Combined application of *Pseudomonas fluorescens* and *Purpureocillium lilacinum* liquid formulations to manage *Globodera* spp on potato

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Abstract: Potato cyst nematodes (PCN), *Globodera rostochiensis* and *Globodera pallida* are major limiting factors to potato cultivation globally. Effective use of nematode antagonistic bio-agents is a potentially important component of the eco-friendly agro-farming. *Pseudomonas fluorescens* and *Purpureocillium lilacinum* are known for their nematode antagonistic potential and plant growth promotion ability. The effect of seed treatment with liquid suspension of *P. fluorescens* at 1 l/ton seeds and soil drenching with suspension of *P. lilacinum* at 5 l/ha, singly and jointly, was studied to minimize the damage caused by PCN in potato plants under field conditions in two regions in India. Both applications showed significantly greater PCN suppression and better plant growth promotion in comparison to solo application. The both application showed the highest reduction of cyst population (75.7%) in soil, female population (79.9%) in root and egg numbers per soil of each location (84%). The potato plants from *P. fluorescens*-seed treatment and *P. lilacinum*-soil drenching both applied plots were 33.5% taller with 45.6% more number of tubers than untreated plants. The tuber yield was also significantly higher (35.9%) in both application than untreated control. There was no significant difference on the root colonization of *P. fluorescens* and *P. lilacinum* in solo and both treatments.

Keywords: Potato cyst nematode, biological control, liquid formulations, *Pseudomonas fluorescens, Purpureocillium lilacinum*

Introduction

Potato *Solanum tuberosum* L. is an important tuber crop that is grown globally to meet food requirement of people in many countries. It is considered fourth important food crop after rice, wheat and maize. It is also used as animal feed and to make commercial starch products. In India, potato is cultivated in 28 states with total area of 20.24 million ha, producing 46.4 million tonnes annually (Welfare, 2016). Among the various pest and diseases associated with this crop, potato cyst nematodes, *Globodera rostochiensis* (Woll) and *Globodera pallida* (Stone) remain a daunting challenge for potato production. PCN are sedentary root endo-parasites. The second-stage juveniles (J2) penetrate through growing tips of roots and form feeding sites or syncytia in vascular tissues which lead to stunted growth, early senescence, proliferation of lateral roots and partial or complete arrest of tuber formation (Devraj et al., 2004). In addition, root damage caused by PCN provides an avenue for
entry of fungal pathogen such as *Rhizoctonia solani* resulting in crop loss due to the synergistic disease complex (Back et al., 2006). In Europe and North America, the yield loss due to PCN has been reported as 9-100% (Pineda et al., 1993). In India, up to 80% yield loss due to PCN was reported from Nilgiris and Kodaikanal hills, Tamil Nadu region (Devrajan et al., 2011), Karnataka, Kerala and Himachal Pradesh (Krishna Prasad and Singh, 1986; Ramana and Mohandas, 1988; Sudershan et al., 2010).

The chemical nematicide carbofuran 3G is frequently used to control nematodes, but its repeated use is required to maintain cyst populations below the damage threshold levels (Seenivasan, 2017). The drawbacks of chemical nematicides such as the potential residue, groundwater contamination, enhanced biodegradation and toxicity to applicators also necessitated to search for alternative method of control. Biological control with fungal or bacterial organism that effectively antagonise the nematodes is an ecologically sound approach that has tremendous prospective to control nematode population build up and thereby reduce the crop damage (Seenivasan and Sundarababu, 2007). The root colonizing plant growth promoting rhizobacteria like *P. fluorescens* have shown better result for the management of various plant parasitic nematodes such as *Hirschmaniella oryza* on rice (Seenivasan and Lakshmanan, 2002), *Globodera rostochiensis* (Devrajan et al., 2004), *Meloidogyne graminicola* on rice (Seenivasan, 2011), *Radopholus similis* on banana (Seenivasan et al., 2013), *Meloidogyne javanica* on tomato (Siddiqui and Shaukat, 2004), *Meloidogyne incognita* on medicinal coleus (Seenivasan and Devrajan, 2008) and jasmine (Seenivasan and Poornima, 2010). The facultative egg parasitic fungus, *Purpureocillium lilacinum* (= *Paecilomyces lilacinus*) has been reported to be effective against *Meloidogyne* spp and many other plant parasitic nematodes in various crops (Rao, 2008; Rao et al., 2012; Crow, 2013; Mohd et al., 2009). Earlier reports by Devrajan et al. (2004) and Seenivasan et al. (2007) demonstrated the biocontrol potential of *P. fluorescens* and *P. lilacinum* against PCN in potato. In the field situation, the performance of bio-control agents is not efficient enough to provide sufficient nematode control as like that of chemical nematicides. Recently, the concept of combined use of different biocontrol agents was attempted on crops of tomato, pumpkin, sugar beet and chickpea and demonstrated successfully against various plant parasitic nematodes (Seenivasan et al., 2012). However, there are no reports on the combined use of these biocontrol agents for the management of PCN on potato. Hence this study aimed to find out the effect of combined use of liquid suspensions of *P. fluorescens* and *P. lilacinum* to manage PCN in the field conditions.

**Materials and Methods**

**Bio-formulations**

The liquid formulation of *P. fluorescens* strain Pf1 containing $5 \times 10^9$ colony forming units (cfu)/ml was obtained from the Department of Plant Pathology, Tamil Nadu Agricultural University, Coimbatore, India. The liquid formulation of *P. lilacinum* strain Ooty1 containing $4 \times 10^9$ cfu/ml was obtained from Horticultural Research Station, Tamil Nadu Agricultural University, Ooty, India.

**Field studies**

Two field trials were conducted in the farmer fields naturally infested with PCN populations at Shenbaganur village (Location I) and Bambharapuram village (Location II), Kodaikanal, Tamil Nadu, India. The mixed populations of *G. rostochiensis* and *G. pallida* existed in both fields. Both trials were laid at similar time period from February 2016 to April 2016. Seed tubers of the potato cv. Kufri Jothi were used for field trials. Both experiments consisted of the following 5 treatments: (1) Seed treatment (ST) with liquid suspension of *P. fluorescens* ($5 \times 10^9$ colony forming units (cfu)/ml) at 1 l/ha seed; (2) Soil drenching (SD) with liquid suspension of *P. lilacinum* ($5 \times 10^9$}
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(Furadan 3G, Cobb, 1918)

first by Cobb’s sieving and decanting method

through 625 mesh (20µm). Then the residue collected was processed by centrifugal floatation technique to separate eggs (Barker and Niblack, 1990). Eggs and juveniles were counted by viewing under a stereo zoom microscope (Kozo Zoom 645) at a magnification of 40x. Since, the juvenile population was very low in each plot and location, juvenile count was added with egg count. Five plants from each plot just before harvest were collected and female population per 2.5 cm root length were recorded under a stereo zoom microscope.

Re-isolation of introduced bio-agents

Root colonization of the introduced P. fluorescens and P. lilacinum was assessed from 1 g root samples from each plot following the serial dilution plate technique as described by Seenivasan (2011). Kings B media and potato dextrose agar media were used for P. fluorescens and P. lilacinum, respectively. Percentage of parasitized cysts by P. lilacinum was also assessed. Ten cysts were hand-picked, rinsed with sterile distilled water two times and plated on potato dextrose agar media in 90 mm Petri plates. The plates were incubated at 28 ± 3 °C for 15 days and fungi parasitization were observed under stereo zoom microscope. The percentage of parasitized eggs was calculated using the formula: (no of cysts infected with fungus/total number of cysts) × 100.

Statistical procedure

The data collected were analyzed for one-way analysis of variance using SPSS 16.0 for Windows software (SPSS Inc., Chicago, IL, USA). The treatment means were compared by Duncan’s multiple range test (DMRT) (Panse and Sukhatme, 1954).

Results

Results showed that PCN cyst density and egg numbers in soil, as well as adult female

General observations

The stem length was measured at 90 days after sowing (DAS) from randomly selected five plants per plot. Plants were harvested on 120 DAS and root tuber yield recorded from all plots. Yield was expressed in tonne (t) per ha. Number of tubers/plant was recorded from five randomly selected plants. The population density of cyst in soil from each plot was determined before treatment and at harvest. Each sample comprising of 10 random cores collected at a depth of 15-20cm and pooled together into a composite sample. A subsample of 200 cm³ from each composite sample was processed by Fenwick’s floatation method (Fenwick, 1940). The population of PCN cysts was counted under a stereoscope microscope. A subsample of 100 g soil was taken from each composite sample after thorough mixing and used for egg estimation. The cysts were extracted from the samples first by Cobb’s sieving and decanting method (Cobb, 1918). The residue containing cysts collected from the 60 mesh (250µ) were crushed by mechanical cyst crusher to release eggs and the macerated suspension was poured through 625 mesh (20µm). Then the residue collected was processed by centrifugal floatation technique to separate eggs (Barker and Niblack, 1990). Eggs and juveniles were counted by viewing under a stereo zoom microscope (Kozo Zoom 645) at a magnification of 40x. Since, the juvenile population was very low in each plot and location, juvenile count was added with egg count. Five plants from each plot just before harvest were collected and female population per 2.5 cm root length were recorded under a stereo zoom microscope.

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Results

Results showed that PCN cyst density and egg numbers in soil, as well as adult female
population in roots were significantly reduced in \textit{P. fluorescens} and \textit{P. lilacinum} treated plots in both fields (Tables 1 and 2). Combined application of seed treatment with \textit{P. fluorescens} and soil drench with \textit{P. lilacinum} was the most effective in controlling PCN. This treatment reduced cyst population in soil by 75.5\% in Location I and 75.8\% in Location II, being significantly superior to their individual applications and also standard chemical carbofuran treatment. The \textit{P. fluorescens} seed treatment and \textit{P. lilacinum} soil drench individually resulted in the smallest reduction of cyst populations (48.3\% and 51.4\%, respectively) over the control. The number of females/2.5 cm root was also significantly less in \textit{P. fluorescens} ST + \textit{P. lilacinum} SD as compared to the individual treatments, carbofuran and untreated plants. The combined treatment reduced the root penetration of PCN by 79.8\% compared to when \textit{P. fluorescens} ST and \textit{P. lilacinum} SD were applied individually.

The number of PCN eggs in soil was significantly higher in control plots (Tables 1 and 2). The egg population in soil from all other treatment plots was found to be less than control. The plots treated with combination of \textit{P. fluorescens} ST + \textit{P. lilacinum} SD had significantly least egg population that was 84\% less than control plots. However, egg population reduction was only 57.3\% in \textit{P. fluorescens} ST and 55.1\% in \textit{P. lilacinum} SD treatments. Plants from untreated plots were smaller and had fewer number of tubers compared to treated plots in both trials (Tables 1 and 2). Seed treatment with \textit{P. fluorescens} accompanied with soil drench with \textit{P. lilacinum} had significantly higher effect on plant growth improvement than all other treatments. The plants in this treatment were 33.5\% taller with 45.6\% more number of tubers than the untreated plants. The growth improvement was lesser in \textit{P. lilacinum} SD treated plots than \textit{P. fluorescens} ST alone and combination of \textit{P. fluorescens} ST and \textit{P. lilacinum} SD plots. The improved plant growth in carbofuran, \textit{P. fluorescens} ST, \textit{P. lilacinum} SD and combination of \textit{P. fluorescens} ST and \textit{P. lilacinum} SD plots resulted in significant increase in potato yield. The maximum tuber yield increase (35.9\%) was noticed in the plots treated with combination of \textit{P. fluorescens} ST and \textit{P. lilacinum} SD followed by carbofuran (21.1\%), \textit{P. fluorescens} ST alone (16.1\%) and \textit{P. lilacinum} SD alone (15.5\%) treated plots.

The introduced \textit{P. fluorescens} and \textit{P. lilacinum} singly or in combination survived in potato roots up to harvest. The colonization of roots by \textit{P. fluorescens} was statistically uniform in plots applied individually or in combination with \textit{P. lilacinum}. Similarly, root colonization and cyst parasitization by \textit{P. lilacinum} were not significantly different compared with combination of \textit{P. fluorescens} (Tables 1 and 2).

\begin{table}[h]
\centering
\begin{tabular}{|c|c|c|c|c|c|c|c|c|c|}
\hline
Treatments & Cyst population & Eggs/100 g soil & Number of females & Plant height & Number of tubers/ plant & Tuber yield & Root colonization & Cyst parasitization by \\
& & & & (2.5 cm root) & & & \textit{P. fluorescens} & \textit{P. lilacinum} \\
& & & & & & & (CFU x 10^8 g\(^{-1}\) root) & (\%) \\
Before & 120 & Before & 120 & & & & & \\
& treatment & DAT & treatment & DAT & & & & \\
\hline
\text{T\textsubscript{1}} & 131 a & 238 b & 1902 a & 5863 b & 13.1 b & 55.3 b & 14.2 b & 13.4 b & 2065 a & - & - \\
\text{T\textsubscript{2}} & 128 a & 224 b & 1856 a & 5623 b & 12.4 b & 54.1 b & 14.9 b & 13.9 b & - & 197 a & 58 a \\
\text{T\textsubscript{1} + T\textsubscript{2}} & 127 a & 112 c & 1894 a & 1989 c & 4.3 c & 58.3 a & 18.7 a & 18.1 a & 2012 a & 192 a & 53 a \\
\text{T\textsubscript{3}} & 125 a & 208 b & 1924 a & 5148 b & 11.6 b & 54.9 b & 15.2 b & 14.7 b & - & - & - \\
\text{UC} & 130 a & 459 a & 1863 a & 12617 a & 21.7 a & 38.7 c & 10.2 c & 11.4 c & - & - & - \\
\hline
\end{tabular}
\caption{Effect of liquid bio-formulation treatments on potato cyst nematode infection, growth and yield of potato cv. Kufri Jothi. (Location I).}
\end{table}

\text{FT: Seed treatment with liquid \textit{P. fluorescens} (1 L/ton seed), T2: Soil drenching with liquid \textit{P. lilacinum} (5 L/ha), T4: Carbofuran 3G (1Kg a.i/ha), UC: Untreated control. DAT: Days after treatment. Means followed by the same letter in columns are not significantly different at P ≤ 0.05 according to Duncan’s multiple range test; CFU: colony forming units.}
Table 2. Effect of liquid bio-formulation treatments on potato cyst nematode infection, growth and yield of potato cv. Kufri Jothi. (Location II).

<table>
<thead>
<tr>
<th>Treatments</th>
<th>Cyst population (100 cm² / soil)</th>
<th>Egg/ 100 g soil</th>
<th>Number of females/2.5 cm root</th>
<th>Plant height (cm)</th>
<th>Number of tubers/plant</th>
<th>Tuber yield (t/ha)</th>
<th>Root colonization (CFU x 10⁸ g⁻¹ root)</th>
<th>Cyst parasitisation by P. lilacinum (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>T1</td>
<td>154 a</td>
<td>273 b</td>
<td>6318 b</td>
<td>15.2 b</td>
<td>56.0 b</td>
<td>13.7 b</td>
<td>12.8 b</td>
<td>3153 a</td>
</tr>
<tr>
<td>T2</td>
<td>162 a</td>
<td>257 b</td>
<td>6072 b</td>
<td>14.5 b</td>
<td>55.1 b</td>
<td>14.3 b</td>
<td>13.3 b</td>
<td>258 a</td>
</tr>
<tr>
<td>T3</td>
<td>152 a</td>
<td>197 a</td>
<td>2148 c</td>
<td>5.1 c</td>
<td>58.9 a</td>
<td>18.1 a</td>
<td>17.5 a</td>
<td>31717 a</td>
</tr>
<tr>
<td>T4</td>
<td>156 a</td>
<td>206 a</td>
<td>5556 b</td>
<td>13.7 b</td>
<td>55.9 b</td>
<td>15.7 b</td>
<td>14.2 b</td>
<td>60 a</td>
</tr>
</tbody>
</table>

T1: Seed treatment with liquid *P. fluorescens* (1 L/ton seed); T2: Soil drenching with liquid *P. lilacinum* (5 L/ha); T4: Carbofuran 3G (1 kg a.i/ha), UC: Untreated control, DAT: Days after treatment.

Means followed by the same letter in columns are not significantly different at P < 0.05 according to Duncan’s multiple range test; CFU: colony forming units.

Discussion

The individual lethal effect of *P. fluorescens* and *P. lilacinum* have been demonstrated against Meloidogyne spp. (Seenivasan and Devrajan, 2008; Seenivasan and Poornima, 2010). In this study, the efficacy of PCN control was significantly higher by the combined application of *P. fluorescens* seed treatment and *P. lilacinum* soil drench when they were used alone. Increasing population of biocontrol agents with antagonistic activities against nematodes in the rhizosphere have been reported to improve soil suppressiveness (Shaukat and Siddiqui, 2001). Similarly, control of *M. incognita* on tomato and bell pepper has been improved by combining *P. fluorescens* and *P. lilacinum* (Rao et al., 2012; Hashem and Abo-Elyour, 2011). Seenivasan (2010) also showed that integration of *P. fluorescens* and *P. lilacinum* effectively reduced *M. incognita* and *Macrophomina phaseolina* disease complex on medicinal coleus. In addition, the synergistic effect between *P. fluorescens* and *P. lilacinum* was reported to provide more efficient and consistent nematode control in gladiolus fields (Sowmya and Rao, 2013). This study confirms that combined application of *P. fluorescens* and *P. lilacinum* is beneficial in the management of PCN.

The mechanism of PCN protection by *P. fluorescens* and *P. lilacinum* is attributed to the following direct or indirect effects. The root colonization by *P. fluorescens* has been reported to alter the root exudates that affect the nematode egg hatching, attraction towards root and root penetration potential (Seenivasan and Lakshmanan, 2002). In their study culture filtrates of *P. fluorescens* strain Pf1 was reported to have nematotoxic principle (Seenivasan and Lakshmanan, 2001). The *P. fluorescens* strain Pf1 also has the ability to induce systemic resistance against nematodes in plants by producing peroxidases, polyphenol oxidases, phenylalanine ammonia lyase and 1-aminoacyclopropane-1-carboxylic acid (ACC) deaminase enzymes (Seenivasan, 2011; Saravanakumar and Samiyappan, 2006). It is a well-established fact that *P. lilacinum* colonizes roots of diverse plants, parasitizes cysts, eggs, juveniles and adult females of *Globodera* spp. by direct hyphal penetration (Jatala, 1986). Apart from direct parasitism, the development of *P. lilacinum* early in the soil might prevent the initial infection resulting in lower level of root penetration. All strains of *P. lilacinum* are reported to produce acetic acid and some metabolites like paeclotoxins and leucinostatins which are found to have detrimental effect on nematode juveniles (Singh et al., 2013). These metabolites may also probably be involved in reduction of PCN juveniles. Furthermore, being a parasite of mature females it would affect their egg production (Jatala, 1986) and *P. lilacinum* is capable of arresting syncitia formation induced by nematodes in plants (Cabanillas et al., 1988).
The results proved that the *P. fluorescens* and *P. lilacinum* enhanced the growth of plants in addition to PCN reduction. Apart from their effect on nematode, *P. fluorescens* and *P. lilacinum* are recognized to possess plant growth promoting effect in many crop plants like pigeon pea and tomato (Siddiqui et al., 1998; Khan and Akram, 2000). The *P. fluorescens* strain Pfi is reported to induce plant growth by producing plant growth regulators like indole acetic acid, gibberellins and cytokinins (Seenivasan, 2011). *P. lilacinum* on the other hand improves plant growth by increasing the available phosphorus in the soil (Lima-Rivera et al., 2016). The results of this study showed that root colonization of *P. fluorescens* did not affect *P. lilacinum* and vice-versa. Similar result was reported in bell pepper in which root colonization by *P. fluorescens* did not affect *P. lilacinum* (Rao et al., 2012).

It is concluded that seed treatment with *P. fluorescens* followed by *P. lilacinum* soil drenching can be recommended for the practical management of *G. rostochiensis* and *G. pallida* infection in potato fields rather than application of a single bio-agent.

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**Disclosure statement**
No potential conflict of interest was reported by the author.

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کاربرد ترکیبی از فرمولاسیون مایع باکتری Pseudomonas fluorescens و قارچ Purpureocillium lilacinum در مدیریت نماتدهای سیب‌زمینی Globodera spp

نامه‌اولین از سیب‌زمینی‌های سیب‌زمینی در پاتولوژی و بیولوژی فرآیند نماتود‌پروری در ناحیه‌های الزام‌گذاری‌ها در زمین‌های سیب‌زمینی در هندوستان مطالعه شد. در این پژوهش برای به دست آوردن رالد نماتدهای نامیده شده در نماتدهای سیب‌زمینی، نمونه‌بندی و تولید رازه‌گیر کیلولگری P. fluorescens به میزان 5 لیتر در هر کیلوگرم خاک و خیساندن هاک با سوسپنژ P. lilacinum به میزان 5 لیتر در هر کیلوگرم خاک و افزایش پرورش بیش از افزایش در نوع هر دو به‌طور یکپارچه در شرایط مزروع، در هنگام نماتود‌پروری در بالعصر جمعیتی P. fluorescens به میزان 5/33 درصد و P. lilacinum به میزان 8/45 درصد در تعداد بیش از گیاهان مقدار نمایندگی نمایندگی آرانگی و تعداد افزایشی P. fluorescens و خیساندن جمعیتی P. lilacinum در اکثریت آرا نمایندگی در نمایی‌های هر دو باعث بهبود رشد گیاهان شد. نتایج نشان داد که نماتدهای نامیده شده در تعداد بیش از اکثریت آرا نمایندگی در نمایی‌های هر دو باعث بهبود رشد گیاهان شد. نتایج نشان داد که نماتدهای نامیده شده در تعداد بیش از اکثریت آرا نمایندگی در نمایی‌های هر دو باعث بهبود رشد گیاهان شد. نتایج نشان داد که نماتدهای نامیده شده در تعداد بیش از اکثریت آرا نمایندگی در نمایی‌های هر دو باعث بهبود رشد گیاهان شد. نتایج نشان داد که نماتدهای نامیده شده در تعداد بیش از اکثریت آرا نمایندگی در نمایی‌های هر دو باعث بهبود رشد گیاهان شد. نتایج نشان داد که نماتدهای نامیده شده در تعداد بیش از اکثریت آرا نمایندگی در نمایی‌های هر دو باعث بهبود رشد گیاهان شد. نتایج نشان داد که نماتدهای نامیده شده در تعداد بیش از اکثریت آرا نمایندگی در نمایی‌های هر دو باعث بهبود رشد گیاهان شد. نتایج نشان داد که نماتدهای نامیده شده در تعداد بیش از اکثریت آرا نمایندگی در نمایی‌های هر دو باعث بهبود رشد گیاهان شد. نتایج نشان داد که نماتدهای نامیده شده در تعداد بیش از اکثریت آرا نمایندگی در نمایی‌های هر دو باعث بهبود رشد گیاهان شد. نتایج نشان داد که نماتدهای نامیده شده در تعداد بیش از اکثریت آرا نمایندگی در نمایی‌های هر دو باعث بهبود رشد گیاهان شد. نتایج نشان داد که نماتدهای نامیده شده در تعداد بیش از اکثریت آرا نمایندگی در نمایی‌های هر دو باعث بهبود رشد گیاهان شد. نتایج نشان داد که نماتدهای نامیده شده در تعداد بیش از اکثریت آرا نمایندگی در نمایی‌های هر دو باعث بهبود رشد گیاهان شد. نتایج نشان داد که نماتدهای نامیده شده در تعداد بیش از اکثریت آرا نمایندگی در نمایی‌های هر دو باعث بهبود رشد گیاهان شد. نتایج نشان داد که نماتدهای نامیده شده در تعداد بیش از اکثریت آرا نمایندگی در نمایی‌های هر دو باعث بهبود رشد گیاهان شد. نتایج نشان داد که نماتدهای نامیده شده در تعداد بیش از اکثریت آرا نمایندگی در نمایی‌های هر دو باعث بهبود رشد گیاهان شد. نتایج نشان داد که نماتدهای نامیده شده در تعداد بیش از اکثریت آرا نمایندگی در نمایی‌های هر دو باعث بهبود رشد گیاهان شد. نتایج نشان داد که نماتدهای نامیده شده در تعداد بیش از اکثریت آرا نمایندگی در نمایی‌های هر دو باعث بهبود رشد گیاهان شد. نتایج نشان داد که نماتدهای نامیده شده در تعداد بیش از اکثریت آرا نمایندگی در نمایی‌های هر دو باعث بهبود رشد گیاهان شد. نتایج نشان داد که نماتدهای نامیده شده در تعداد بیش از اکثریت آرا نمایندگی در نمایی‌های Henshaw نشان داد که نماتدهای نامیده شده در تعداد بیش از اکثریت آرا نمایندگی در نمایی‌های Henshaw نشان داد که نماتدهای نامیده شده در تعداد بیش از اکثریت آرا نمایندگی در نمایی‌های Henshaw نشان داد که نماتدهای نامیده شده در تعداد بیش از اکثریت آرا نمایندگی در نمایی‌های Henshaw نشان داد که Nematology, 2017, Vol. 6 (4)