

**Table 2** Total number and frequency % of fungal isolates

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6.25

5.35

3.57

3.57

3.57

1.78

1.78

1.78

1.78

#### Pathogenicity test

F. longipes F.avenaceum

F. subglutinans

F. sambucinum

F. lateritium

F. nygamai

Pythium sp. B. sorokiniana

F. semitectum

Results showed that Fusarium pseudograminearum and F. culmorum caused root and crown rot under normal irrigation regime, whereas all of the other F. spp. caused disease only after water stress. R. solani and Bipolaris sorokiniana also produced visible symptoms. In all replications of pathogenicity experiments with visible symptoms, each casual agent was re-isolated from infected tissues.

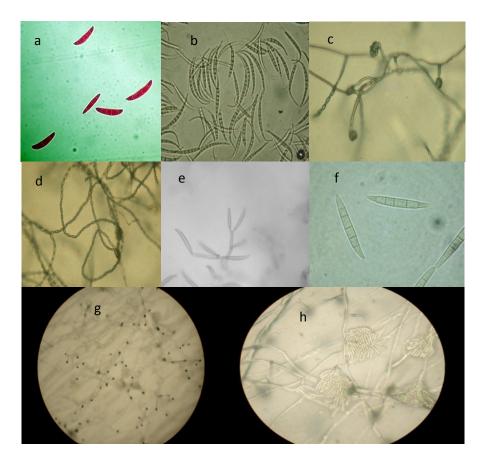
# **Frequencies and distribution of species**

F. solani and F. equiseti were the most frequent species while the F. nygamani, F. semitectum, Pythium sp. and B. sorokiniana were the least frequent ones (Table 2).

The two most common species contained about 32 % of the isolates. The distribution of the prevailing species with relation to different regions was calculated as 92 % for F. solani and 69 % for F. eqiseti (Fig. 2).

## Discussion

Samples were taken from north and northwest, south and southwest, northeast and southeast of Khuzestan. Drought is a serious problem in south and south east such as Ahvaz and Dashte Azadegan and also in some regions of northeast such as Masjedsoleyman.



**Figure 1** Microscopic characteristics of some *Fusarium* isolates: a) macroconidia of *Fusarium culmorum* 40x, b) macroconidia of *Fusarium equeseti* 40x, c) false heads of *Fusarium proliferatum* 40x, d) spore chain of *Fusarium proliferatum* 40x, e) macroconidia of *Fusarium semitectum* 40x, f) macroconidia of *Fusarium semitectum* 40x, g) false heads of *Fusarium solani* 10x and h) macroconidia of *Fusarium solani* 10x.

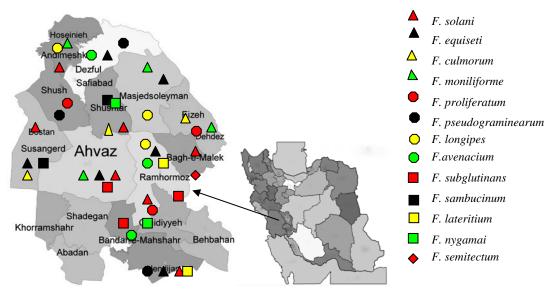


Figure 2 Distribution map of *Fusarium* species in Khuzestan province Iran.

This study showed that Fusarium species were the predominant pathogens causing root and crown rot of wheat in Khuzestan province, while other fungi such as B. sorokiniana, R. solani and Pythium sp. were observed as well. Some of Fusarium species are commonly associated with warm dry condition and are greatly influenced by temperature and rainfall (Chekali al., 2011). et Fusairum pseudograminearum and F. culmorum were pathogenic and infected the seedlings without any water stress while the other Fusarium spp. caused disease symptoms in roots of plants only under water stress condition. Among the identified Fusarium species F. equiseti, F. solani and F. culmorum were more frequent. F. equiseti was observed in all areas including regions with drought and water stress.

The three species of F. equiseti, F. solani and F. culmorum have been collected from stubbles in South Africa (Klassen et al., 1991) as well. They were limiting factors for wheat production in Nigeria (Los, et al., 1994). F. equiseti has been reported as a potential infective agent (Burgees et al., 1995). It was collected from root and crown rot of wheat and was more frequent than other species in Lorestan province, Iran (Darvishnia et al., 2007). F. solani is cosmopolitan and causes crown and root rot on most plants (Burgess et al., 1995). F. culmorum is widespread seed borne pathogen and the causal agent of crown and root rot of wheat and barley (Burgess et al., 1994). This species was isolated from crown and root of wheat and barley in Iran (Vafaie et al., 2001) and its pathogenicity was confirmed on date palm (Mosavi-jorof et al., 1999) and faba bean in Khuzestan (Azimi et al., 2005). F. avenacium has been mostly isolated from barley and wheat (Nelson et al., 1983). F. nygamai, F. lateritium, F. equiseti, F. solani, F. culmorum, F. moniliforme, F. proliferatum, F. longipes, F. sambucinum and F. avenaceum have previously been reported from Iran (Zare and Ershad, 1997).

*F. pseudograminearum* and *F.culmorum* were mostly isolated in late tillering stage and early heading stage but in seedling stage other species of *Fusarium* along with *Pythium* and *Rhizoctonia* could be isolated. These results indicated that the host may be weakened by the mentioned fungi in early stages of growth and its susceptibility to other pathogens is thereby increased. Isolation of fungi in different growth stages showed that the greatest number of casual agents were different *Fusarium* species which probably become more aggressive by water stress at different growth stages of the crop. Distribution of *Fusarium* species was not uniform and depended on water stress so that a species that was not previously seen in a region could be observed after water stress.

Pathogenicity tests of *B. sorokiniana* and *R. solani* were positive but in the case of *Pythium* sp. despite being pathogenic, no distinguishable symptoms similar to those of *B. sorokiniana*, *R. solani* or *Fusarium* spp. were observed. *Gaumannomyces graminis* var. *graminis* was not observed in cultures. Most soils in Khuzestan are often calcareous and it seems this fungus can not grow in such soils.

Since the rotation of wheat, canola and corn is a common agricultural practice in Khuzestan province, the effect of rotation in combination with water stress on prevalence of wheat crown and root pathogenic fungi may be considered for future research.

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# قارچهای همراه با پوسیدگی طوقه و ریشه گندم در استان خوزستان

محمدرضا اصلاحي

اهواز – بلوار گلستان - مرکز تحقیقات کشاورزی و منابع طبیعی خوزستان پست الکترونیکی مسئول مکاتبه: mr\_eslahi@yahoo.com

چکیده: برای شناسایی قارچهای همراه با طوقه و ریشه گندم در استان خوزستان، نمونههای بیمار در همه مراحل رشد در سه فصل کشت در سالهای ۱۳۸۳ تا ۱۳۸۶ جمع آوری شدند. قطعاتی از قسمتهای آلوده ریشه و طوقه ضدعفونی سطحی شد و روی محیط کشت سیبزمینی- دکستروز- آگار اسیدی و غیر اسیدی کشت گردید. یک دو پنجاه جدایه از نمونههای کشت داده شده به دست آمد و براساس خصوصیات میکروسکوپی و گردید. یک دو پنجاه جدایه از نمونههای کشت داده شده به دست آمد و براساس خصوصیات میکروسکوپی و ماکروسکوپی و کلیدهای معتبر گونههای کشت داده شده به دست آمد و براساس خصوصیات میکروسکوپی و *K. subglutinans, F. moniliforme F. equisetiFusarium solani* معتبر گونههای *Comparent Journal of the equiser for a seconder of the equiser of the equiser for a seconder of the equiser of the equiption of the equipti. The equiption of the equ* 

واژگان کلیدی: Fusarium pseudograminearum, F. culmorum, F. equiseti و تنش آبی