

Research Article

Evaluation of fungal isolates as possible biocontrol agents against *Striga hermonthica*

Victor Kagot^{1*}, Sheila Okoth¹, Fred Kanampiu², Peter Okoth³ and Edna Mageto²

1. School of Biological Sciences, University of Nairobi P. O. Box: 2586–00200 Nairobi, Kenya.

2. International Maize and Wheat Improvement Centre (CIMMYT) P. O. Box: 1041- 00621 Nairobi, Kenya.

3. International Centre for Tropical Agriculture (CIAT) P. O: Box: 30677- 00100 Nairobi, Kenya.

Abstract: *Striga hermonthica* is a parasitic weed which largely constrains maize and sorghum production in Western Kenya. The weed mostly invades small scale farms and depending on severity, it may cause damage ranging from 10% to complete crop failure thereby aggravating the food insecurity in that region. This study aimed at evaluating fungal isolates as possible biocontrol agents against the weed. Fungi were isolated from diseased *Striga hermonthica* and their virulence efficacy against the weed tested in a greenhouse. All the fungal isolates tested caused infection and consequently death of the weed. *Fusarium incarnatum* had the highest infection rate of 92% followed by *Gibberella intricans* and *F. chlamydosporum* at 90% each. *Fusarium oxysporium* caused the highest mortality of 60% with *Gibberella intricans*, causing the least mortality of 36%. *F. oxysporium* was the most aggressive and potent fungal isolate against the weed hence a suitable candidate for exploitation as a mycoherbicide against the weed.

Keywords: *Striga hermonthica*, *Fusarium*, Biocontrol, Mycoherbicide

Introduction

Striga hermonthica is a root parasitic weed which causes severe constrains in cereal crop production in the sub Saharan Africa by parasitizing the roots of the host crop. The parasite, attaches itself to the roots of its host from where it siphons nutrients and water intended for the plant to grow (Jamil *et al.*, 2012; Ndambi *et al.*, 2011). Several species of *Striga* have been identified worldwide with *S. hermonthica* being the most notorious and causing serious damage to the agro economic systems in Western Kenya (Gacheru *et al.*,

2002). Agriculture is the mainstay economic activity in Western Kenya region with cereals being the main source of food and income. Infestation by *S. hermonthica* has become a real threat to maize and sorghum cultivation in the region. In the Lake Victoria Basin of Kenya alone, 0.24 million hectares of arable land, which is 15% of the total arable land in the region is infested. This causes yield losses between 10-100% and or monetary losses of upto US\$ 41 million annually (Jamil *et al.*, 2012). In many cases, the damage caused by the weed is so severe that a farmer can only harvest a paltry 0.5tons per hectare of maize instead of the potential 5tons, or zero yields under heavy infestation (Olwenya, 2012).

The *S. hermonthica* menace is worsening due to continuous mono-cropping of maize and sorghum, little effort to control the weed and

Handling Editor: Dr. Mohammad Mehrabadi

* **Corresponding author**, e-mail: kagovic14@yahoo.com

Received: 6 August 2013, Accepted: 27 January 2014

Published online: 27 January 2014

the increasing population which is increasing pressure on the already limited arable land. In return, this leads to a decline in the soil fertility, thus creating favourable conditions for the weed to flourish (Esilaba, 2006). This weed is reported to be spreading rapidly to high land areas previously thought to be immune to it (Gacheru *et al.*, 2002). The currently recommended control strategies against the weed are ineffective due to the big disparity between the cost of the control strategies and the farmers' socioeconomic status (Atera *et al.*, 2013). Hence alternative, affordable and easy to use control measures should be developed to alleviate the *Striga* menace. This project aimed at evaluating fungal isolates as possible biocontrol agents against *Striga hermonthica*.

Methods

Isolation of fungal species infecting *Striga hermonthica*

Diseased *S. hermonthica* plants showing necrosis, spots on the leaves and stem, wilting and other abnormalities were uprooted from farms in Kibos–Kisumu (GPS coordinates: latitude 0.03861, longitude 34.815965, elevation 1196) and Alupe–Busia (GPS coordinates: latitude 0.50372514, longitude 34.1214814, elevation 1157 meters) research stations in Kenya, placed in brown paper bags and transported to the laboratory. The diseased parts were cut using a laboratory scalpel into pieces of about 4-7mm, surface-sterilized with 1% sodium hypochlorite (NaOCl) for 1½ minutes and rinsed 3 times in sterile distilled water. The cut plant parts were then dried with sterilized filter papers and some placed on Peptone PCNB Agar (PPA) while others on potato dextrose agar (PDA) half strength (Rahjoo *et al.*, 2008). For purification and subsequent morphological identification of the fungus which grew, the isolates were transferred on to potato-dextrose agar (PDA), Spezieller Nährstoffarmer Agar (SNA) and Carnation Leaf Agar CLA (Kwasna and Bateman, 2007). All the cultures were incubated at 25 °C for two to four weeks.

Cultural characters were assessed by eye and by microscopic examination. Colony morphology and colour (reverse and front) were recorded from cultures grown on PDA. The morphology of macroconidia was assessed from cultures grown on CLA while the morphologies of microconidia, conidiogenous cells and chlamydospores were assessed from cultures grown on SNA. Morphological identifications of the *Fusarium* isolates were made using the criteria of Gerlach and Nirenberg (1982) and Leslie and Summerell (2006). The non *Fusarium* isolates were identified using the criteria of Dugan (2006).

Molecular characterization of the isolated fungi

Molecular characterization of the isolated fungi was done to confirm their morphological identities. DNA was extracted from fungal isolates grown on PDA plates for 7 days. Their mycelia were harvested and resuspended in nuclease free water. Total DNA was extracted from the resuspended mycelia of each isolate (50-100 mg wet weight) using a ZR Fungal/Bacterial DNA MiniPrep Kit (Zymo research, South Africa) according to the manufacturer's instructions.

PCR amplification

The PCR amplifications were performed using the DreamTaq Green PCR Master mix (Thermo Scientific). The amplification reactions were carried out in volumes of 50 µL containing; 25 µL DreamTaq Green PCR Master mix, 1 µM of each primer i.e the forward primer and the reverse primer, 1µg of the template DNA and 23 µL of nuclease free water. The PCR reaction was carried out in a thermal cycler as follows: 1) 1 cycle of initial denaturation at 95 °C for 3 minutes; 2) 35 cycles of the following: denaturation at 95 °C for 30 seconds, annealing at 57 °C for 30 seconds and extension at 72 °C for 1 min; 3) final extension at 72 °C for 10 min, followed by cooling at 4 °C until recovery of the samples. Amplification products were visualized in 1.2% agarose gels stained with

ethidium bromide. The gel was photographed under UV light at 254 nm (Kwasna and Bateman, 2007). The primers used in the PCR were; TEF primers (TEF1 5'-ATGGGTAAGGARGACAAGAC-3' and TEF2 5'-GGARGTACCAGTSATCATGTT-3') for the *Fusarium* species and ITS primers (ITS F 5'-AACTCCCAAACCCCTGTGAACATA-3' and ITS R 5'-TTTAACGGCGTGGCCGC-3') for the non *Fusarium* species. The PCR products were then sequenced and edited using the Geneious program. The obtained DNA sequences were blasted using the NCBI BLAST (National Center for Biotechnology Information - Basic Local Alignment Search Tool) to reveal their identities. To reveal the relatedness of the isolates alignment of all the sequences was done using Clustalx 2.1 software and Phylogenetic analyses were conducted using MEGA version 5.1 using the Neighbour Joining (NJ) method (Tamura *et al.*, 2011). In the NJ analysis, distances were calculated using the Kimura 2-Parameter model and bootstrap tests performed with 1,000 replications (Tamura *et al.*, 2004; Felsenstein, 1985).

Virulence efficacy of the fungal isolates on *Striga hermonthica*

The most frequently isolated species were tested for their efficacy as biocontrol agents against *S. hermonthica*. The inoculums were prepared by scraping the fungal hyphae into Armstrong medium and incubating it in a shaker at 25 °C at 100rpm for 5 days to produce spores. The spores were filtered through two layers of sterile cheesecloth into a 50ml falcon tube and centrifuged at 3500rpm for 10minutes, the supernatant was discarded and the pellet (conidia) washed twice with de-ionized autoclaved water. The spores were suspended in 350ml sterile distilled water. With the aid of a haemocytometer the conidia concentration was adjusted to 2×10^6 conidia per ml. Tween 20 surfactant (Polyoxyethylene 20-sorbitan monolaurate) was added to the conidial suspension before inoculation at the rate of 3 drops per liter (Booth, 1971).

Green house trials

Maize was grown in 5- litre plastic pots, in a screenhouse at Kibos–Kisumu research stations. The pots were filled with 5 kg of pure, filtered soil. Five replicates were used for each fungal isolate being tested. Prior to sowing, the pots were infested by mixing about 2000 to 3000 *S. hermonthica* seeds (in *Striga* seeds/ sand mixture) into the soil layer 5-10 cm below the surface. Prior to inoculation, all the non - *S. hermonthica* weeds growing in the pots were uprooted. Excess *S. hermonthica* were also uprooted so that each pot had only 10 emerged *S. hermonthica* plants. Eight weeks after planting, when the *S. hermonthica* plants were approximately 5-15 cm tall, they were inoculated with the different fungal isolates. For each fungal isolate, 50 *S. hermonthica* plants were inoculated. The inoculum was sprayed throughout each *S. hermonthica* plant. Following inoculation, the *S. hermonthica* plants were assessed for disease symptoms at a 5 day interval for 3weeks. Plants with observed lesions and abnormal colour (maroon) changes on the leaves and stems were designated as infected. At the end of the experiment, the fungal isolates were then re-isolated from the infected *S. hermonthica* onto PDA plates to confirm that they were the ones previously inoculated. The most infectious fungal isolate was determined by counting the infected plants and then expressing it in percent. The number of dead plants after 22 days was used to determine the mortality rate and therefore the most virulent isolate.

Results

Fungal species isolated

Ten fungal isolates were recovered from diseased *S. hermonthica* collected from Kibos and Alupe. *Fusarium* spp. were isolated from *S. hermonthica* collected from Alupe as well as from Kibos. *Hypocrea stellata* was isolated from *S. hermonthica* collected from Alupe, while *Colletotrichum gloeosporioides* and *Arthroderma otae* were isolated from *S. hermonthica* collected from Kibos. *Fusarium oxysporium* was the most

frequent fungal species isolated followed by *F. chlamydosporium* then *F. equiseti*. The least frequent species were *Colletotrichum gloeosporioides* and *Hypocrea stellata* (Table 1).

Table 1 Frequency of isolation of the fungal species isolated from diseased *Striga hermonthica*.

Identity of the Fungus as in the Ncbi Blast	Origin (Area)	Origin (Part of Plant)	Frequency of Isolation
<i>Fusarium equiseti</i>	Kibos	Leaves	11
	Alupe	Leaves	9
<i>Fusarium verticilloides</i>	Kibos	Leaves	8
	Alupe	Leaves	10
<i>Fusarium oxysporum</i>	Kibos	Leaves	12
	Alupe	Leaves	14
	Kibos	Flowers	4
<i>Fusarium incarnatum</i>	Kibos	Leaves	8
	Alupe	Leaves	8
<i>Fusarium chlamydosporium</i>	Kibos	Leaves	10
	Alupe	Leaves	12
<i>Gibberella moniliformis</i>	Kibos	Stem	8
	Alupe	Stem	9
<i>Gibberella intricans</i>	Kibos	Stem	9
	Alupe	Stem	6
<i>Colletotrichum gloeosporioides</i>	Kibos	Stem	1
	Alupe		0
<i>Hypocrea stellata</i>	Kibos		0
	Alupe	Roots	1
<i>Arthroderma otae</i>	Kibos	Roots	2
	Alupe		0

Molecular characterization of the fungal isolates and their phylogenetic analysis

Isolates morphologically identified as *Fusarium* sp. and *Gibberella* sp. were positively amplified using TEF1 (20 base pairs) and TEF2 (21 base pairs) primers while the isolates morphologically identified as *Hypocrea stellata* and *Colletotrichum gloeosporioides*, were amplified using the ITS1 (24 base pairs) and ITS2 (17 base pairs) primers as shown in Fig. 1.

The blasted sequences of the extracted DNA showing positive bands in Fig. 1, agreed with the morphological identifications. Alignment of the sequences grouped the *Fusarium* sp. isolates into 3 clades. *Fusarium* spp. isolates demonstrated relatedness; however, they showed no relationship with *Arthroderma otae*. Close relationships were observed; amongst the different strains of *F. oxysporum* with the exception of the strain IBSD-GF13 which was in a different clade maybe because it was isolated from flowers while the other strains were isolated from leaves of *S. hermonthica*. Relatedness was also observed amongst the different strains of *F. Chlamydosporum*. Close relationships were observed between *F. equiseti* and its sexual state and between *F. verticilloides* and its sexual state. A relationship was also observed amongst the *Fusarium* spp. isolated from the leaves of *S. hermonthica*, however no relationship was observed amongst the *Gibberella* sp. isolated from the stems (Fig. 2).

Infection and mortality rates of the fungal isolates on *Striga hermonthica*

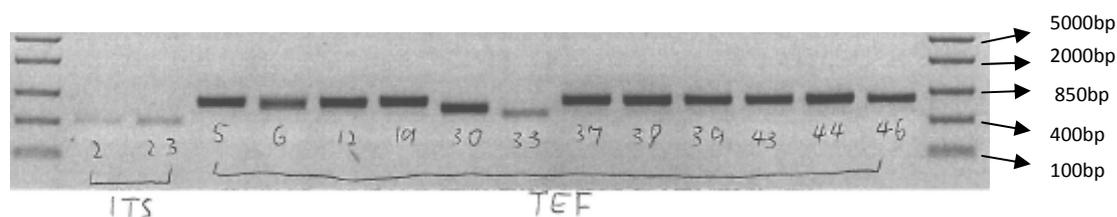
All the fungal isolates tested for their efficacy against *S. hermonthica* caused infections on the weed producing different symptoms as shown in Table 2. The number of weeds with observed lesions on the leaves and stems increased progressively with time. *Fusarium oxysporum* had the highest number of weeds with observed lesions after 5 days followed by *F. verticilloides* then *F. chlamydosporum*. After 20 days, *F. incarnatum* had the highest number of plants with observed lesions while *F. oxysporum* had the least. *Fusarium oxysporum* had the highest number of dead weeds followed by *F. equiseti* then *F. verticilloides* (Fig. 3). All the fungal isolates demonstrated significant infection rates (significance level less than 0.001 at $p = 0.05$) with *F. incarnatum* having the highest infection rate of 92%, followed by *Gibberella intricans* and *F. chlamydosporum* at 90% each. Consequently the fungal isolates lead to mortality of the weed ($P < 0.05$). *Fusarium oxysporum* strains had the highest mortality rates of 60% and 58% followed by *F.*

equiseti at 46%. *Gibberella intricans*, on the other hand, had the least mortality rate at 36% (Fig. 4). Levene's test for equality of variances revealed that the means of infection and mortality rates of the fungal isolates were not significantly different (Significance level was 0.056 at $p = 0.05$). Infected *S. hermonthica* plants which survived were emaciated and developed fewer flowers and consequently seeds than usual.

Discussion

Fusarium sp. was the most abundant fungal species isolated from *S. hermonthica*. All the *Fusarium* spp. were isolated from the leaves and flowers of *S. hermonthica* with *Colletotrichum gloeosporioides* being isolated from the stems and *Hypocrea stellata* and *Arthroderma otae* being isolated from the roots. Molecular characterization of the fungal isolates revealed that the amplified genomic DNA of the fungal isolates ranged between 100bp to 850bp, indicating their potential in genetic modification in producing more virulent strains for use as biocontrol agents. Phylogenetic analysis of the isolates revealed that the *Fusarium* spp. were related to each other, however, no relatedness was observed in terms of virulence.

In the screen house, all the fungal isolates tested showed high virulence efficacy on the weed revealing a wide variety of choice in developing a biological control against the weed. All the fungal isolates demonstrated infection rates greater than 75% indicating that the isolates tested could easily colonize the weed. However, it was only the two strains *Fusarium oxysporum* which had mortality rates greater than 50%, one having 60% and the other having 58% agreeing with the findings of Elzein *et al.*, (2008). This demonstrated the suitability of *F. oxysporum* as a biocontrol agent against the weed. The less than 50% mortality rate exhibited by most of the isolates can be attributed to the time of inoculation of the weed with the fungal isolates. The fungal isolates were inoculated when the *S. hermonthica* were 8 weeks old, probably a higher mortality rate could have been achieved with early inoculation, because perhaps young *S. hermonthica* plants are more susceptible. Infection of *S. hermonthica* with the fungal isolates reduced the growth vigour and consequently the biomass of the weed. The infected weeds demonstrated a great reduction in the number of flowers and seeds produced agreeing with the findings of Yonli *et al.*, (2010). Thus, the use of *Fusarium* spp. as biocontrol agents against *S. hermonthica* can limit the increase of the weed's soil seed bank.



KEY:

- | | | |
|------------------------------------|-------------------------------------------|------------------------------------|
| 2- <i>Hypocrea stellata</i> | 23- <i>Colletotrichum gloeosporioides</i> | 5- <i>Gibberella monoliformis</i> |
| 6- <i>Fusarium equiseti</i> | 12- <i>Fusarium incarnatum</i> | 19- <i>Fusarium oxysporum</i> |
| 30- <i>Gibberella intricans</i> | 33- <i>Arthroderma otae</i> | 37- <i>Fusarium oxysporum</i> |
| 38- <i>Fusarium oxysporum</i> | 39- <i>Fusarium oxysporum</i> | 43- <i>Fusarium chlamydosporum</i> |
| 44- <i>Fusarium chlamydosporum</i> | 46- <i>Fusarium verticilloides</i> | |

Figure 1 Micrograph showing amplified DNA bands.

Table 2 Table of symptomatology as portrayed by *Striga hermonthica* infected with the fungal isolates.

Isolate name	Symptoms	Illustrations	Isolate name	Symptoms	Illustrations
<i>Gibberella moniliformis</i>	Caused lesions on both the stem and leaves of the weed. The leaves turned dark maroon, followed by blackening of the stems and eventual drying and death of the weed.		<i>Fusarium incarnatum</i>	Caused lesions on the leaves of the weed. The leaves became twisted and developed an ashy burned appearance at the tip. The weed then dried and died.	
<i>Gibberella intricans</i>	Caused the leaves to turn pale, then maroon and then twisted. The stems darkened from the ground up the plant this was followed by drying and eventual death of the weed.		<i>Fusarium equiseti</i>	Caused lesions on the leaves. The leaves became pale, rough textured and developed spots. Some leaves also curled and became twisted. The weed then dried and died.	
<i>Fusarium oxysporum</i>	Caused lesions on the leaves and the stem. The leaves turned pale then maroon with curling and twisting. Some leaves developed spots which perforated them. The stem darkened from the ground up the plant, followed by drying then death of the weed.		<i>Fusarium chlamydosporum</i>	Caused lesions on the leaves and on the stem. The leaves turned brownish, developed whitish spots, curled, became twisted and then dried. The weed dried from the tip down the stem and eventually died.	
<i>Fusarium verticilloides</i>	Caused lesions on the leaves. The leaves turned maroon, developed whitish spots and an ashy burned appearance at the tip and on the edges of the leaves. The plant then dried and died.				

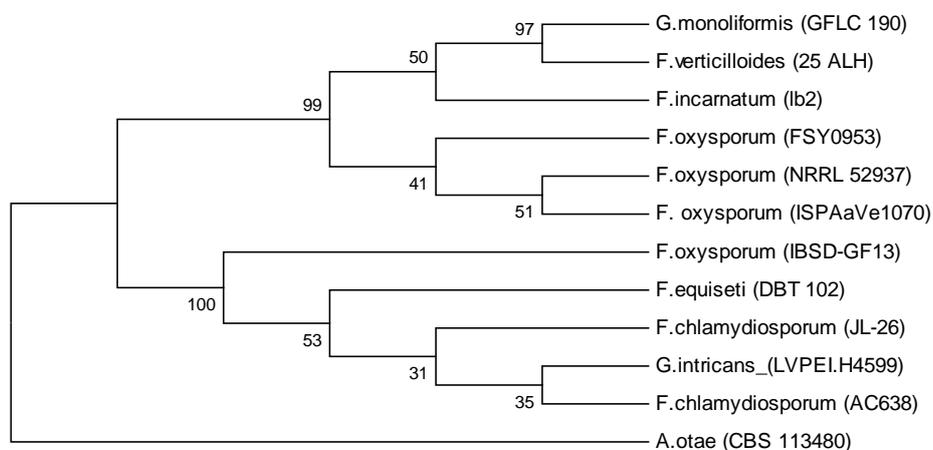


Figure 2 Phylogenetic analysis of the isolated fungal species.

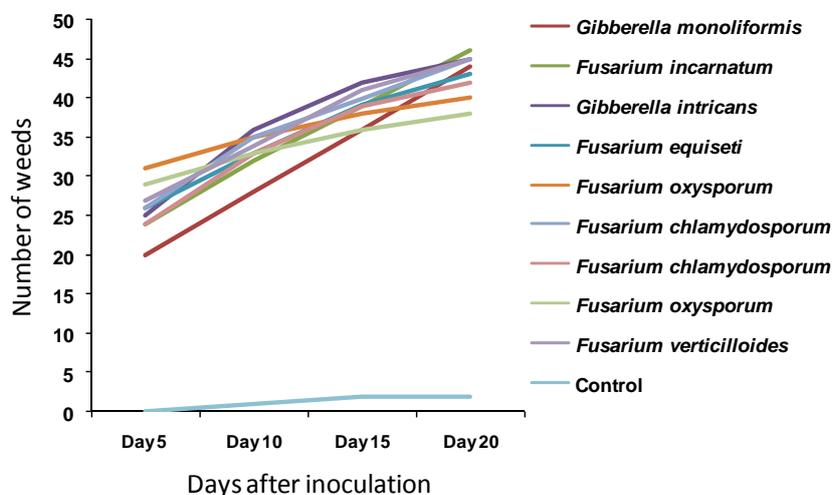


Figure 3 Number of weeds, having necrosis and lesions in *Striga hermonthica* after inoculation with the fungal isolates.

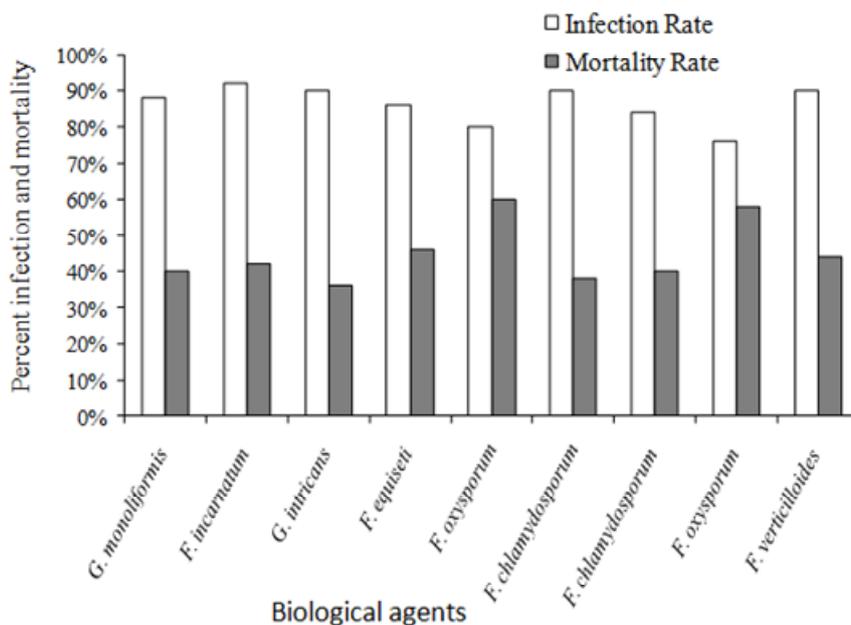


Figure 4 Infection and mortality rates of *Striga hermonthica* 22 days after inoculation with fungal isolates.

Conclusion

Though all the fungal isolates tested against the weed could cause infection and consequently kill the weed, *Fusarium oxysporum* was the most aggressive recording the highest mortality rate of

more than 50% within the shortest time. The others took longer which is not advantageous in the development of a biological control agent. Further research should be done on the fungal isolates tested as mycoherbicides so as to elucidate their maximum potential, safety and to find out how best

they can be used and exploited as biocontrol agents against the weed on a commercial scale.

Acknowledgement

The author expresses deep and sincere gratitude to the International Maize and Wheat Improvement Centre (CIMMYT) and International Centre for Tropical Agriculture (CIAT) for jointly funding this research work. A mentioned also goes to the University of Nairobi for providing the technical support in piecing this work together.

References

- Atera, E. A., Ishii, T., Onyango, J. C., Itoh, K. and Azuma, T. 2013. *Striga* Infestation in Kenya: Status, Distribution and Management Options. *Sustainable Agriculture Research*, 2 (2): 99.
- Booth, C. 1971. *The Genus Fusarium*. Commonwealth Mycological Institute, Kew, Surrey, England.
- Dugan, F. M. 2006. *The Identification of Fungi: an illustrated introduction with keys, glossary, and guide to literature*. American Phytopathological Society.
- Elzein, A., Brändle, F., Cadisch, G., Kroschel, J., Marley, P. and Thines, M. 2008. *Fusarium oxysporum* strains as potential *Striga* mycoherbicides: Molecular characterization and evidence for a new forma specialis. *The Open Mycology Journal*, 2: 89-93.
- Esilaba, A. O. 2006. Options for *Striga* management in Kenya: Kenya Agricultural Research Institute (KARI). Technical Note No. 19.
- Felsenstein, J. 1985. Phylogenies and the Comparative Method. *American Naturalist*, 1-15.
- Gacheru, E., Odhiambo, G. D., and Bashir, J. 2002. *Striga* biology and participatory control approaches: A Facilitators/Extension guide: KARI-CIMMYT-ICRAF-ICIPE together with FAO 2nd edition.
- Gerlach, W. and Nirenberg, H. 1982. *The Genus Fusarium—A Pictorial Atlas*. Mitteilungen Aus Der Biologischen Bundesanstalt Fur Land- Und Forstwirtschaft Berlin-Dahlem, 209.
- Jamil, M., Kanampiu, F. K., Karaya, H., Charnikhova, T. and Bouwmeester, H. J. 2012. *Striga hermonthica* parasitism in maize in response to N and P Fertilisers. *Field Crops Research*, 134: 1-10.
- Kwaśna, H. and Bateman, G. L. 2007. *Heteroconium* sp. nov. from roots of *Triticum aestivum* in the United Kingdom. *Mycologia*, 99 (5): 777-785.
- Leslie, J. F. and Summerell, B. A. 2006. *The Fusarium Laboratory Manual*. Blackwell Publishing.
- Ndambi, B., Cadisch, G., Elzein, A. and Heller, A. 2011. Colonization and Control of *Striga hermonthica* by *Fusarium oxysporum* f. sp. *strigae*, a Mycoherbicide Component: An anatomical study. *Biological Control*, 58 (2): 149-159.
- Olwenya G. 2012. *Striga* weed destroys sh1.8 billion of maize in farms. *Standard Digital*, Retrieved on 9th June 2012 from: Retrieved from www.standardmedia.co.ke
- Rahjoo, V., Zad, J., Javan-Nikkhah, M., Mirzadi Gohari, A., Okhovvat, S. M., Bihamta, M. R. and Klemsdal, S. S. 2008. Morphological and molecular identification of *Fusarium* isolated from maize ears in Iran. *Journal of Plant Pathology*, 90 (3): 463-468.
- Tamura, K., Nei, M. and Kumar, S. 2004. Prospects for inferring very large phylogenies by using the neighbor-joining method. *Proceedings of the National Academy of Sciences of the United States of America*, 101 (30): 11030-11035.
- Tamura, K., Peterson, D., Peterson, N., Stecher, G., Nei, M. and Kumar, S. 2011. MEGA5: Molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. *Molecular Biology and Evolution*, 28 (10): 2731-2739.
- Yonli, D., Traoré, H., Sérémé, P., Hess, D. E. and Sankara, P. 2010. Pathogenicity of *Fusarium* isolates to *Striga hermonthica* in Burkina Faso. *Pakistan Journal of Biological Sciences*, 13 (5): 201.

ارزیابی جدایه‌های قارچی به‌عنوان عوامل بیوکنترل علف جادو

ویکتور کاگوت^{۱*}، شیلا اُکوت^۱، فرد کانامپیو^۲، پیتر اُکوت و ادنا ماگتو^۲

۱- دانشگاه علوم زیستی، دانشگاه نایروبی، کنیا.

۲- مرکز بین‌المللی اصلاح گندم (سیمیت CIMMYT) صندوق پستی: ۰۰۶۲-۱۰۴۱، نایروبی، کنیا.

۳- مرکز بین‌المللی کشاورزی مناطق گرمسیری (CIAT) صندوق پستی: ۰۰۱۰۰-۳۰۶۷، نایروبی، کنیا.

* پست الکترونیکی نویسنده مسئول مکاتبه: kagovic14@yahoo.com

دریافت: ۱۵ مرداد ۱۳۹۲؛ پذیرش: ۷ بهمن ۱۳۹۲

چکیده: علف جادو (*Striga hermonthica*) علف هرز انگلی می‌باشد که عمدتاً در تولید محصول ذرت و سورگوم در غرب کنیا مزاحمت ایجاد می‌کند. این علف هرز بیشتر مزارع با اندازه کوچک را مورد حمله قرار می‌دهد و بستگی به شدت ممکن است بازه‌ای از ۱۰ درصد تا کل محصول را از بین ببرد و عدم اطمینان از خودکفایی مواد غذایی در منطقه را تشدید می‌کند. هدف این بررسی ارزیابی جدایه‌های قارچی به‌عنوان عوامل بیوکنترل احتمالی علیه این علف هرز بود. قارچ‌ها از علف جادوی آلوده جداسازی شد و کارایی بیماری‌زایی آنها علیه علف جادو در گلخانه بررسی شد. همه‌ی جدایه‌های قارچی آزمایش شده سبب آلودگی و در نتیجه مرگ علف هرز شد. *Fusarium incarnatum* بیشترین آلودگی در حدود ۹۲٪ داشت و به‌دنبال آن *Gibberella intricans* و *F. chlamdosporum* هر کدام ۹۰٪ آلودگی داشتند. *F. oxysporium* سبب بیشترین مرگ‌ومیر با ۶۰٪ و *G. intricans* با ۳۶٪ مرگ‌ومیر کمترین اثر را داشت. *F. oxysporium* متهاجم‌ترین و پرتوان‌ترین جدایه قارچی علیه علف هرز بود بنابراین نامزد مناسبی برای بهره‌برداری به‌عنوان یک علف‌کش قارچی علیه علف هرز جادوگر می‌باشد.

واژگان کلیدی: بیوکنترل، علف‌کش قارچی، *Fusarium*، *Striga hermonthica*